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(54) Title: SCARECROW GENE, PROMOTER AND USES THEREOF (57) Abstract The structure and function of a regulatory gene, <i>SCARECROW</i> (<i>SCR</i>), is described. The <i>SCR</i> gene is expressed specifically in root progenitor tissues of embryos, and in roots and stems of seedlings and plants. <i>SCR</i> expression controls cell division of certain cell types in roots and affects the organization of root tissues, and affects gravitropism of aerial structures. The invention relates to the <i>SCARECROW</i> (<i>SCR</i>) gene, <i>SCR</i> gene products, (including but not limited to transcriptional products such as mRNAs, antisense, and ribozyme molecules, and translational products such as the <i>SCR</i> protein, polypeptides, peptides and fusion proteins related thereto), antibodies to the <i>SCR</i> product, <i>SCR</i> promoters and regulatory regions and the use of the foregoing to improve agronomically valuable plants.		

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SCARECROW GENE, PROMOTER AND USES THEREOF

This application is a continuation-in-part of co-pending Application No. 08/638,617, filed April 26, 1996, the disclosure of which is incorporated by reference in its entirety.

This invention was made with government support under grant number: GM43778 awarded by the National Institute of Health. The government may have certain rights in the invention.

1. INTRODUCTION

The present invention generally relates to the SCARECROW (SCR) gene family and their promoters. The invention more particularly relates to ectopic expression of members of the SCARECROW gene family in transgenic plants to artificially modify plant structures. The invention also relates to utilization of SCARECROW promoter for tissue and organ specific expression of heterologous gene products.

2. BACKGROUND OF THE INVENTION

Asymmetric cell divisions, in which a cell divides to give two daughters with different fates, play an important role in the development of all multicellular organisms. In plants, because there is no cell migration, the regulation of asymmetric cell divisions is of heightened importance in determining organ morphology. In contrast to animal embryogenesis, most plant organs are not formed during embryogenesis. Rather, cells that form the apical meristems are set aside at the shoot and root poles. These reservoirs of stem cells are considered to be the source of all post-embryonic organ development in plants. A fundamental question in developmental biology is how meristems function to generate plant organs.

2.1. ROOT DEVELOPMENT

Root organization is established during embryogenesis. This organization is propagated during postembryonic development by the root meristem. Following 5 germination, the development of the postembryonic root is a continuous process, a series of initials or stem cells continuously divide to perpetuate the pattern established in the embryonic root (Steeves & Sussex, 1972, Patterns in Plant Development, Englewood Cliffs, NJ: Prentice-Hall, Inc.).

10 Due to the organization of the Arabidopsis root it is possible to follow the fate of cells from the meristem to maturity and identify the progenitors of each cell type (Dolan et al., 1993, Development 119:71-84). The Arabidopsis root is a relatively simple and well characterized organ.

15 The radial organization of the mature tissues in the Arabidopsis root has been likened to tree rings with the epidermis, cortex, endodermis and pericycle forming radially symmetric cell layers that surround the vascular cylinder (FIG. 1A). See also Dolan et al., 1993, Development

20 119:71-84. These mature tissues are derived from four sets of stem cells or initials: i) the columella root cap initial; ii) the pericycle/vascular initial; iii) the epidermal/lateral root cap initial; and iv) the cortex/endodermal initial (Dolan et al., 1993, Development

25 119:71-84). It has been shown that these initials undergo asymmetric divisions (Scheres et al., 1995, Development 121:53-62). The cortex/endodermal initial, for example, first divides anticlinally (in a transverse orientation) (FIG. 1B). This asymmetric division produces another initial

30 and a daughter cell. The daughter cell, in turn, expands and then divides periclinally (in the longitudinal orientation) (FIG. 1B). This second asymmetric division produces the progenitors of the endodermis and the cortex cell lineages (FIG. 1B).

35

2.2. GENES REGULATING ROOT STRUCTURE

Mutations that disrupt the asymmetric divisions of the cortex/endodermal initial have been identified and characterized (Benfey et al., 1993, Development 119:57-70; Scheres et al., 1995, Development 121:53-62). *short-root* (*shr*) and *scarecrow* (*scr*) mutants are missing a cell layer between the epidermis and the pericycle. In both types of mutants the cortex/endodermal initial divides anticlinally, but the subsequent periclinal division that increases the number of cell layers does not take place (Benfey et al., 1993, Development 119:57-70; Scheres et al., 1995, Development 121:53-62). The defect is first apparent in the embryo and it extends throughout the entire embryonic axis which includes the embryonic root and hypocotyl (Scheres et al., 1995, Development 121:53-62). This is also true for the other radial organization mutants characterized to date, suggesting that radial patterning that occurs during embryonic development may influence the post-embryonic pattern generated by the meristematic initials (Scheres et al., 1995, Development 121:53-62).

Characterization of the mutant cell layer in *shr* indicated that two endodermal-specific markers were absent (Benfey et al., 1993, Development 119:57-70). This provided evidence that the wild-type *SHR* gene may be involved in specification of endodermis identity.

2.3. GEOTROPISM

In plants, the capacity for gravitropism has been correlated with the presence of amyloplast sedimentation. See, e.g., Volkmann and Sievers, 1979, Encyclopedia Plant Physiol., N.S. vol 7, pp. 573-600; Sack, 1991, Intern. Rev. Cytol. 127:193-252; Björkmann, 1992, Adv. Space Res. 12:195-201; Poff et al., in The Physiology of Tropisms, Meyerowitz & Somerville (eds); Cold Spring Harbor Laboratory Press, Plainview, NY (1994) pp. 639-664; Barlow, 1995, Plant Cell Environ. 18:951-962. Amyloplast sedimentation only occurs in cells in specific locations at distinct developmental stages.

That is, when and where sedimentation occurs is precisely regulated (Sack, 1991, Intern. Rev. Cytol. 127:193-252). In roots, amyloplast sedimentation only occurs in the central (columella) cells of the rootcap; as these cells mature into 5 peripheral cap cells, the amyloplasts no longer sediment (Sack & Kiss, 1989, Amer. J. Bot. 76:454-464; Sievers & Braun, in The Root Cap: Structure and Function, Wassail et al. (eds.), New York: M. Dekker (1996) pp. 31-49). In stems of many plants, including Arabidopsis, amyloplast 10 sedimentation occurs in the starch sheath (endodermis) especially in elongating regions of the stem (von Guttenberg, Die Physiologischen Scheiden, Handbuch der Pflanzenanatomie; K. Linsbauer (ed.), Berlin: Gebruder Borntraeger, vol. 5 (1943) p. 217; Sack, 1987, Can. J. Bot. 65:1514-1519; Sack, 15 1991, Intern. Rev. Cytol. 127:193-252; Caspar & Pickard, 1989, Planta 177:185-197; Volkmann et al., 1993, J. Pl. Physiol. 142:710-6).

Gravitropic mutants have been studied for evidence that proves the role of amyloplast sedimentation in gravity 20 sensing. However, many gravitropic mutations affect downstream events such as auxin sensitivity or metabolism (Masson, 1995, BioEssays 17:119-127). Other mutations seem to affect gene products that process information from gravity sensing. For example, the lazy mutants of higher plants and 25 comparable mutants in mosses can clearly sense and respond to gravity, but the mutations reverse the normal polarity of the gravitropic response (Gaiser & Lomax, 1993, Plant Physiol. 102:339-344; Jenkins et al., 1986, Plant Cell Environ 9:637-644). Other mutations appear to affect gravitropism of 30 specific organs. For example, *sgr* mutants have defective shoot gravitropism (Fukaki et al., 1996, Plant Physiol. 110:933-943; Fukaki et al., 1996, Plant Physiol. 110:945-955; Fukaki et al., 1996, Plant Res. 109:129-137).

Citation or identification of any reference herein 35 shall not be construed as an admission that such reference is available as prior art to the present invention.

3. SUMMARY OF THE INVENTION

The structure and function of a regulatory gene, *SCARECROW* (*SCR*), is described. The *SCR* gene is expressed specifically in root progenitor tissues of embryos, and in
5 certain tissues of roots and stems. *SCR* expression controls cell division of certain cell types in roots, and affects the organization of root and stem. The invention relates to the *SCARECROW* (*SCR*) gene (which encompasses the Arabidopsis *SCR* gene and its orthologs and paralogs), *SCR* gene products,
10 (including but not limited to transcriptional products such as mRNAs, antisense and ribozyme molecules, and translational products such as the *SCR* protein, polypeptides, peptides and fusion proteins related thereto), antibodies to *SCR* gene products, *SCR* regulatory regions and the use of the foregoing
15 to improve agronomically valuable plants.

The invention is based, in part, on the discovery, identification and cloning of the gene responsible for the *scarecrow* phenotype. In contrast to the prevailing view that the *SCR* gene was likely to be involved in the specification
20 of endodermis, the inventors have determined that the mutant cell layer in roots of *scr* mutants has differentiated characteristics of both cortex and endodermis. This is consistent with a role for *SCR* in the regulation of the asymmetric cell division rather than in specification of the
25 identity of either cortex or endodermis. The inventors have also determined that *SCR* expression affects the gravitropism of plant aerial structures such as the stem.

One aspect of the invention relates to the heterologous expression of *SCR* genes and related nucleotide
30 sequences, and specifically the Arabidopsis *SCR* genes, in stably transformed higher plant species. Modulation of *SCR* expression levels can be used to advantageously modify root and aerial structures of transgenic plants and enhance the agronomic properties of such plants.

35 Another aspect of the invention relates to the use of promoters of *SCR* genes, and specifically the use of Arabidopsis *SCR* promoter to control the expression of protein

and RNA products in plants. Plant SCR promoters have a variety of uses, including but not limited to expressing heterologous genes in the embryo, root, root nodule, and stem of transformed plants.

5 The invention is illustrated by working examples described *infra* which demonstrate the isolation of the Arabidopsis SCR gene using insertion mutagenesis. More specifically, T-DNA tagging of genomic and cDNA clones of the Arabidopsis SCR gene are described. Additional working
10 examples include the isolation of SCR sequences from plant genomes using PCR amplification in combination with screening of genomic libraries, and heterologous gene expression in transgenic plants using SCR promoter expression constructs.

 Structural analysis of the deduced amino acid
15 sequence of Arabidopsis SCR protein indicates that SCR encodes a transcription factor. Northern analysis, *in situ* hybridization analysis and enhancer trap analysis show highly localized expression of Arabidopsis SCR in embryos and roots. Genetic analysis shows SCR expression also affects
20 gravitropism of aerial structures (e.g., stems). This indicates that SCR is also expressed in those structures.

 Computer analysis of the deduced amino acid sequence of Arabidopsis SCR protein with those of Expressed Sequence Tag (EST) sequences in GenBank reveals the existence
25 of at least thirteen SCR genes in Arabidopsis, one SCR gene in maize, four SCR genes in rice, and one SCR gene in Brassica. A further aspect of the invention relates to the use of such EST sequences to obtain larger and/or complete clones of the corresponding SCR gene.

30 The various embodiments of the claimed invention presented herein are by the way of illustration and are not meant to limit the invention.

3.1. DEFINITIONS

35 As used herein, the terms listed below will have the meanings indicated.

- 35S = cauliflower mosaic virus promoter for the 35S transcript
- cDNA = complementary DNA
- cis-regulatory
5 element = A promoter sequence 5' upstream of the TATA box that confers specific regulatory response to a promoter containing such an element. A promoter may contain one or more cis-regulatory elements, each responsible for a particular regulatory response
- 10 coding
sequence = sequence that encodes a complete or partial gene product (e.g., a complete protein or a fragment thereof)
- DNA = deoxyribonucleic acid
- EST = expression tagged
- 15 functional
portion = a functional portion of a promoter is any portion of a promoter that is capable of causing transcription of a linked gene sequence, e.g., a truncated promoter
- 20 gene
fusion = a gene construct comprising a promoter operably linked to a heterologous gene, wherein said promoter controls the transcription of the heterologous gene
- gene
25 product = the RNA or protein encoded by a gene sequence
- gene
sequence = sequence that encodes a complete gene product (e.g., a complete protein)
- GUS = 1,3- β -Glucuronidase
- 30 gDNA = genomic DNA
- heterologous
gene = In the context of gene constructs, a heterologous gene means that the gene is linked to a promoter that said gene is not naturally linked to. The heterologous gene may or may not be from the organism contributing said promoter. The heterologous gene may encode messenger RNA (mRNA), antisense RNA or ribozymes
- 35

homologous
promoter = a native promoter of a gene that selectively hybridizes to the sequence of a *SCR* gene described herein

mRNA = messenger RNA

5 operably
linked = A linkage between a promoter and gene sequence such that the transcription of said gene sequence is controlled by said promoter

10 ortholog = related gene in a different plant (e.g., maize *ZCARECROW* gene is an ortholog of the Arabidopsis *SCR* gene)

paralog = related gene in the same plant (e.g., Arabidopsis *SRPa1* is a paralog of Arabidopsis *SCR* gene)

RNA = ribonucleic acid

15 RNase = ribonuclease

SCR
(italic) = *SCARECROW* gene or gene product, encompasses *SCR* and *ZCR* genes and their orthologs and paralogs

20 *SCR* = *SCARECROW* protein

scr
(lower case) = *scarecrow* mutant (e.g., *scr1*)

ZCR = maize *ZCARECROW* gene, a paralog of, for example, the Arabidopsis *SCR* gene

25 *SCR* protein means a protein containing sequences or a domain substantially similar to one or more motifs (i.e., Motif I-VI), preferably MOTIF III (VHIID), of Arabidopsis *SCR* protein as shown in FIGS. 13A-F and FIGS. 15A-S. *SCR* proteins include *SCR* ortholog and paralog proteins having the structure and activities described herein.

30 *SCR* polypeptides and peptides include deleted or truncated forms of the *SCR* protein, and fragments corresponding to the *SCR* motifs described herein.

35 *SCR* fusion proteins encompass proteins in which the *SCR* protein or an *SCR* polypeptide or peptide is fused to a heterologous protein, polypeptide or peptide.

SCR gene, nucleotides or coding sequences means nucleotides, e.g., gDNA or cDNA encoding SCR protein, SCR polypeptides or peptides, or SCR fusion proteins.

SCR gene products include transcriptional products such as mRNAs, antisense and ribozyme molecules, as well as translational products of the SCR nucleotides described herein including but not limited to the SCR protein, polypeptides, peptides and/or SCR fusion proteins.

SCR promoter means the regulatory region native to the SCR gene in a variety of species, which promotes the organ and tissue specific pattern of SCR expression described herein.

4. BRIEF DESCRIPTION OF THE FIGURES

FIGS. 1A-B. Schematic of Arabidopsis root anatomy. FIG. 1A. Transverse section showing the four tissues, epidermis, cortex, endodermis and pericycle that surround the vascular tissue. In the longitudinal section, the epidermal/lateral root cap initials and the cortex/endodermal initials are shown at the base of their respective cell files. FIG. 1B. Schematic of division pattern of the cortex/endodermal initial. The initial expands then divides anticlinally to reproduce itself and a daughter cell. The daughter then divides periclinally to produce the progenitors of the endodermis and cortex cell lineages. Abbreviations: C, cortex; Da, daughter cell; E, endodermis; In, initial.

FIGS. 2A-F. Phenotype of *scr* mutant plants.

FIG. 2A. Shown left to right are 12-day *scr-2*, *scr-1* and wild-type seedlings grown vertically on nutrient agar medium. FIG. 2B. 21-day *scr-2* mutant plants in soil. FIG. 2C. Transverse section through primary root of 7-day *scr-2*. FIG. 2D. Transverse section through primary root of 7-day wild-type (WT). FIG. 2E. Transverse section through lateral root of 12-day *scr-1* mutant seedling. FIG. 2F. Transverse section through root regenerated from *scr-1* callus. Bar, 50 μ m. Abbreviations: C, cortex; En, endodermis; Ep, epidermis; M, mutant cell layer; P, pericycle; V, vascular tissue.

FIGS. 3A-F. Characterization of the cellular identity of the mutant cell layer. FIG. 3A. Endodermis-specific Casparian band staining of transverse sections through the primary root of 7-day *scr-1* mutant. (Note: the histochemical stain also reveals xylem cells in the vascular cylinder.) FIG. 3B. Casparian band staining of transverse sections through the primary root of 7-day wild-type (WT). FIG. 3C. Immunostaining with the endodermis (and a subset of vascular tissue) specific JIM13 monoclonal antibodies on transverse root sections of *scr-2* mutant. FIG. 3D. Immunostaining with JIM13 monoclonal antibodies on transverse root sections of WT. FIG. 3E. Immunostaining with the JIM7 monoclonal antibody that stains all cell walls on transverse root sections of *scr-2* mutant. FIG. 3F. Immunostaining with JIM7 monoclonal antibodies on transverse root sections of WT. Bar, 25 μ m. Abbreviations are same as those for description of FIGS. 2A-2F and: Ca, casparian strip.

FIGS. 4A-F. Immunostaining. FIG. 4A. Immunostaining with the cortex (and epidermis) specific CCRC-M2 monoclonal antibodies on transverse root sections of *scr-1* mutant. FIG. 4B. Immunostaining with CCRC-M2 antibodies on transverse root sections of *scr-2* mutant. FIG. 4C. Immunostaining with CCRC-M2 antibodies on transverse root sections of wild-type (WT). FIG. 4D. Immunostaining with the CCRC-M1 monoclonal antibodies (specific to a cell wall epitope found on all cells) on transverse root sections of *scr-1*. FIG. 4E. Immunostaining with CCRC-M1 antibodies on transverse root sections of *scr-2*. FIG. 4F. Immunostaining with CCRC-M1 antibodies on transverse root sections of WT. Bar, 30 μ m. Abbreviations are same as those for description of FIGS. 2A-2F.

FIG. 5A-E. Structure of the Arabidopsis SCARECROW gene. FIG. 5A. Nucleic acid sequence and deduced amino acid sequence of the Arabidopsis SCR genomic region (SEQ ID NO:1) and (SEQ ID NO:2), respectively. Regulatory sequences including: (i) TATA box, (ii) ATG start codon, and (iii) potential polyadenylation sequence are underlined. Within

the deduced amino acid sequence homopolymeric repeats are underlined. FIG. 5B. Schematic diagram of genomic clone indicating possible functional motifs, T-DNA insertion sites and subclones used as probes. Abbreviations: Q,S,P,T, region with homopolymeric repeats of these amino acids; b, region with similarity to the basic region of bZIP factors; I and II, regions with leucine heptad repeats; E, acidic region. FIG. 5C. Comparison of the charged region found in Arabidopsis SCR protein with that found in bZIP transcription factors, SCR bZIP-like domain (SEQ ID NO:3), GCN4 (SEQ ID NO:4), TGA1 (SEQ ID NO:5), C-Fos (SEQ ID NO:6), c-JUN (SEQ ID NO:7), CREB (SEQ ID NO:8), Opaque-2 (SEQ ID NO:9), OBF2 (SEQ ID NO:10), RAF-1 (SEQ ID NO:11). FIG. 5D. Translations of EST clones encoding putative peptide having similarities to the VHIID domain region of Arabidopsis SCR protein (SEQ ID NO:12), F13896 (SEQ ID NO:13), Z37192 (SEQ ID NO:14), and Z25645 (SEQ ID NO:15) are from Arabidopsis, T18310 (SEQ ID NO:17) is from maize and D41474 (SEQ ID NO:16) is from rice. FIG. 5E. The deduced amino acid sequence of the Arabidopsis SCARECROW gene (SEQ ID NO:2).

FIGS. 6A-B. Expression of the Arabidopsis SCARECROW gene. FIG. 6A. Northern blot of total RNA from wild-type siliques (Si), roots (R), leaves (L) and whole seedlings (Sd) hybridized with Arabidopsis SCR probe a and with a probe from the Arabidopsis glutamine dehydrogenase (GDH) gene (Melo-Oliveira et al., 1996, Proc. Natl. Acad. Sci. USA 93:4718-4723) as a control for RNA integrity. (GDH expression is lower in siliques than in vegetative tissues.) The 1.6 kb band corresponds to the GDH gene and the approximately 2.5 kb band corresponds to SCR. Ribosomal RNA is shown as a loading control. FIG. 6B. Northern blot of Arabidopsis wild-type, scr-1 and scr-2 total RNA, probed with Arabidopsis SCR probe "a" corresponding to a cDNA sequence shown in FIG. 5B, and with the GDH probe. In scr-2 mutant additional bands of 4.1 kb and 5.0 kb were detected.

FIGS. 7A-G. In situ hybridization and enhancer trap analyses of Arabidopsis SCR expression. FIG. 7A. SCR

RNA expression detected by *in situ* hybridization of *SCR* antisense probe to a longitudinal section through the root meristem. FIG. 7B. *In situ* hybridization of *SCR* antisense probe to a transverse section in the meristematic region.

5 FIG. 7C. *In situ* hybridization of *SCR* antisense probe to late torpedo stage embryo. FIG. 7D. Negative control *in situ* hybridization using a *SCR* sense probe to a longitudinal section through the root meristem. FIG. 7E. GUS expression in a whole mount in the enhancer trap line, ET199 in primary
10 root tip. FIG. 7F. GUS expression in the ET199 line in transverse root section in the meristematic region. FIG. 7G. GUS expression in ET199 detected in a section through the root meristem. GUS expression is observed in the cortex/endodermal initial, and in the first cell in the
15 endodermal cell lineage but not in the first cell of the cortex lineage. Expression in two endodermal layers is observed higher up in the root because the section was not median at that point. Bar, 50 μ m. Abbreviations are same as those in the description of FIGS. 2A-2F.

20 FIG. 8. Partial nucleotide sequence (SEQ ID NO:18) and deduced amino acid sequence (SEQ ID NO:19) of the Arabidopsis *SRPa4* gene.

FIG. 9. Partial nucleotide sequence (SEQ ID NO:20) and deduced amino acid sequence (SEQ ID NO:21) of the
25 Arabidopsis *SRPa3* gene.

FIG. 10. Partial nucleotide sequence (SEQ ID NO:22) of the Arabidopsis *SRPa1* gene.

FIG. 11A. Nucleotide sequence (SEQ ID NO:24) and deduced amino acid sequence (SEQ ID NO:25) of the maize Zm-
30 *Sc11* fragment.

FIG. 11B. Partial nucleotide sequence (SEQ ID NO:25) and deduced amino acid sequence (SEQ ID NO:26) of the maize *SRPm1* gene (Zm-*Sc12*).

FIG. 12A-B. Nucleotide sequence of rice *SRPo3* EST
35 clone. FIG. 12A. Sequence of 5' end of EST clone (SEQ ID NO:28). FIG. 12B. Sequence of 3' end of EST clone (SEQ ID NO:29).

FIGS. 13A-F. Comparison of the amino acid sequence of members of the *SCARECROW* family of genes. Conserved Motifs I through VI are indicated by dashed line above the aligned sequences. Consensus sequences are shown in bold. See Table 1 for the identity and sequence identifier number of each of the sequences shown in this Figure. Hu-scr-1 = Human SCR paralog (SEQ ID NO:40).

FIG. 14. Restriction map of the approximately 8.8 kb Eco RI insert DNA of lambda clone, t643, containing the Arabidopsis SCR gene. The locations of the approximately 5.6 kb HindIII-SacI fragment subcloned in plasmid LIG 1-3/SAC+MoB₂ 1SAC, and the SCR coding region are indicated below the restriction map. The location of the translational initiation site of the SCR gene is at the Nco I site at the left end of the indicated coding region. The SCR coding sequence begins at the translation initiation site and extends approximately 1955 nucleotides to its right. *E. coli* DH5 α containing plasmid pLIG1-3/SAC+MoB₂ 1SAC, has the ATCC accession number 98031.

FIGS. 15A-S. Comparison of the partial and complete amino acid sequences of several plant members of the *SCARECROW* family of genes. The amino acid sequences are aligned in a manner that maximizes amino acid sequence similarity and identity among SCR family members. Each sequence shown is continuous except where noted otherwise; the dots are inserted between two sequence segments in order to align homologous segments. "X" in the middle of a sequence indicates ambiguity in the corresponding nucleotide sequence and, possible termination of the ORF at the "X" residue site. "X" at the end of a sequence indicates termination of the ORF at the "X" residue site. The numbering of the amino acid residues is shown at the bottom of each figure and is based on the Arabidopsis SCR amino acid sequence. Conserved Motifs I through VI are indicated by the various dashed lines above the figures. The new and old names of the family members are shown in FIG. 15A. The sequences of SCR, Tf1 and Tf4 are of the complete SCR

protein. See Table 1 for the identity and the sequence identifier number of each sequence shown in these figures.

FIGS. 16A-M. The partial nucleotide sequences of several plant members of the *SCARECROW* family of genes. "N" indicates an unknown base. See Table 1 for the identity and the sequence identifier number of each sequence shown in these figures.

FIG. 17A. The partial nucleotide sequence (SEQ ID NO:66) of the maize ZCR gene.

10 FIG. 17B. The partial amino acid sequence (SEQ ID NO:67) of the maize ZCR gene. The underlined sequence shares approximately 80% sequence identity with a corresponding sequence of Arabidopsis SCR protein.

FIG. 18. Comparison of the partial amino acid sequences of several SCR ortholog sequences amplified from the genomes of carrot, soybean and spruce. The *SRPd1* and *SRPp1* sequences each were obtained by PCR amplification using a combination of 1F and 1R primers. The *SRPg1* sequence was obtained by PCR amplification using a combination of 1F and 20 WP primers. The amino acid sequences are aligned in a manner that maximizes amino acid sequence identity and similarity amongst these sequences. Each sequence shown is continuous except where noted otherwise; the dashes are inserted between two sequence segments in order to allow 25 alignment of homologous segments. "x" in the middle of a sequence indicates ambiguity in the corresponding nucleotide sequence and, possible termination of the ORF or existence of an intron at the "x" residue site. See Table 1 for the identity and the sequence identifier number of each sequence 30 shown in this figure.

FIG. 19. Comparison of promoter activities in transgenic lines and roots. Panel a. A stably transformed line containing four copies of the B2 subdomain of the 35S promoter of CaMV upstream of GUS (Benfey et al., 1990). GUS 35 is expressed in the root tip. Panel b. Roots emerging from callus transformed with four copies of the B2 subdomain of the 35S promoter fused to GUS. GUS expression can be seen in

the emerging root tips (arrows). Panel c. Higher magnification of a root emerging from the callus in panel b. GUS is clearly restricted to the root tip. The morphology of roots regenerated from calli often appears abnormal. Panel d. A transgenic plant regenerated from the calli and roots shown in panel b. GUS expression in this plants appears to be similar to that of the original line shown in panel a. Panel e. ET199, a stably transformed line that contains an enhancer trapping construct with a minimal promoter fused to the GUS coding region inserted 1 kb upstream from the SCR coding region. GUS expression is primarily in the endodermal layer of the root. Panel f. Roots emerging from calli transformed with the SCR promoter::GUS construct. Expression of the GUS gene appears to be limited to an internal layer (arrows). Panel g. SCR promoter::GUS transformed root in liquid culture. Roots shown in panel f were excised and transferred to liquid cultures. GUS expression is primarily found in the endodermal layer as in ET199. The expression of GUS in the quiescent center, as seen here, is also sometimes observed in ET199. Bar, 50 μ m.

FIG. 20. Analysis of SCR promoter activity in the scr mutant background. Panel a. Roots emerging from scr calli transformed with the SCR promoter::GUS construct. Roots regenerated from scr calli are very short. GUS expression appears to be limited to an internal layer of the root (arrows). Panel b. Root regenerated from transformed scr calli and transferred to liquid culture. The scr phenotype, a single layer between the epidermis and pericycle, is easily seen. GUS expression is limited to this mutant layer. E, Epidermis. M, Mutant Layer. P, Pericycle. Bar, 50 μ m.

FIG. 21. Molecular Complementation of the scr mutant. Panels a, c and e. scr transformed with the SCR promoter::GUS construct. Panels b, d and f. scr transformed with the SCR promoter::SCR coding region construct. Panels a and b. Roots emerging from scr calli. Arrows point to several very short roots among many fine root hairs in the

scr calli transformed with the SCR promoter::GUS construct. In contrast, roots from scr calli transformed with the SCR promoter::SCR coding region construct appeared to be wild-type in length, suggesting molecular complementation by the transgene. Panels c and d. Transgenic roots in liquid culture. The scr roots transformed with the SCR promoter::GUS construct appeared short, while those transformed with the SCR promoter::SCR coding region construct appeared of wild-type length. Panels e and f. Transverse sections through roots emerging from calli. Whereas there is only a single cell layer between the epidermis and stele in the SCR promoter::GUS transformed root, the radial organization of the root transformed with the SCR promoter::SCR coding region appeared identical to wild-type, with both cortex and endodermal layers. E, epidermis. M, mutant layer. C, cortex. En, Endodermis. P, Pericycle. Bar, 50µm

FIG. 22. Expression of ZCR in maize root tips.

Left Panel. Expression of ZCR is in the endodermal layer and extends down through the region of the quiescent center. Right Panel. Higher magnification showing expression in a single cell layer through the quiescent center.

5. DETAILED DESCRIPTION OF THE INVENTION

The invention relates to the SCARECROW (SCR) gene, SCR gene products, including but not limited to transcriptional products such as mRNAs, antisense and ribozyme molecules, and translational products such as the SCR protein, polypeptides, peptides and fusion proteins related thereto; antibodies to SCR gene products; SCR regulatory regions; and the use of the foregoing to improve agronomically valuable plants.

In summary, the data described herein show the identification of SCR, a gene involved in the regulation of a specific asymmetric division, in controlling gravitropic response in aerial structures, and in controlling pattern

formation in roots. Sequence analysis shows that the SCR protein has many hallmarks of transcription factors. *In situ* and marker line expression studies show that SCR is expressed in the cortex/endodermal initial of roots before asymmetric division occurs, and in quiescent center of regenerating roots. Together, these findings indicate that SCR gene regulates key events that establish the asymmetric division that generates separate cortex and endodermal cell lineages, and that affect tissue organization of roots. The establishment of these lineages is not required for cell differentiation to occur, because in the absence of division the resulting cell acquires mature characteristics of both cortex and endodermal cells. However, it is possible that SCR functions to establish the polarity of the initial before cell division, or that it is involved in generating an external polarity that has an effect on asymmetric cell division.

Genetic analysis indicates that SCR expression affects gravitropism of plant stems and hypocotyls. This indicates that SCR is also expressed in these aerial structures of plants.

The SCR genes and promoters of the present invention have a number of important agricultural uses. The SCR promoters of the invention may be used in expression constructs to express desired heterologous gene products in the embryo, root, root nodule, and starch sheath layer in stem of transgenic plants transformed with such constructs. For example, SCR promoters may be used to express disease resistance genes such as lysozymes, cecropins, maganins, or thionins for anti-bacterial protection or the pathogenesis-related (PR) proteins such as glucanases and chitinases for anti-fungal protection. SCR promoters also may be used to express a variety of pest resistance genes in the aforementioned plant structures and tissues. Examples of useful gene products for controlling nematodes or insects include *Bacillus thuringiensis* endotoxins, protease

inhibitors, collagenases, chitinase, glucanases, lectins, and glycosidases.

Gene constructs that express or ectopically express *SCR*, and the *SCR*-suppression constructs of the invention may
5 be used to alter the root and/or stem structure, and the gravitropism of aerial structures of transgenic plants. Since *SCR* regulates root cell divisions, overexpression of *SCR* can be used to increase division of certain cells in roots and thereby form thicker and stronger roots. Thicker
10 and stronger roots are beneficial in preventing plant lodging. Conversely, suppression of *SCR* expression can be used to decrease cell division in roots and thereby form thinner roots. Thinner roots are more efficient in uptake of soil nutrients. Since *SCR* affects gravitropism of aerial
15 structures, overexpression of *SCR* may be used to develop "straighter" transgenic plants that are less susceptible to lodging.

Further, *SCR* gene sequence may be used as a molecular marker for a qualitative trait, e.g., a root or
20 gravitropism trait, in molecular breeding of crop plants.

For purposes of clarity and not by way of limitation, the invention is described in the subsections below in terms of (a) *SCR* genes and nucleotides; (b) *SCR* gene products; (c) antibodies to *SCR* gene products; (d) *SCR*
25 promoters and promoter elements; (e) transgenic plants which ectopically express *SCR*; (f) transgenic plants in which endogenous *SCR* expression is suppressed; and (g) transgenic plants in which expression of a transgene of interest is controlled by *SCR* promoter.

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5.1. SCR GENES

The *SCARECROW* genes and nucleotide sequences of the invention include: (a) a gene listed below in Table 1 (hereinafter, a gene comprising any one of the nucleotide
35 sequences shown in FIG. 5A, FIG. 8, FIG. 9, FIG. 10, FIGS. 11A-B, FIGS. 12A-B, FIGS. 16A-M, or FIG. 17A, or a segment of such nucleotide sequences), or as contained in the clones

described herein and deposited with the ATCC (see Section 13, infra); (b) nucleotide sequence that encodes a protein comprising any one of the amino acid sequences shown in FIG. 5A, FIG. 5D, FIG. 5E, FIG. 8, FIG. 9, FIGS. 11A-B, FIGS. 13A-5 F, FIGS. 15A-S, FIG. 17B or FIG. 18 or a segment of such amino acid sequences, or that is encoded by any one of the genes and/or nucleotide sequences listed by their sequence identifier numbers in Table 1, or any segment of such genes and/or nucleotide sequences, or contained in any one of the 10 clones described herein and deposited with the ATCC (see Section 13, infra); (c) any gene comprising nucleotide sequence that hybridizes to the complement of any one of the genes and/or nucleotide sequences listed by their sequence identifier numbers in Table 1, or any segment of such genes 15 and/or nucleotide sequences, or as contained in any one of the clones described herein and deposited with the ATCC, under highly stringent conditions, e.g., hybridization to filter-bound DNA in 0.5 M NaHPO₄, 7% sodium dodecyl sulfate (SDS), 1 mM EDTA at 65°C, and washing in 0.1xSSC/0.1% SDS at 20 68°C (Ausubel F.M. et al., eds., 1989, Current Protocols in Molecular Biology, Vol. I, Green Publishing Associates, Inc., and John Wiley & sons, Inc., New York, at p. 2.10.3) and that encodes a gene product functionally equivalent to SCR gene product encoded completely or partly by any one of the genes 25 and/or sequences listed in Table 1 or any segment of such genes and nucleotide sequences, or as contained in any one of the clones deposited with the ATCC; (d) any gene comprising nucleotide sequence that hybridizes to the complement of any one of the sequences listed by their sequence identifier 30 numbers in Table 1, or any segment of such nucleotide sequences, or as contained in any one of the clones described herein and deposited with the ATCC, under less stringent conditions, such as moderately stringent conditions, e.g., washing in 0.2xSSC/0.1% SDS at 42°C (Ausubel et al., 1989, 35 supra), and which encodes a functionally equivalent SCR gene product; (e) any gene comprising nucleotide sequence that hybridizes to the complement of any one of the sequences

listed by their sequence identifier numbers in Table 1 or any segment of such nucleotide sequences, or as contained in any one of the clones described herein and deposited with the ATCC, under the following low stringency conditions: pre-
5 hybridization in hybridization solution (HS) containing 43% formamide, 5xSSC, 1% SDS, 10% dextran sulfate, 0.1% sarkosyl, 2% block (Genius kit, Boehringer-Mannheim), followed by hybridization overnight at 30 to 33°C using as a probe a DNA molecule of approximately 1.6 kb of SEQ ID NO:1 at a
10 concentration of 20 ng/ml, followed by washing in 2xSSC/0.1% SDS two times for 15 minutes at room temperature and then two times at 50°C, and which encodes a functionally equivalent SCR gene product; and/or (f) any gene comprising nucleotide sequence that encodes a polypeptide or protein containing the
15 consensus sequence for SCR (i.e., MOTIF III or VHIIID) shown in FIGS. 13B-D or a segment of such polypeptide or protein. The partial and complete nucleotide and amino acid sequences of SCR genes and encoded proteins and polypeptides included in the invention are listed in Table 1 below.

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Table 1. SCR ORTHOLOGS AND PARALOGS

	<u>New Name</u>	<u>Old Name</u>	<u>EST Clone¹</u>	<u>SEQ ID NOS</u> <u>Nucleotide³</u>	<u>Amino Acid</u>
5	<u>ARABIDOPSIS</u>				
	SRPa1	1110	Z25645/33772	22	23
	SRPa2	Tf4	Z34599	--	35*
	SRPa3	3935	Z37192/1 N96166	20	21
10	SRPa4	4818	F13896/7	18	19
	SRPa5	4871	F13949	45	46
	SRPa6	12398	R29793	51	52
	SRPa7	3635	T21627 H76979 N96767	55	56
15	SRPa8	Tf1	T46205 (9468) N96653 (21711)	--	34*
	SRPa9	10964	T78186 T44774	47	48
	SRPa10	11261	T76483	49	50
20	SRPa11	18652	N37425	53	54
	SRPa12	23196	W43803 W435138 AA042397	57	58
	SRPa13	33/08	T46008	--	41
25	SCR	Scr	N.A. ²	1 ⁺	2*
	<u>RICE</u>				
	SRPo1	713	D15490	--	43
	SRPo2	2504	D40482 D40607 D40800 D41389	--	44
30	SRPo3	3989	D41474	--	36
	SRPo4	11846	C20324	--	59
	<u>MAIZE</u>				
35	SRPm1	18310	T18310	--	37
	<u>BRASSICA</u>				
	SRPb1	174	H74669	--	42

Table 1. (Continued)

	<u>New Name</u>	<u>Old Name</u>	<u>EST Clone¹</u>	<u>SEQ ID NOS</u> <u>Nucleotide²</u>	<u>Amino Acid</u>
	<u>CARROT</u>				
	<i>SRPd1</i>	N.A.	N.A.	60	61
5	<u>SOYBEAN</u>				
	<i>SRPg1</i>	N.A.	N.A.	62	63
	<u>SPRUCE</u>				
	<i>SRPp1</i>	N.A.	N.A.	64	65

¹ Each EST clone is identified by its GenBank accession number. Each EST clone corresponds to a deposit of a cDNA sequence that matches a part of the nucleotide sequence of the corresponding SCR ortholog or paralog.

² N.A. = not applicable.

³ The partial or complete nucleotide sequence of the SCR orthologs and paralogs listed here are shown in FIGS. 5A, 8, 9, 10, 11A-B, 12A-B, 16A-M and 17A.

⁴ Contains the complete coding sequence of Arabidopsis SCR gene.

⁵ Contains the complete amino acid sequence of Arabidopsis SRPa2, SRPa8, or SCR protein.

Functional equivalents of the *SCR* gene product include any plant gene product that regulates plant embryo or root development, or, preferably, that regulates root cell division or root tissue organization, or affects gravitropism of plant aerial structures (e.g., stems and hypocotyls). Functional equivalents of the *SCR* gene product include naturally occurring *SCR* gene products, and mutant *SCR* gene products, whether naturally occurring or engineered.

The invention also includes nucleic acid molecules, preferably DNA molecules, that hybridize to, and are therefore the complements of the nucleotide sequences (a) through (f), in the first paragraph of this section. Such hybridization conditions may be highly stringent, less highly stringent, or low stringency as described above. In instances wherein the nucleic acid molecules are oligonucleotides ("oligos"), highly stringent conditions may refer, e.g., to washing in 6xSSC/0.05% sodium pyrophosphate at 37°C (for 14-base oligos), 48°C (for 17-base oligos), 55°C (for 20-base oligos), and 60°C (for 23-base oligos). These nucleic acid molecules may act as *SCR* antisense molecules, useful, for example, in *SCR* gene regulation and/or as antisense primers in amplification reactions of *SCR* gene and/or nucleic acid sequences. Further, such sequences may be used as part of ribozyme and/or triple helix sequences, also useful for *SCR* gene regulation. Still further, such molecules may be used as components in probing methods whereby the presence of a *SCARECROW* allele may be detected.

The invention also includes nucleic acid molecules, preferably DNA molecules, which are amplified using the polymerase chain reaction under conditions described in Section 5.1.1., *infra*, and that encode a gene product functionally equivalent to a *SCR* gene product encoded by any one of the genes and sequences listed in Table 1 or as contained in any one of the clones described herein and deposited with the ATCC.

The invention also encompasses (a) DNA vectors that contain any of the foregoing gene and/or coding sequences

and/or their complements (i.e., antisense or ribozyme molecules); (b) DNA expression vectors that contain any of the foregoing gene and/or coding sequences operatively associated with a regulatory element that directs the expression of the gene and/or coding sequences; and (c) genetically engineered host cells that contain any of the foregoing gene and/or coding sequences operatively associated with a regulatory element that directs the expression of the gene and/or coding sequences in the host cell. As used herein, regulatory elements include but are not limited to inducible and non-inducible promoters, enhancers, operators and other elements known to those skilled in the art that drive and regulate expression.

The invention also encompasses nucleotide sequences that encode mutant SCR gene products, peptide fragments of the SCR gene product, truncated SCR gene products, and SCR fusion proteins. These gene products include, but are not limited to, nucleotide sequences encoding mutant SCR gene products; polypeptides or peptides corresponding to one or more of the Motifs I-VI as shown in FIGS. 13A-F and FIGS. 15A-S, or the bZIP, VHIID, or leucine heptad domains of the SCR, or portions of these motifs and domains; truncated SCR gene products in which one or more of the motifs or domains is deleted, e.g., a truncated, nonfunctional SCR lacking all or a portion of the Motifs I-VI as shown in FIGS. 13A-F and FIGS. 15A-S, or the bZIP, VHIID, or leucine heptad domains of the SCR. Nucleotides encoding fusion proteins may include but are not limited to full length SCR, truncated SCR or peptide fragments of SCR fused to an unrelated protein or peptide, such as for example, an enzyme, fluorescent protein, or luminescent protein which can be used as a marker.

In particular, the invention includes, for example, fragments of SCR genes encoding one or more of the following

domains as shown in FIG. 5E: amino acids 1-264, 265-283, 287-316, 410-473, 436-473, and 473-653.

In addition to the gene and/or coding sequences described above, homologous *SCR* genes, and other genes related by DNA sequence, may be identified and may be readily isolated, without undue experimentation, by molecular biological techniques well known in the art. More specifically, such homologs include, for example, paralogs (*i.e.*, members of the *SCR* gene family occurring in the same plant) as well as orthologs (*i.e.*, members of the *SCR* gene family which occur in a different plant species) of the *Arabidopsis SCR* gene.

A specific embodiment of a *SCR* gene and coding sequence of the invention is *Arabidopsis SCR* (FIGS. 5A and 5E). Other specific embodiments include the various *SCR* genes and coding sequences listed in Table 1, *supra*.

Methods for isolating *SCR* genes and coding sequences are described in detail in Section 5.2, below.

SCR genes share substantial amino acid sequence similarities at the protein level and nucleotide sequence similarities in their encoding genes. The term "substantially similar" or "substantial similarity" when used herein with respect to two amino acid sequences means that the two sequences have at least 75% identical residues, preferably at least 85% identical residues and most preferably at least 95% identical residues. The same term when used herein with respect to two nucleotide sequences means that the two sequences have at least 70% identical residues, preferably at least 85% identical residues and most preferably at least 95% identical residues. Determining whether two sequences are substantially similar may be carried out using any methodologies known to one skilled in the art, preferably using computer assisted analysis. For example, the alignments showed herein were initially accomplished by a BLAST search (NCBI using the BLAST network server). The final alignments of *SCR* family members were done manually.

Moreover, SCR genes show highly localized expression in embryos and, particularly, roots. Such expression patterns may be ascertained by Northern hybridizations and in situ hybridizations using antisense probes.

5.1.1. ISOLATION OF SCR GENES

The following methods can be used to obtain SCR genes and coding sequences from a wide variety of plants, including but not limited to *Arabidopsis thaliana*, *Zea mays*, *Nicotiana tabacum*, *Daucus carota*, *Oryza*, *Glycine max*, *Lemna gibba*, and *Picea abies*.

Nucleotide sequences encoding an SCR gene or a portion thereof may be obtained by PCR amplification of plant genomic DNA or cDNA. Useful cDNA sources include "free" cDNA preparations (i.e., the products of cDNA synthesis) and cloned cDNA in cDNA libraries. Root cDNA preparations or libraries are particularly preferred.

The amplification may use, as the 5'-primer (i.e., forward primer), a degenerate oligonucleotide that corresponds to a segment of a known SCR amino acid sequence, preferably from the amino-terminal region. The 3'-primer (i.e., reverse primer) may be a degenerate oligonucleotide that corresponds to a distal segment of the same known SCR amino acid sequence (i.e., carboxyl to the sequence that corresponds to the 5'-primer). For example, the amino acid sequence of the *Arabidopsis* SCR protein (SEQ ID NO:2) may be used to design useful 5' and 3' primers. Preferably, the primers corresponds to segments in the Motif III or VHIID domain of SCR protein (see FIGS. 13B-D and FIGS. 15K-L). The sequence of the optimal degenerate oligonucleotide probe corresponding to a known amino acid sequence may be determined by standard algorithms known in the art. See for example, Sambrook et al., Molecular Cloning: A Laboratory Manual, 2nd ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, Vol 2 (1989).

Further, for amplification from cDNA sources, the 3'-primer may be an oligonucleotide comprising an 3' oligo(dT) sequence. The amplification may also use as primers nucleotide sequences of *SCR* genes or coding sequences (e.g., any one of the *scr* sequences and EST sequences listed in Table 1).

PCR amplification can be carried out, e.g., by use of a Perkin-Elmer Cetus thermal cycler and Taq polymerase (Gene Amp[®]). One can choose to synthesize several different degenerate primers for use in the PCR reactions. It is also possible to vary the stringency of hybridization conditions used in priming the PCR reactions, to allow for greater or lesser degrees of nucleotide sequence similarity between the degenerate primers and the corresponding sequences in the cDNA library. One of ordinary skill in the art will know that the appropriate amplification conditions and parameters depend, in part, on the length and base composition of the primers and that such conditions may be determined using standard formulae. Protocols for executing all PCR procedures discussed herein are well known to those skilled in the art, and may be found in references such as Gelfand, 1989, PCR Technology, Principles and Applications for DNA Amplification, H.A. Erlich, ed., Stockton Press, New York; and Current Protocols In Molecular Biology, Vol. 2, Ch. 15, Ausubel et al., eds 1988, New York, Wiley & Sons, Inc.

A PCR amplified sequence may be molecularly cloned and sequenced. The amplified sequence may be utilized as a probe to isolate genomic or cDNA clones of a *SCR* gene, as described below. This, in turn, will permit the determination of a *SCR* gene's complete nucleotide sequence, including its promoter, the analysis of its expression, and the production of its encoded protein, as described *infra*.

In a preferred embodiment, PCR amplification of *SCR* gene and/or coding sequences can be carried out according to the following procedure:

PRIMERS:**Forward:**

Name: SCR5AII (23-mer, 2 inosines, 64-mix)
 A.A. code: HFTANQAI
 5 DNA Sequence: 5' CAT/C TTT/C ACI GCI AAT/C CAA/G GCN AT 3'

Name: SCR5B (29-mer, 1 inosine, 144-mix)
 A.A. code: VHIID(L/F)D
 DNA Sequence: 5' ACGTCTCGA GTI CAT/C ATA/C/T ATA/C/T GAT/C
 TTN GA 3'

10 Name: 1F
 A.A. code: LQCAEAV
 DNA Sequence: (T/C)TI CA(A/G) TG(T/C GCI GA(A/G) GCN GT

Reverse:

Name: SCR3AII (23-mer, 2 inosines, 128-mix)
 A.A. code: PGGPP(H/N/K)(V/L/F)R'
 15 DNA Sequence: 5' CG/T CCA/C GTG/T TGG IGG ICC NCC NGG 3'

Name: 1R
 A.A. code: AFQVFNGI
 DNA Sequence: AT ICC (A/G)TT (A/G)AA IAC (C/T)TG (A/G)AA NGC

Name: 4R
 A.A. code: QWPGLFHI
 20 DNA Sequence: AT (A/G)TG (A/G)AA IA(A/G) NCC IGG CCA (C/T)TG

I = inosine
 N = A/C/G/T

Useful primer combinations include the following:
 SCR5AII+SCR3AII; SCR5B+SCR3AII; IF+IR; and IF+4R

25

PCR:

Reaction mixture (volume 50 μ l):

- 5 μ l 10X amplification buffer containing Mg (Boehringer-Mannheim)
- 30 -1 μ l 10 mM dNTP's
- 1 μ l forward primer (stock concentration: 80 pmol/ μ l)
- 1 μ l reverse primer (80 pmol/ μ l)
- DNA (100-300 ng).

Begin reaction with "hot start" in which the enzyme is added to the mix only after a brief denaturation at a high temperature (80°C)

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Cycles:

94°C 30 sec - brief denaturation (to prevent non-specific priming)
 80°C 5 min - apply the enzyme to the tubes (30 tubes/round at maximum)
 94°C 5 min - thorough denaturation
 2 times: 94°C 1 min
 5 64°C 5 min
 72°C 2 min
 2 times: 94°C 1 min
 62°C 5 min
 72°C 2 min
 2 times: 94°C 1 min
 60°C 5 min
 10 72°C 2 min
 (reduce the annealing temperature 2°C in every second round), until 44°C is reached after that:
 40 times: 94°C 20 sec
 48°C 1 min
 72°C 2 min
 15 finally, let cool down to 15°C.

A SCR gene coding sequence may also be isolated by screening a plant genomic or cDNA library using a SCR nucleotide sequence (e.g., the sequence of any of the SCR genes and sequences and EST clone sequences listed in Table 1.) as hybridization probe. For example, the whole or a segment of the Arabidopsis SCR nucleotide sequence (FIG. 5A) may be used. Alternatively, a SCR gene may be isolated from such libraries using as probe a degenerate oligonucleotide that corresponds to a segment of a SCR amino acid sequence. For example, degenerate oligonucleotide probe corresponding to a segment of the Arabidopsis SCR amino acid sequence (FIG. 5E) may be used.

In preparation of cDNA libraries, total RNA is isolated from plant tissues, preferably roots. Poly(A)+ RNA is isolated from the total RNA, and cDNA prepared from the poly(A)+ RNA, all using standard procedures. See, for example, Sambrook et al., Molecular Cloning: A Laboratory Manual, 2d ed., Vol. 2 (1989). The cDNAs may be synthesized with a restriction enzyme site at their 3'-ends by using an appropriate primer and further have linkers or adaptors

attached at their 5'-ends to facilitate the insertion of the cDNAs into suitable cDNA cloning vectors. Alternatively, adaptors or linkers may be attached to the cDNAs after the completion of cDNA synthesis.

5 In preparation of genomic libraries, plant DNA is isolated and fragments are generated, some of which will encode parts of the whole SCR protein. The DNA may be cleaved at specific sites using various restriction enzymes. Alternatively, one may use DNase in the presence of manganese
10 to fragment the DNA, or the DNA can be physically sheared, as for example, by sonication. The DNA fragments can then be separated according to size by standard techniques, including but not limited to, agarose and polyacrylamide gel electrophoresis, column chromatography and sucrose gradient
15 centrifugation.

The genomic DNA or cDNA fragments can be inserted into suitable vectors, including but not limited to, plasmids, cosmids, bacteriophages lambda or T₄, and yeast artificial chromosome (YAC) [See, for example, Sambrook et
20 al., Molecular Cloning: A Laboratory Manual, 2d ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York (1989); Glover, D.M(ed.), DNA Cloning: A Practical Approach, MRL Press, Ltd., Oxford, U.K., Vols. I and II (1985)].

The SCR nucleotide probe, DNA or RNA, should be at
25 least 17 nucleotides, preferably at least 26 nucleotides, and most preferably at least 50 nucleotides in length. The nucleotide probe is hybridized under moderate stringency conditions and washed under moderate, preferably high stringency conditions. Clones in libraries with insert DNA
30 having substantial homology to the SCR probe will hybridize to the probe. Hybridization of the nucleotide probe to genomic or cDNA libraries is carried out using methods known in the art. One of ordinary skill in the art will know that the appropriate hybridization and wash conditions depend on
35 the length and base composition of the probe and that such conditions may be determined using standard formulae. See, for example, Sambrook et al., Molecular Cloning: A Laboratory

Manual, 2nd ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, Vol. 2, (1989) pp 11.45-11.57 and 15.55-15.57.

The identity of a cloned or amplified SCR gene sequence can be verified by comparing the amino acid sequences of its three open reading frames with the amino acid sequence of a SCR gene (e.g., Arabidopsis SCR protein [SEQ ID No:2]). A SCR gene or coding sequence encodes a protein or polypeptide whose amino acid sequence is substantially similar to that of a SCR protein or polypeptide (e.g., the amino acid sequence of any one of the SCR proteins and/or polypeptides shown in FIG. 5A, 5E, FIG. 8, FIG. 9, FIGS. 11A-B, FIGS. 15A-S, FIG. 17B and FIG. 18). The identity of the cloned or amplified SCR gene sequence may be further verified by examining its expression pattern, which should show highly localized expression in the embryo and/or root of the plant from which the SCR gene sequence was isolated.

Comparison of the amino acid sequences encoded by a cloned or amplified sequence may reveal that it does not contain the entire SCR gene or its promoter. In such a case the cloned or amplified SCR gene sequence may be used as a probe to screen a genomic library for clones having inserts that overlap the cloned or amplified SCR gene sequence. A complete SCR gene and its promoter may be reconstructed by splicing the overlapping SCR gene sequences.

5.1.2. EXPRESSION OF SCR GENE PRODUCTS

SCR proteins, polypeptides and peptide fragments, mutated, truncated or deleted forms of SCR and/or SCR fusion proteins can be prepared for a variety of uses, including but not limited to the generation of antibodies, as reagents in assays, the identification of other cellular gene products involved in regulation of root development; etc.

SCR translational products include, but are not limited to those proteins and polypeptides encoded by the SCR gene sequences described in Section 5.1, above. The

invention encompasses proteins that are functionally equivalent to the *SCR* gene products described in Section 5.1. Such a *SCR* gene product may contain one or more deletions, additions or substitutions of *SCR* amino acid residues within
5 the amino acid sequence encoded by any one of the *SCR* gene sequences described, above, in Section 5.1, but which result in a silent change, thus producing a functionally equivalent *SCR* gene product. Amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility,
10 hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues involved.

For example, nonpolar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan, and methionine; polar neutral
15 amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine; positively charged (basic) amino acids include arginine, lysine, and histidine; and negatively charged (acidic) amino acids include aspartic acid and glutamic acid. "Functionally equivalent", as
20 utilized herein, refers to a protein capable of exhibiting a substantially similar *in vivo* activity as the endogenous *SCR* gene products encoded by the *SCR* gene sequences described in Section 5.1, above. Alternatively, "functionally equivalent" may refer to peptides capable of regulating gene expression
25 in a manner substantially similar to the way in which the corresponding portion of the endogenous *SCR* gene product would.

The invention also encompasses mutant *SCR* proteins and polypeptides that agree not functionally equivalent to
30 the gene products described in Section 5.1. Such a mutant *SCR* protein or polypeptide may contain one or more deletions, additions or substitutions of *SCR* amino acid residues within the amino acid sequence encoded by any one the *SCR* gene sequences described above in Section 5.1., and which result
35 in loss of one or more functions of the *SCR* protein (e.g., recognition of a specific nucleic sequence, binding of an transcription factor, etc.), thus producing a *SCR* gene

product not functionally equivalent to the wild-type SCR protein.

While random mutations can be made to SCR DNA (using random mutagenesis techniques well known to those skilled in the art) and the resulting mutant SCRs tested for activity, site-directed mutations of the SCR gene and/or coding sequence can be engineered (using site-directed mutagenesis techniques well known to those skilled in the art) to generate mutant SCRs with increased function, (e.g., resulting in improved root formation), or decreased function (e.g., resulting in suboptimal root function). In particular, mutated SCR proteins in which any of the domains shown in FIGS. 13A-F are deleted or mutated are within the scope of the invention. Additionally, peptides corresponding to one or more domains of the SCR (e.g., shown in FIGS. 13A-F), truncated or deleted SCRs, as well as fusion proteins in which the full length SCR, a SCR polypeptide or peptide fused to an unrelated protein are also within the scope of the invention and can be designed on the basis of the SCR nucleotide and SCR amino acid sequences disclosed in Section 5.1. above.

While the SCR polypeptides and peptides can be chemically synthesized (e.g., see Creighton, 1983, *Proteins: Structures and Molecular Principles*, W.H. Freeman & Co., N.Y.) large polypeptides derived from SCR and the full length SCR may advantageously be produced by recombinant DNA technology using techniques well known to those skilled in the art for expressing nucleic acid sequences.

Methods which are well known to those skilled in the art can be used to construct expression vectors containing SCR protein coding sequences and appropriate transcriptional/translational control signals. These methods include, for example, *in vitro* recombinant DNA techniques, synthetic techniques and *in vivo* recombination/genetic recombination. See, for example, the techniques described in Sambrook et al., 1989, *supra*, and Ausubel et al., 1989, *supra*. Alternatively, RNA capable of encoding SCR protein

sequences may be chemically synthesized using, for example, synthesizers. See, for example, the techniques described in "Oligonucleotide Synthesis", 1984, Gait, M.J. ed., IRL Press, Oxford.

5 A variety of host-expression vector systems may be utilized to express the *SCR* gene products of the invention. Such host-expression systems represent vehicles by which the *SCR* gene products of interest may be produced and subsequently recovered and/or purified from the culture or
10 plant (using purification methods well known to those skilled in the art), but also represent cells which may, when transformed or transfected with the appropriate nucleotide coding sequences, exhibit the *SCR* protein of the invention *in situ*. These include but are not limited to microorganisms
15 such as bacteria (e.g., *E. coli*, *B. subtilis*) transformed with recombinant bacteriophage DNA, plasmid DNA or cosmid DNA expression vectors containing *SCR* protein coding sequences; yeast (e.g., *Saccharomyces*, *Pichia*) transformed with recombinant yeast expression vectors containing the *SCR*
20 protein coding sequences; insect cell systems infected with recombinant virus expression vectors (e.g., baculovirus) containing the *SCR* protein coding sequences; plant cell systems infected with recombinant virus expression vectors (e.g., cauliflower mosaic virus, CaMV; tobacco mosaic virus,
25 TMV) or transformed with recombinant plasmid expression vectors (e.g., Ti plasmid) containing *SCR* protein coding sequences; or mammalian cell systems (e.g., COS, CHO, BHK, 293, 3T3) harboring recombinant expression constructs containing promoters derived from the genome of mammalian
30 cells (e.g., metallothionein promoter) or from mammalian viruses (e.g., the adenovirus late promoter; the vaccinia virus 7.5K promoter; the cytomegalovirus promoter/enhancer; etc.).

In bacterial systems, a number of expression
35 vectors may be advantageously selected depending upon the use intended for the *SCR* protein being expressed. For example, when a large quantity of such a protein is to be produced,

for the generation of antibodies or to screen peptide libraries, for example, vectors which direct the expression of high levels of fusion protein products that are readily purified may be desirable. Such vectors include, but are not
5 limited, to the *E. coli* expression vector pUR278 (Ruther et al., 1983, EMBO J. 2:1791), in which the SCR coding sequence may be ligated individually into the vector in frame with the lac Z coding region so that a fusion protein is produced; pIN vectors (Inouye & Inouye, 1985, Nucleic Acids Res. 13:3101-
10 3109; Van Heeke & Schuster, 1989, J. Biol. Chem. 264:5503-5509); and the like. pGEX vectors may also be used to express foreign polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified from lysed cells by
15 adsorption to glutathione-agarose beads followed by elution in the presence of free glutathione. The pGEX vectors are designed to include thrombin or factor Xa protease cleavage sites so that the cloned target gene protein can be released from the GST moiety.

20 In one such embodiment of a bacterial system, full length cDNA sequences are appended with in-frame Bam HI sites at the amino terminus and Eco RI sites at the carboxyl terminus using standard PCR methodologies (Innis et al., 1990, *supra*) and ligated into the pGEX-2TK vector (Pharmacia,
25 Uppsala, Sweden). The resulting cDNA construct contains a kinase recognition site at the amino terminus for radioactive labelling and glutathione S-transferase sequences at the carboxyl terminus for affinity purification (Nilsson, et al., 1985, EMBO J. 4: 1075; Zabeau and Stanley, 1982, EMBO J. 1:
30 1217.

The recombinant constructs of the present invention may include a selectable marker for propagation of the construct. For example, a construct to be propagated in bacteria preferably contains an antibiotic resistance gene,
35 such as one that confers resistance to kanamycin, tetracycline, streptomycin, or chloramphenicol. Suitable

vectors for propagating the construct include plasmids, cosmids, bacteriophages or viruses, to name but a few.

In addition, the recombinant constructs may include plant-expressible, selectable, or screenable marker genes for isolating, identifying or tracking plant cells transformed by these constructs. Selectable markers include, but are not limited to, genes that confer antibiotic resistance, (e.g., resistance to kanamycin or hygromycin) or herbicide resistance (e.g., resistance to sulfonylurea, phosphinothricin, or glyphosate). Screenable markers include, but are not be limited to, genes encoding β -glucuronidase (Jefferson, 1987, Plant Mol. Biol. Rep. 5:387-405), luciferase (Ow et al., 1986, Science 234:856-859), B protein that regulates anthocyanin pigment production (Goff et al., 1990, EMBO J 9:2517-2522).

In embodiments of the present invention which utilize the *Agrobacterium tumefaciens* system for transforming plants (see *infra*), the recombinant constructs may additionally comprise at least the right T-DNA border sequences flanking the DNA sequences to be transformed into the plant cell. Alternatively, the recombinant constructs may comprise the right and left T-DNA border sequences flanking the DNA sequence. The proper design and construction of such T-DNA based transformation vectors are well known to those skilled in the art.

5.1.3. ANTIBODIES TO SCR PROTEINS AND POLYPEPTIDES

Antibodies that specifically recognize one or more epitopes of SCR, or epitopes of conserved variants of SCR, or peptide fragments of the SCR are also encompassed by the invention. Such antibodies include but are not limited to polyclonal antibodies, monoclonal antibodies (mAbs), humanized or chimeric antibodies, single chain antibodies, Fab fragments, F(ab'), fragments, fragments produced by a Fab expression library, anti-idiotypic (anti-Id) antibodies, and epitope-binding fragments of any of the above.

For the production of antibodies, various host animals may be immunized by injection with the SCR protein, an SCR peptide (e.g., one corresponding to a functional domain of the protein), a truncated SCR polypeptide (SCR in which one or more domains has been deleted), functional equivalents of the SCR protein, or mutants of the SCR protein. Such SCR proteins, polypeptides, peptides or fusion proteins can be prepared and obtained as described in Section 5.1.2. *supra*. Host animals may include but are not limited to rabbits, mice, and rats, to name but a few. Various adjuvants may be used to increase the immunological response, depending on the host species, including but not limited to Freund's (complete and incomplete), mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanin, dinitrophenol, and potentially useful human adjuvants such as BCG (bacille Calmette-Guerin) and *Corynebacterium parvum*. Polyclonal antibodies are heterogeneous populations of antibody molecules derived from the sera of the immunized animals.

Monoclonal antibodies, which are homogeneous populations of antibodies to a particular antigen, may be obtained by any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique of Kohler and Milstein, (Nature 256:495-497 [1975]; and U.S. Patent No. 4,376,110), the human B-cell hybridoma technique (Kosbor et al., 1983, Immunology Today 4:72; Cole et al., 1983, Proc. Natl. Acad. Sci. USA 80:2026-2030), and the EBV-hybridoma technique (Cole et al., 1985, Monoclonal Antibodies And Cancer Therapy, Alan R. Liss, Inc., pp. 77-96). Such antibodies may be of any immunoglobulin class including IgG, IgM, IgE, IgA, IgD and any subclass thereof. The hybridoma producing the mAb of this invention may be cultivated *in vitro* or *in vivo*. Production of high titers of mAbs *in vivo* makes this the presently preferred method of production.

In addition, techniques developed for the production of "chimeric antibodies" (Morrison et al., 1984, Proc. Natl. Acad. Sci., 81:6851-6855; Neuberger et al., 1984, Nature, 312:604-608; Takeda et al., 1985, Nature, 314:452-5 454) by splicing the genes from a mouse antibody molecule of appropriate antigen specificity together with genes from a human antibody molecule of appropriate biological activity can be used. A chimeric antibody is a molecule in which different portions are derived from different animal species, 10 such as those having a variable region derived from a murine mAb and a human immunoglobulin constant region.

In addition, techniques have been developed for the production of humanized antibodies. (See, e.g., Queen, U.S. Patent No. 5,585,089.) An immunoglobulin light or heavy 15 chain variable region consists of a "framework" region interrupted by three hypervariable regions, referred to as complementarily determining regions (CDRs). The extent of the framework region and CDRs have been precisely defined (see, "Sequences of Proteins of Immunological Interest", 20 Kabat, E. et al., U.S. Department of Health and Human Services (1983). Briefly, humanized antibodies are antibody molecules from non-human species having one or more CDRs from the non-human species and a framework region from a human immunoglobulin molecule.

25 Alternatively, techniques described for the production of single chain antibodies (U.S. Patent 4,946,778; Bird, 1988, Science 242:423-426; Huston et al., 1988, Proc. Natl. Acad. Sci. USA 85:5879-5883; and Ward et al., 1989, Nature 334:544-546) can be adapted to produce single chain 30 antibodies against SCR proteins or polypeptides. Single chain antibodies are formed by linking the heavy and light chain fragments of the Fv region via an amino acid bridge, resulting in a single chain polypeptide.

Antibody fragments which recognize specific 35 epitopes may be generated by known techniques. For example, such fragments include but are not limited to: the F(ab')₂ fragments which can be produced by pepsin digestion of the

antibody molecule and the Fab fragments which can be generated by reducing the disulfide bridges of the F(ab')₂ fragments. Alternatively, Fab expression libraries may be constructed (Huse et al., 1989, Science, 246:1275-1281) to
5 allow rapid and easy identification of monoclonal Fab fragments with the desired specificity.

Antibodies to a SCR protein and/or polypeptide can, in turn, be utilized to generate anti-idiotypic antibodies that "mimic" SCR, using techniques well known to those
10 skilled in the art. (See, e.g., Greenspan & Bona, 1993, FASEB J 7(5):437-444; and Nissinoff, 1991, J. Immunol. 147(8):2429-2438).

15 5.1.4. SCR GENE OR GENE PRODUCTS AS MARKERS FOR QUALITATIVE TRAIT LOCI

Any of the nucleotide sequences (including EST clone sequences) described in §§ 5.1 and 5.1.1. and/or listed in Table 1, and/or polypeptides and proteins described in §§ 5.1.2. and/or listed in Table 1, can be used as markers
20 for qualitative trait loci in breeding programs for crop plants. To this end, the nucleic acid molecules, including but not limited to full length SCR coding sequences, and/or partial sequences (ESTs), can be used in hybridization and/or DNA amplification assays to identify the endogenous SCR
25 genes, scr mutant alleles and/or SCR expression products in cultivars as compared to wild-type plants. They can also be used as markers for linkage analysis of qualitative trait loci. It is also possible that the SCR gene may encode a product responsible for a qualitative trait that is desirable
30 in a crop breeding program. Alternatively, the SCR protein, peptides and/or antibodies can be used as reagents in immunoassays to detect expression of the SCR gene in cultivars and wild-type plants.

5.2. SCR PROMOTERS

According to the present invention, *SCR* promoters and functional portions thereof described herein refer to regions of the *SCR* gene which are capable of promoting tissue-specific expression in embryos and/or roots of an operably linked coding sequence in plants. The *SCR* promoter described herein refers to the regulatory elements of *SCR* genes, i.e., regulatory regions of genes which are capable of selectively hybridizing to the nucleic acids described in Section 5.1, or regulatory sequences contained, for example, in the region between the translational start site of the Arabidopsis *SCR* gene and the HindIII site approximately 2.5 kb upstream of the site in plasmid pLIG1-3/SAC+Mob21SAC (see FIGS. 5A and 14) in hybridization assays, or which are homologous by sequence analysis (containing a span of 10 or more nucleotides in which at least 50 percent of the nucleotides are identical to the sequences presented herein). Homologous nucleotide sequences refer to nucleotide sequences including, but not limited to, *SCR* promoters in diverse plant species (e.g., promoters of orthologs of Arabidopsis *SCR*) as well as genetically engineered derivatives of the promoters described herein.

Methods which could be used for the synthesis, isolation, molecular cloning, characterization and manipulation of *SCR* promoter sequences are well known to those skilled in the art. See, e.g., the techniques described in Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2nd. ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, New York (1989).

According to the present invention, *SCR* promoter sequences or portions thereof described herein may be obtained from appropriate plant or mammalian sources from cell lines or recombinant DNA constructs containing *SCR* promoter sequences, and/or by chemical synthetic methods. *SCR* promoter sequences can be obtained from genomic clones containing sequences 5' upstream of *SCR* coding sequences. Such 5' upstream clones may be obtained by screening genomic

libraries using SCR protein coding sequences, particularly those encoding SCR N-terminal sequences, from SCR gene clones obtained as described in Sections 5.1. and 5.2. Standard methods that may be used in such screening include, for example, the method set forth in Benton & Davis, 1977, Science 196:180 for bacteriophage libraries; and Grunstein & Hogness, 1975, Proc. Nat. Acad. Sci. U.S.A. 72:3961-3965 for plasmid libraries.

The full extent and location of SCR promoters within such 5' upstream clones may be determined by the functional assay described below. In the event a 5' upstream clone does not contain the entire SCR promoter as determined by the functional assay, the insert DNA of the clone may be used to isolate genomic clones containing sequences further 5' upstream of the SCR coding sequences. Such further upstream sequences can be spliced on to existing 5' upstream sequences and the reconstructed 5' upstream region tested for functionality as a SCR promoter (i.e., promoting tissue-specific expression in embryos and/or roots of an operably linked gene in plants). This process may be repeated until the complete SCR promoter is obtained.

The location of the SCR promoter within genomic sequences 5' upstream of the SCR gene isolated as described above may be determined using any method known in the art. For example, the 3'-end of the promoter may be identified by locating the transcription initiation site, which may be determined by methods such as RNase protection (e.g., Liang et al., 1989, J. Biol. Chem. 264:14486-14498), primer extension (e.g., Weissenborn & Larson, 1992, J. Biol. Chem. 267:6122-6131), and/or reverse transcriptase/PCR. The location of the 3'-end of the promoter may be confirmed by sequencing and computer analysis, examining for the canonical AGGA or TATA boxes of promoters that are typically 50-60 base pairs (bp) and 25-35 bp 5'-upstream of the transcription initiation site. The 5'-end promoter may be defined by deleting sequences from the 5'-end of the promoter containing fragment, constructing a transcriptional or translational

fusion of the resected fragment and a reporter gene, and examining the expression characteristics of the chimeric gene in transgenic plants. Reporter genes that may be used to such ends include, but are not limited to, GUS, CAT, luciferase, β -galactosidase and C1 and R gene controlling anthocyanin production.

According to the present invention, a SCR promoter is one that confers to an operably linked gene in a transgenic plant tissue-specific expression in roots, root nodules, stems and/or embryos. A SCR promoter comprises the region between about -5,000 bp and +1 bp upstream of the transcription initiation site of SCR gene. In a particular embodiment, the Arabidopsis SCR promoter comprises the region between positions -2.5 kb and +1 in the 5' upstream region of the Arabidopsis SCR gene (see FIGS. 5A and 14).

5.2.1. CIS-REGULATORY ELEMENTS OF SCR PROMOTERS

According to the present invention, the cis-regulatory elements within a SCR promoter may be identified using any method known in the art. For example, the location of cis-regulatory elements within an inducible promoter may be identified using methods such as DNase or chemical footprinting (e.g., Meier et al., 1991, Plant Cell 3:309-315) or gel retardation (e.g., Weissenborn & Larson, 1992, J. Biol. Chem. 267-6122-6131; Beato, 1989, Cell 56:335-344; Johnson et al., 1989, Ann. Rev. Biochem. 58:799-839). Additionally, resectioning experiments may also be employed to define the location of the cis-regulatory elements. For example, an inducible promoter-containing fragment may be resected from either the 5' or 3'-end using restriction enzyme or exonuclease digests.

To determine the location of cis-regulatory elements within the sequence containing the inducible promoter, the 5'- or 3'-resected fragments, internal fragments to the inducible promoter containing sequence, or inducible promoter fragments containing sequences identified by footprinting or gel retardation experiments may be fused

to the 5'-end of a truncated plant promoter, and the activity of the chimeric promoter in transgenic plant examined. Useful truncated promoters to these ends comprise sequences starting at or about the transcription initiation site and 5 extending to no more than 150 bp 5' upstream. These truncated promoters generally are inactive or are only minimally active. Examples of such truncated plant promoters may include, among others, a "minimal" CaMV 35S promoter whose 5' end terminates at position -46 bp with respect to 10 the transcription initiation site (Skriver et al., Proc. Natl. Acad. Sci. USA 88:7266-7270); the truncated "-90 35S" promoter in the X-GUS-90 vector (Benfey & Chua, 1989, Science 244:174-181); a truncated "-101 nos" promoter derived from the nopaline synthase promoter (Aryan et al., 1991, Mol. Gen. 15 Genet. 225:65-71); and the truncated maize Adh-1 promoter in pADcat 2 (Ellis et al., 1987, EMBO J. 6:11-16).

According to the present invention, a cis-regulatory element of a SCR promoter is a sequence that confers to a truncated promoter tissue-specific expression in 20 embryos, stems, root nodules and/or roots.

5.2.2. SCR PROMOTER-DRIVEN EXPRESSION VECTORS

The properties of the nucleic acid sequences are varied as are the genetic structures of various potential 25 host plant cells. In the preferred embodiments of the present invention, described herein, a number of features which an artisan may recognize as not being absolutely essential, but clearly advantageous are used. These include methods of isolation, synthesis or construction of gene 30 constructs, the manipulation of the gene constructs to be introduced into plant cells, certain features of the gene constructs, and certain features of the vectors associated with the gene constructs.

Further, the gene constructs of the present 35 invention may be encoded on DNA or RNA molecules. According to the present invention, it is preferred that the desired, stable genotypic change of the target plant be effected

through genomic integration of exogenously introduced nucleic acid construct(s), particularly recombinant DNA constructs. Nonetheless, according to the present invention, such genotypic changes can also be effected by the introduction of 5 episomes (DNA or RNA) that can replicate autonomously and that are somatically and germinally stable. Where the introduced nucleic acid constructs comprise RNA, plant transformation or gene expression from such constructs may proceed through a DNA intermediate produced by reverse 10 transcription.

The present invention provides for use of recombinant DNA constructs which contain tissue-specific and developmental-specific promoter fragments and functional portions thereof. As used herein, a functional portion of a 15 *SCR* promoter is capable of functioning as a tissue-specific promoter in the embryo, stem, root nodule and/or root of a plant. The functionality of such sequences can be readily established by any method known in the art. Such methods include, for example, constructing expression vectors with 20 such sequences and determining whether they confer tissue-specific expression in the embryo, stem, root nodule and/or root to an operably linked gene. In a particular embodiment, the invention provides for the use of the *Arabidopsis SCR* promoter contained in the sequences depicted in FIGS. 5A and 25 14 and the insert DNA of plasmid pGEX-2TK⁺.

The *SCR* promoters of the invention may be used to direct the expression of any desired protein, or to direct the expression of a RNA product, including, but not limited to, an "antisense" RNA or ribozyme. Such recombinant 30 constructs generally comprise a native *SCR* promoter or a recombinant *SCR* promoter derived therefrom, ligated to the nucleic acid sequence encoding a desired heterologous gene product.

A recombinant *SCR* promoter is used herein to refer 35 to a promoter that comprises a functional portion of a native *SCR* promoter or a promoter that contains native promoter sequences that is modified by a regulatory element from a *SCR*

promoter. Alternatively, a recombinant inducible promoter derived from the *scr* promoter may be a chimeric promoter, comprising a full-length or truncated plant promoter modified by the attachment of one or more *SCR* cis-regulatory elements.

5 The manner of chimeric promoter constructions may be any well known in the art. For examples of approaches that can be used in such constructions, see Section 5.1.2., above and Fluhr et al., 1986, *Science* 232:1106-1112; Ellis et al., 1987, *EMBO J.* 6:11-16; Strittmatter & Chua, 1987, *Proc.*
10 *Natl. Acad. Sci. USA* 84:8986-8990; Poulsen & Chua, 1988, *Mol. Gen. Genet.* 214:16-23; Comai et al., 1991, *Plant Mol. Biol.* 15:373-381; Aryan et al., 1991, *Mol. Gen. Genet.* 225:65-71.

 According to the present invention, where a *SCR* promoter or a recombinant *SCR* promoter is used to express a
15 desired protein, the DNA construct is designed so that the protein coding sequence is ligated in phase with the translational initiation codon downstream of the promoter. Where the promoter fragment is missing 5' leader sequences, a DNA fragment encoding both the protein and its 5' RNA leader
20 sequence is ligated immediately downstream of the transcription initiation site. Alternatively, an unrelated 5' RNA leader sequence may be used to bridge the promoter and the protein coding sequence. In such instances, the design should be such that the protein coding sequence is ligated in
25 phase with the initiation codon present in the leader sequence, or ligated such that no initiation codon is interposed between the transcription initiation site and the first methionine codon of the protein.

 Further, it may be desirable to include additional
30 DNA sequences in the protein expression constructs. Examples of additional DNA sequences include, but are not limited to, those encoding: a 3' untranslated region; a transcription termination and polyadenylation signal; an intron; a signal peptide (which facilitates the secretion of the protein); or
35 a transit peptide (which targets the protein to a particular cellular compartment such as the nucleus, chloroplast, mitochondria, or vacuole).

5.3. PRODUCTION OF TRANSGENIC PLANTS AND PLANT CELLS

According to the present invention, a desirable plant or plant cell may be obtained by transforming a plant cell with the nucleic acid constructs described herein. In some instances, it may be desirable to engineer a plant or plant cell with several different gene constructs. Such engineering may be accomplished by transforming a plant or plant cell with all of the desired gene constructs simultaneously. Alternatively, the engineering may be carried out sequentially. That is, transforming with one gene construct, obtaining the desired transformant after selection and screening, transforming the transformant with a second gene construct, and so on.

In an embodiment of the present invention, *Agrobacterium* is employed to introduce the gene constructs into plants. Such transformations preferably use binary *Agrobacterium* T-DNA vectors (Bevan, 1984, Nuc. Acid Res. 12:8711-8721), and the co-cultivation procedure (Horsch et al., 1985, Science 227:1229-1231). Generally, the *Agrobacterium* transformation system is used to engineer dicotyledonous plants (Bevan et al., 1982, Ann. Rev. Genet. 16:357-384; Rogers et al., 1986, Methods Enzymol. 118:627-641). The *Agrobacterium* transformation system may also be used to transform, as well as transfer, DNA to monocotyledonous plants and plant cells (see Hernalsteen et al., 1984, EMBO J 3:3039-3041; Hooykass-Van Slogteren et al., 1984, Nature 311:763-764; Grimsley et al., 1987, Nature 325:1677-179; Boulton et al., 1989, Plant Mol. Biol. 12:31-40.; Gould et al., 1991, Plant Physiol. 95:426-434).

In other embodiments, various alternative methods for introducing recombinant nucleic acid constructs into plants and plant cells may also be utilized. These other methods are particularly useful where the target is a monocotyledonous plant or plant cell. Alternative gene transfer and transformation methods include, but are not limited to, protoplast transformation through calcium-,

polyethylene glycol (PEG)- or electroporation-mediated uptake of naked DNA (see Paszkowski et al., 1984, EMBO J 3:2717-2722, Potrykus et al., 1985, Mol. Gen. Genet. 199:169-177; Fromm et al., 1985, Proc. Natl. Acad. Sci. USA 82:5824-5828; 5 Shimamoto, 1989, Nature 338:274-276), and electroporation of plant tissues (D'Halluin et al., 1992, Plant Cell 4:1495-1505). Additional methods for plant cell transformation include microinjection, silicon carbide mediated DNA uptake (Kaeppeler et al., 1990, Plant Cell Reporter 9:415-418), and 10 microprojectile bombardment (see Klein et al., 1988, Proc. Natl. Acad. Sci. USA 85:4305-4309; Gordon-Kamm et al., 1990, Plant Cell 2:603-618).

According to the present invention, a wide variety of plants may be engineered for the desired physiological and 15 agronomic characteristics described herein using the nucleic acid constructs of the instant invention and the various transformation methods mentioned above. In preferred embodiments, target plants for engineering include, but are not limited to, crop plants such as maize, wheat, rice, 20 soybean, tomato, tobacco, carrots, peanut, potato, sugar beets, sunflower, yam, Arabidopsis, rape seed, and petunia; and trees such as spruce.

According to the present invention, desired plants and plant cells may be obtained by engineering the gene 25 constructs described herein into a variety of plant cell types, including but not limited to, protoplasts, tissue culture cells, tissue and organ explants, pollen, embryos as well as whole plants. In an embodiment of the present invention, the engineered plant material is selected or 30 screened for transformants (i.e., those that have incorporated or integrated the introduced gene construct(s)) following the approaches and methods described below. An isolated transformant may then be regenerated into a plant. Alternatively, the engineered plant material may be 35 regenerated into a plant, or plantlet, before subjecting the derived plant, or plantlet, to selection or screening for the marker gene traits. Procedures for regenerating plants from

plant cells, tissues or organs, either before or after selecting or screening for marker gene(s), are well known to those skilled in the art.

A transformed plant cell, callus, tissue or plant
5 may be identified and isolated by selecting or screening the engineered plant material for traits encoded by the marker genes present on the transforming DNA. For instance, selection may be performed by growing the engineered plant material on media containing inhibitory amounts of the
10 antibiotic or herbicide to which the transforming marker gene construct confers resistance. Further, transformed plants and plant cells may also be identified by screening for the activities of any visible marker genes (e.g., the β -glucuronidase, luciferase, B or C1 genes) that may be present
15 on the recombinant nucleic acid constructs of the present invention. Such selection and screening methodologies are well known to those skilled in the art.

Physical and biochemical methods may also be used to identify a plant or plant cell transformant containing the
20 gene constructs of the present invention. These methods include but are not limited to: 1) Southern analysis or PCR amplification for detecting and determining the structure of the recombinant DNA insert; 2) Northern blot, S-1 RNase protection, primer-extension or reverse transcriptase-PCR
25 amplification for detecting and examining RNA transcripts of the gene constructs; 3) enzymatic assays for detecting enzyme or ribozyme activity, where such gene products are encoded by the gene construct; 4) protein gel electrophoresis, western blot techniques, immunoprecipitation, or enzyme-linked
30 immunoassays, where the gene construct products are proteins; 5) biochemical measurements of compounds produced as a consequence of the expression of the introduced gene constructs. Additional techniques, such as *in situ* hybridization, enzyme staining, and immunostaining, may also
35 be used to detect the presence or expression of the recombinant construct in specific plant organs and tissues.

The methods for doing all these assays are well known to those skilled in the art.

5.3.1. TRANSGENIC PLANTS THAT ECTOPICALLY EXPRESS SCR

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In accordance to the present invention, a plant that expresses a recombinant SCR gene may be engineered by transforming a plant cell with a gene construct comprising a plant promoter operably associated with a sequence encoding SCR protein or a fragment thereof. (Operably associated is used herein to mean that transcription controlled by the "associated" promoter would produce a functional messenger RNA, whose translation would produce the enzyme.) The plant promoter may be constitutive or inducible. Useful constitutive promoters include, but are not limited to, the CaMV 35S promoter, the T-DNA mannopine synthetase promoter, and their various derivatives. Useful inducible promoters include but are not limited to the promoters of ribulose biphosphate carboxylase (RUBISCO) genes, chlorophyll a/b binding protein (CAB) genes, heat shock genes, the defense responsive gene (e.g., phenylalanine ammonia lyase genes), wound induced genes (e.g., hydroxyproline rich cell wall protein genes), chemically-inducible genes (e.g., nitrate reductase genes, gluconase genes, chitinase genes, PR-1 genes etc.), dark-inducible genes (e.g., asparagine synthetase gene (Coruzzi and Tsai, U.S. Patent 5,256,558, October 26, 1993, Gene Encoding Plant Asparagine Synthetase) developmentally regulated genes (e.g., Shoot Meristemless gene) to name just a few.

In yet another embodiment of the present invention, it may be advantageous to transform a plant with a gene construct operably linking a modified or artificial promoter to a sequence encoding SCR protein or a fragment thereof. Typically, such promoters, constructed by recombining structural elements of different promoters, have unique expression patterns and/or levels not found in natural promoters. See, e.g., Salina et al., 1992, Plant Cell

4:1485-1493, for examples of artificial promoters constructed from combining cis-regulatory elements with a promoter core.

In a preferred embodiment of the present invention, the associated promoter is a strong and root, root nodule, 5 stem and/or embryo-specific plant promoter such that the SCR protein is overexpressed in the transgenic plant. Examples of root- and root nodules-specific promoters include but are not limited to the promoters of *SCR* genes, *SHR* genes, leghemoglobin genes, nodulin genes and root-specific 10 glutamine synthetase genes (See e.g., Tingey et al., 1987, EMBO J. 6:1-9; Edwards et al., 1990, Proc. Nat. Acad. Sci. USA 87:3459-3463).

In yet another preferred embodiment of the present invention, the overexpression of SCR protein in roots may be 15 engineered by increasing the copy number of the SCR gene. One approach to producing such transgenic plants is to transform with nucleic acid constructs that contain multiple copies of the complete SCR gene (i.e., with its own native scr promoter). Another approach is repeatedly transform 20 successive generations of a plant line with one or more copies of the complete SCR gene. Yet another approach is to place a complete SCR gene in a nucleic acid construct containing an amplification-selectable marker (ASM) gene such as the glutamine synthetase or dihydrofolate reductase gene. 25 Cells transformed with such constructs is subjected to culturing regimes that select cell lines with increased copies of complete SCR gene. See, e.g., Donn et al., 1984, J. Mol. Appl. Genet. 2:549-562, for a selection protocol used to isolate of a plant cell line containing amplified copies 30 of the GS gene. Because the desired gene is closely linked to the ASM, cell lines that amplified the ASM gene are also likely to have amplified the SCR gene. Cell lines with amplified copies of the SCR gene can then be regenerated into transgenic plants.

35

5.3.2. TRANSGENIC PLANTS THAT SUPPRESS ENDOGENOUS SCR EXPRESSION

In accordance with the present invention, a desired plant may be engineered by suppressing *SCR* activity. In one embodiment, the suppression may be engineered by transforming a plant with a gene construct encoding an antisense RNA or ribozyme complementary to a segment or the whole of *SCR* RNA transcript, including the mature target mRNA. In another embodiment, *SCR* gene suppression may be engineered by transforming a plant cell with a gene construct encoding a ribozyme that cleaves the *SCR* mRNA transcript.

Alternatively, the plant can be engineered, e.g., via targeted homologous recombination to inactive or "knock-out" expression of the plant's endogenous *SCR*.

For all of the aforementioned suppression constructs, it is preferred that such gene constructs express specifically in the root, root nodule, stem and/or embryo tissues. Alternatively, it may be preferred to have the suppression constructs expressed constitutively. Thus, constitutive promoters, such as the nopaline, CaMV 35S promoter, may also be used to express the suppression constructs. A most preferred promoter for these suppression constructs is a *SCR* or *SHR* promoter.

In accordance with the present invention, desired plants with suppressed target gene expression may also be engineered by transforming a plant cell with a co-suppression construct. A co-suppression construct comprises a functional promoter operatively associated with a complete or partial *SCR* gene sequence. It is preferred that the operatively associated promoter be a strong, constitutive promoter, such as the CaMV 35S promoter. Alternatively, the co-suppression construct promoter can be one that expresses with the same tissue and developmental specificity as the *scr* gene.

According to the present invention, it is preferred that the co-suppression construct encodes a incomplete *SCR* mRNA, although a construct encoding a fully functional *SCR*

mRNA or enzyme may also be useful in effecting co-suppression.

In accordance with the present invention, desired plants with suppressed target gene expression may also be engineered by transforming a plant cell with a construct that can effect site-directed mutagenesis of the SCR gene. (See, e.g., Offringa et al., 1990, EMBO J. 9:3077-84; and Kanevskii et al., 1990, Dokl. Akad. Nauk. SSSR 312:1505-1507) for discussions of nucleic constructs for effecting site-directed mutagenesis of target genes in plants.) It is preferred that such constructs effect suppression of SCR gene by replacing the endogenous SCR gene sequence through homologous recombination with none or inactive SCR protein coding sequence.

15

5.3.3. TRANSGENIC PLANTS THAT EXPRESS A TRANSGENE CONTROLLED BY THE SCR PROMOTER

In accordance with the present invention, a desired plant may be engineered to express a gene of interest under the control of the SCR promoter. SCR promoters and functional portions thereof refer to regions of the nucleic acid sequence which are capable of promoting tissue-specific transcription of an operably linked gene of interest in the embryo, stem, root nodule and/or root of a plant. The SCR promoter described herein refers to the regulatory elements of SCR genes as described in Section 5.2.

Genes that may be beneficially expressed in the roots and/or root nodules of plants include genes involved in nitrogen fixation or cytokines or auxins, or genes which regulate growth, or growth of roots. In addition, genes encoding proteins that confer on plants herbicide, salt, or pest resistance may be engineered for root specific expression. The nutritional value of root crops may also be enhanced through SCR promoter driven expression of nutritional proteins. Alternatively, therapeutically useful proteins may be expressed specifically in root crops.

Genes that may be beneficially expressed in the stems of plants include those involved in starch lignin or cellulose biosynthesis.

In accordance with the present invention, desired plants which express a heterologous gene of interest under the control of the *SCR* promoter may be engineered by transforming a plant cell with *SCR* promoter driven constructs using those techniques described in Section 5.2.2. and 5.3., *supra*.

10

5.3.4. SCREENING OF TRANSFORMED PLANTS FOR THOSE HAVING DESIRED ALTERED TRAITS

It will be recognized by those skilled in the art that in order to obtain transgenic plants having the desired engineered traits, screening of transformed plants (*i.e.*, those having an gene construct of the invention) having those traits may be required. For example, where the plants have been engineered for ectopic overexpression of *SCR* gene, transformed plants are examined for those expressing the *SCR* gene at the desired level and in the desired tissues and developmental stages. Where the plants have been engineered for suppression of the *SCR* gene product, transformed plants are examined for those expressing the *SCR* gene product (*e.g.*, RNA or protein) at reduced levels in various tissues. The plants exhibiting the desired physiological changes, *e.g.*, ectopic *SCR* overexpression or *SCR* suppression, may then be subsequently screened for those plants that have the desired structural changes at the plant level (*e.g.*, transgenic plants with overexpression or suppression of *SCR* gene having the desired altered root structure). The same principle applies to obtaining transgenic plants having tissue-specific expression of a heterologous gene in embryos and/or roots by the use of a *SCR* promoter driven expression construct.

Alternatively, the transformed plants may be directly screened for those exhibiting the desired structural and functional changes. In one embodiment, such screening may be for the size, length or pattern of the root of the

transformed plants. In another embodiment, the screening of the transformed plants may be for altered gravitropism or decreased susceptibility to lodging. In other embodiments, the screening of the transformed plants may be for improved agronomic characteristics (e.g., faster growth, greater vegetative or reproductive yields, or improved protein contents, etc.), as compared to unengineered progenitor plants, when cultivated under various growth conditions (e.g., soils or media containing different amount of nutrients, water content).

According to the present invention, plants engineered with *SCR* overexpression may exhibit improved vigorous growth characteristics when cultivated under conditions where large and thicker roots are advantageous. Plants engineered for *SCR* suppression may exhibit improved vigorous growth characteristics when cultivated under conditions where thinner roots are advantageous.

Engineered plants and plant lines possessing such improved agronomic characteristics may be identified by examining any of following parameters: 1) the rate of growth, measured in terms of rate of increase in fresh or dry weight; 2) vegetative yield of the mature plant, in terms of fresh or dry weight; 3) the seed or fruit yield; 4) the seed or fruit weight; 5) the total nitrogen content of the plant; 6) the total nitrogen content of the fruit or seed; 7) the free amino acid content of the plant; 8) the free amino acid content of the fruit or seed; 9) the total protein content of the plant; and 10) the total protein content of the fruit or seed. The procedures and methods for examining these parameters are well known to those skilled in the art.

According to the present invention, a desired plant is one that exhibits improvement over the control plant (i.e., progenitor plant) in one or more of the aforementioned parameters. In an embodiment, a desired plant is one that shows at least 5% increase over the control plant in at least one parameter. In a preferred embodiment, a desired plant is one that shows at least 20% increase over the control plant

in at least one parameter. Most preferred is a plant that shows at least 50% increase in at least one parameter.

6. EXAMPLE 1: ARABIDOPSIS SCR GENE

5 This example describes the cloning and structure of the Arabidopsis SCR gene and its expression. The deduced amino acid sequence of the Arabidopsis SCR gene product contains a number of potential functional domains similar to those found in transcription factors. Closely related
10 sequences have been found in both dicots and monocots indicating that Arabidopsis SCR is a member of a new protein family. The expression pattern of the SCR gene was characterized by means of *in situ* hybridization and by an enhancer trap insertion upstream of the SCR gene (described
15 in more detail in Section 7). The expression pattern is consistent with a key role for Arabidopsis SCR in regulating the asymmetric division of the cortex/endodermis initial which is essential for generating the radial organization of the root.

20

6.1. MATERIALS AND METHODS

6.1.1. PLANT CULTURE

Arabidopsis ecotypes Wassilewskija (Ws), Columbia (Col), and Landsberg erecta (Ler) were obtained from Lehle.
25 Arabidopsis seeds were surface sterilized and grown as described previously (Benfey et al., 1993, Development 119:57-70). Generation of the enhancer trap lines is described in Section 7.

30

6.1.2. GENETIC ANALYSIS

For the *scr-1* allele, co-segregation of the mutant phenotype and kanamycin resistance conferred by the inserted T-DNA was determined as described previously (Aeschbacher et al., 1995, Genes & Development 9:330-340). Because kanamycin
35 affects root growth, 1557 seeds from heterozygous lines were germinated on non-selective media, scored for the appearance of the mutant phenotype, and subsequently transferred to

selective media. All (284) phenotypically mutant seedlings showed resistance to the antibiotic, whereas 834 of 1273 phenotypically wild-type seedlings showed resistance to kanamycin, respectively. Phenotypically wild type plants (83) were also transferred to soil and allowed to set seeds. The progeny of these plants were plated on selective and non-selective media, and scored for the co-segregation of the mutant phenotype and antibiotic resistance. A majority (48) of the plants segregated for the mutant phenotype and for kanamycin resistance, whereas 35 were wild-type and sensitive to kanamycin. Due to a mis-identified cross, *scr-2* was originally thought to be non-allelic and was named *pinocchio* (Scheres et al., 1995, Development 121:53-62). Subsequent mapping results placed it in an identical chromosomal location as *scr-1*. The original *scr-2* line contained at least two T-DNA inserts. Co-segregation analysis revealed a lack of linkage between the antibiotic resistance marker carried by the T-DNA and the mutant phenotype. Antibiotic sensitive lines were identified that segregated for mutants. These lines were crossed to *scr-1*. All F1 antibiotic resistant progeny exhibited a mutant phenotype. All F2 progeny (from independent lines) were mutant, and there was a 3:1 segregation for antibiotic resistance indicating that the two mutations were allelic. Antibiotic sensitive lines of *scr-2* were found to contain a rearranged T-DNA insert as determined by Southern blots and PCR using T-DNA specific probes and primers respectively. The presence of this T-DNA in the *SCR* gene was confirmed by Southern blots using *SCR* probes. A combination of T-DNA and *SCR* specific primers was used to amplify T-DNA/*SCR* junctions. The PCR fragments were cloned using the TA cloning kit (Invitrogen) and sequenced. The insertion points were determined for both 5' and 3' T-DNA/*SCR* junctions.

6.1.3. MAPPING

Mutant plants of *scr-2* (WS background) were crossed to Col WT. DNA from mutant F2 individual plants were analyzed for co-segregation with microsatellite (Bell & Ecker, 1994, Genomics 18:137-144) and CAPS markers (Konieczny & Ausubel, 1993, Plant J. 4:403-410). The closest linkage was found to two CAPS markers located at the bottom of chromosome III. Only one out of 238 mutant chromosomes was recombinant for the BGL1 marker (Konieczny & Ausubel, 1993, Plant J. 4:403-410) and one out of 210 chromosomes was recombinant for the *cdc2b* marker.

A RFLP for the *SCR* gene was identified between Col and Ler ecotypes with Xho I endonuclease. Genomic DNAs from independent R1 lines (Jarvis et al., 1994, Plant Mol. Biol. 24:685-687) were digested with Xho I and blots were hybridized to *SCR*. Using the segregation data obtained for 25 R1 lines, the *SCR* gene was mapped relative to molecular markers by CLUSTER. The *SCR* gene was assigned to the bottom of chromosome III closest to BGL1.

20

6.1.4. PHENOTYPIC ANALYSIS

Morphological characterization of the mutant roots was performed as follows: 7 to 14 days post-germination phenotypically mutant seedlings were fixed in 4.0% formaldehyde in PIPES buffer pH 7.2. After fixation the samples were dehydrated in ethanol followed by infiltration with Histoiresin (Jung-Leica, Heidelberg, Germany). Plastic sections were mounted on superfrost slides (Fisher). The sections were either stained with 0.05% toluidine blue and photographed using Kodak 160T film or used for Casparian strip detection or antibody staining.

Casparian strip detection was performed as described previously (Scheres et al., 1995, Development 121:53-62), with the following modifications. Plastic sections were used and the counterstaining was done in 0.1% aniline blue for 5 to 15 min. The sections were visualized with a Leitz fluorescent microscope with FITC filter.

Pictures were taken using a Leitz camera attached to the microscope and Kodak HC400 film. Slides were digitized with a Nikon slide scanner and manipulated in Adobe Photoshop.

For antibody staining, sections were blocked for 2 hours at room temperature in 1% BSA in PBS containing 0.1% Tween 20 (PBT). Samples were incubated with primary antibodies at 4° C in 1% BSA in PBT overnight, and then washed 3 times 5 minutes each with PBT. Samples were incubated for two hours with biotinylated secondary antibodies (Vector Laboratories) in PBT, and washed as above. Samples were incubated with Texas Red conjugated avidin D for 2 hours at room temperature, washed as before, and mounted in Citifluor. Immunofluorescence was observed with a fluorescent microscope equipped with a Rhodamine filter.

15 Staining with the CCRC antibodies was performed as described previously (Freshour et al., 1996, Plant Physiol. 110:1413-1429).

6.1.5. MOLECULAR TECHNIQUES

20 Genomic DNA preparation was performed using the Elu-Quik kit (Schleicher & Schuell) protocol. Radioactive and non-radioactive DNA probes were labeled with either random primed labeling or PCR-mediated synthesis according to the Genius kit manual (Boehringer Mannheim). *E. coli* and

25 *Agrobacterium tumefaciens* cells were transformed using a BIO-RAD gene pulser. Plasmid DNA was purified using the alkaline lysis method (Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor, New York: Cold Spring Harbor Laboratory, 1982).

30 A probe made from a rescued fragment of 1.2 kb was used to screen a wild-type genomic library made from WS plants. One genomic clone containing an insert of approximately 23 kb was isolated. A 3.0 kb Sac I fragment from the genomic clone, which hybridized to the 1.2 kb probe,

35 was subcloned and sequenced (FIG. 5A). Comparison of the nucleotide sequence between the genomic clone and the rescued plasmid revealed the site of the T-DNA insertion.

Approximately 600,000 plaques from a cDNA library, obtained from inflorescences and siliques (*Col ecotype*), and therefore enriched in embryos, were screened with the 1.2 kb probe. Four cDNA clones were isolated. The dideoxy sequencing
5 method was performed using the Sequenase kit (United States Biochemical Corp.). Sequence-specific internal primers were synthesized and used to sequence the *Sac I* genomic as well the cDNA clones. Total RNA from plant tissues was obtained using phenol/chloroform extractions as described in (Berry et
10 al., 1985, *Mol. Cell. Biol.* 5:2238-2246) with minor modifications. Northern hybridization and detection were performed according to the Genius kit manual (Boehringer Mannheim).

To identify the site of insertion of the enhancer-
15 trap T-DNA, genomic DNA from ET199 homozygous plants was amplified using primers specific for the T-DNA left border and the *SCR* gene. An approximately 2.0 kb fragment was amplified. This fragment was sequenced and the site of insertion was found to be approximately 1 kb from the ATG
20 start codon.

6.1.6. IN SITU HYBRIDIZATION

Antisense and sense *SCR* riboprobes were labeled with digoxigenin-11-UTP (Boehringer Mannheim) using T7
25 polymerase following the manufacturer's protocol. Probes contained a 1.1 kb 3' portion of the cDNA. Probe purification, hydrolysis and quantification were performed as described in the *Boehringer Mannheim Genius System user's guide*.

30 Tissue samples were fixed in 4 % formaldehyde overnight at 4°C and rinsed two times in PBS (Jackson et al., 1991, *Pl. Cell* 3:115-125). They were subsequently pre-embedded in 1 % agarose in PBS. The fixed tissue was dehydrated in ethanol, cleared in Hemo-De (Fisher Scientific,
35 Pittsburgh, PA) and embedded in ParaplastPlus (Fisher Scientific). Tissue sections (10µm thick) were mounted on SuperfrostPlus slides (Fisher Scientific). Section

pretreatment and hybridization were performed according to (Lincoln et al., 1994, Plant Cell 6:1859-1876) except that proteinase K was used at 30 mg/ml and a two hour prehybridization step was included. Probe concentration of 5 50 ng/ml/kb was used in the hybridization.

Slides were washed and the immunological detection was performed according to (Coen et al., 1990, Cell 63:1311-1322) with the following modifications. Slides were first washed 5 h in 5xSSC, 50% formamide. After RNase treatment 10 slides were rinsed three times (20 min each) in the buffer (0.5 M NaCl, 10 mM Tris-HCl pH 8.0, 5.0 mM EDTA). In the immunological detection, antibody was diluted 1:1000, levamisole (240 ng/ml) was included in the detection buffer, and after stopping the reaction in 10 mM Tris, 1 mM EDTA, 15 sections were mounted directly to Aqua-Poly/Mount (Polysciences, Warrington, PA).

6.2. RESULTS

6.2.1. CHARACTERIZATION OF THE SCR PHENOTYPE

20 The scarecrow mutant *scr-1* was isolated in a screen of T-DNA transformed Arabidopsis lines (Feldmann, K.A., 1991, Plant J. 1:71-82), as a seedling with greatly reduced root length compared to wild-type (Scheres et al., 1995, Development 121:53-62). A second mutant *scr-2* with a similar 25 phenotype was subsequently identified among T-DNA transformed lines. Analysis of co-segregation between the mutant phenotype and antibiotic resistance carried by the T-DNA indicated tight linkage for *scr-1* and no linkage for *scr-2* (see Experimental Procedures). An antibiotic sensitive line 30 of *scr-2* was isolated and crossed with *scr-1*. The F2 progeny of this cross were all mutant and segregated 3:1 for antibiotic resistance confirming allelism (see Materials & Methods). The principal phenotypic difference between the two alleles was that *scr-1* root growth was more retarded than 35 that of *scr-2*, suggesting that it is the stronger allele (FIG. 2A). For both alleles the aerial organs appeared similar to wild-type and the flowers were fertile (FIGS. 2A

and 2B). The progeny of backcrosses of *scr-1* or *scr-2* to wild-type plants segregated 3:1 for the root phenotype for both alleles, indicating that each mutation is monogenic and recessive.

5 Analysis of transverse sections through the primary root of seedlings revealed only a single cell layer between the epidermis and the pericycle (FIG. 2C) instead of the normal radial organization consisting of cortex and endodermis (FIG. 2D). This radial organization defect was
10 not limited to the primary root, but was also present in secondary roots (FIG. 2E) and in roots regenerated from calli (FIG. 2F). Occasionally defects were observed in the number of cells in the remaining cell layer (more than the invariant 8 found in wild-type). Abnormal placement or numbers of
15 epidermal cells were also observed (see FIG. 2E). These abnormalities were more frequently observed in *scr-1* than in *scr-2*. Nevertheless, organization of the mutant root closely resembles that of wild-type except for the consistent reduction in the number of cell layers. Because the
20 endodermis and cortex are normally generated by an asymmetric division of the cortex/endodermal initial, this indicates that the primary defect in *scr* is disruption of this asymmetric division.

 It has been shown that the radial organization
25 defect in *scr-1* first appears in the developing embryo at the early torpedo stage and manifests itself as a failure of the embryonic ground tissue to undergo the asymmetric division into cortex and endodermis (Scheres et al., 1995, Development 121:53-62). This defect extends the length of the embryonic
30 axis which encompasses the embryonic root and hypocotyl. Other embryonic tissues appear similar to wild-type (Scheres et al., 1995, Development 121:53-62). In seedling hypocotyls of the *scarecrow* phenotype, two cell layers instead of the normal three layers (two cortex and one endodermis) between
35 epidermis and stele were found. This would be the expected result of the lack of the division of the embryonic ground tissue. Similar results were obtained for *scr-2*. Hence,

this mutant identifies a gene involved in the asymmetric division that produces cortex and endodermis from ground tissue in the embryonic root and hypocotyl and from the cortex/endodermal initials in primary and secondary roots.

5

6.2.2. CHARACTERIZATION OF CELL IDENTITY IN SCR ROOTS

To understand the role of the Arabidopsis *SCR* gene in regulating this asymmetric division, it was necessary to determine the identity of the mutant cell layer. Tissue-specific markers were used to distinguish between several possibilities. The cell layer could have differentiated attributes of either cortex or endodermis. Alternatively, it could have an undifferentiated, initial-cell identity or it could have a chimeric identity with differentiated attributes of both endodermis and cortex in the same cell.

Transverse sections of *scr-1* and *scr-2* roots were assayed for the presence of tissue-specific markers. The casparian strip, a deposition of suberin between radial cell walls, is specific to the endodermal cells and is believed to act as a barrier to the entry of solutes into the vasculature (Esau, K. Anatomy of Seed Plants, New York: John Wiley & Sons, 1977, Ed. 2, pp. 1-550). Histochemical staining revealed the presence of a casparian strip in the mutant cell layer (FIG. 3A, compare to wild-type, FIG. 3B). It is noted that in the vascular cylinder, this histochemical stain also reveals the presence of lignin, indicating the presence of differentiated xylem cells in mutant (FIG. 3A) and wild-type (FIG. 3B). Another marker of the differentiated endodermis is the arabinogalactan epitope recognized by the monoclonal antibody, JIM13 (Knox et al., 1990, Planta 181:512-521). The mutant cell layer showed staining with this antibody (FIG. 3C, compare with wild-type, FIG. 3B). As a positive control, the JIM7 antibody that recognizes pectin epitopes in all cell walls was used (FIGS. 3E and 3F). These results indicate that the cell layer between the epidermis and the pericycle has differentiated attributes of the endodermis.

As a marker for the cortex, the CCRC-M2 monoclonal antibody was used. This antibody recognizes a cell wall oligosaccharide epitope, found only on differentiated cortex and epidermis cells. In sections from the differentiation zone of *scr-1* and *scr-2*, both cortex and epidermal cells showed staining (FIG. 4A and 4B) that was similar to that of wild-type (FIG. 4C). In *scr-1*, staining of both cell types was apparent, but staining of cortex was somewhat weaker than wild-type. The positive control used the CCRC-M1 monoclonal antibody which recognizes an oligosaccharide epitope found on all cells (FIGS. 4D-F).

With the CCRC-M2 antibody an interesting difference was observed between the staining pattern of the mutants as compared to wild-type. The appearance of this epitope correlates with differentiation in these two cell types. Normally, in sections close to the root tip there is no staining. In sections higher up in the root, atrichoblasts (epidermal cells that do not make root hairs) stain. In sections from more mature root tissue, all epidermal cells as well as cortex cells stain for this epitope. In both *scr-1* and *scr-2*, sections could be found in which all epidermal cells stained while there was little detectable staining of cortex cells. Although not precisely identical to the wild-type staining pattern, the fact that the mutant cell layer clearly stains for this cortex marker indicates that there are cortex differentiated attributes expressed in these cells.

Taken together, these results indicate that the mutant cell layer has differentiated attributes of both the endodermis and cortex. The possibility that there has been a simple deletion of a cell type, or that the resulting cell type remains in an undifferentiated initial-like stage can be ruled out. This result is consistent with a role for the *scr* gene in regulating this asymmetric division rather than a role in directing cell specification.

6.2.3. MOLECULAR CLONING OF THE SCR GENE

To further elucidate the function of the Arabidopsis *SCR* gene the inserted T-DNA sequences were used to clone the gene. Plant DNA flanking the insertion site was
5 obtained from *scr-1* by plasmid rescue and used to isolate the corresponding wild-type genomic DNA. Several cDNA clones were isolated from a library made from silique tissue. Comparison of the sequence of the longest cDNA and the corresponding genomic region revealed an open reading frame
10 (ORF) interrupted by a single small intron. (FIG. 5A). A potential TATA box and polyadenylation signal that matched the consensus sequences for plant genes were also identified (Joshi, C.P., 1987, Nucl. Acids Res. 15:6643-6653); Heidecker & Messing, 1986, Ann. Rev. Plant Physiol. 37:439-466); Mogen
15 et al., 1990, Plant Cell 2:1261-1272).

Comparison of the nucleotide sequence between the genomic clone and the rescued plasmid placed the site of the T-DNA insertion in *scr-1* at codon 470 (FIGS. 5A and 5B). For *scr-2*, although no linkage was found between the mutant
20 phenotype and antibiotic resistance, DNA blot and PCR analysis of antibiotic sensitive lines revealed the presence of T-DNA sequences that co-segregated with the mutant phenotype. The insertion position in *scr-2* was determined by cloning and sequencing the PCR products amplified from its
25 genomic DNA using a combination of T-DNA and *SCR* specific primers at both sides of the insertion (FIG. 5B). In *scr-2* the T-DNA insertion point is at codon 605 (FIG. 5A and 5B). To verify linkage between the cloned gene and the mutant phenotype, we identified the chromosomal location of both the
30 *scr* locus and the *SCR* gene. To map the *scr* locus, molecular markers were used on F2 progeny of crosses between *scr-2* (ecotype Wassilewskija, Ws) and Colombia (Col) WT. These placed the *scr* locus at the bottom of chromosome III, approximately 0.5 cM away from each of the two closest
35 markers available, *cdc2b* and *BGL1* (Konieczny and Ausubel, 1993, Plant J. 4:403-410). To map the *SCR* gene, we identified a polymorphism between Col and Landsberg (Ler)

ecotypes using the SCR probe b (FIG. 5B). Southern analysis of 25 recombinant inbred lines (Jarvis et al., 1994, Plant Mol. Biol. 24:685-687) mapped the cloned gene to the same location as the SCR locus on chromosome III.

5 The determination of the molecular defects in two independent alleles and the co-localization of the cloned gene and the mutant locus confirms that we have identified the SCR gene.

10 6.2.4. THE SCR GENE HAS MOTIFS THAT INDICATE IT IS A TRANSCRIPTION FACTOR

 The Arabidopsis SCR gene product is a 653 amino acid polypeptide that contains several domains (FIG. 5B). The amino-terminus has homopolymeric stretches of glutamine, 15 serine, threonine, and proline residues, which account for 44% of the first 267 residues. Domains rich in these residues have been shown to activate transcription and may serve such a role in SCR (Johnson et al., 1993, J. Nutr. Biochem 4:386-398). A charged region between residues 265 20 and 283 has similarity to the basic domain of the bZIP family of transcriptional regulatory proteins (FIG. 5C) (Hurst, H.C., 1994, Protein Profile 1:123-168). The basic domains from several bZIP proteins have been shown to act as nuclear localization signals (Varagona et al., 1992, Plant Cell 25 4:1213-1227), and this region in SCR may act similarly. This charged region is followed by a leucine heptad repeat (residues 291-322). A second leucine heptad repeat is found toward the carboxy-terminus (residues 436 to 473). As leucine heptad repeats have been demonstrated to mediate 30 protein-protein interactions in other proteins (Hurst, H.C., 1994, Protein Profile 1:123-168), the existence of these motifs suggests that SCR may function as a dimer or a multimer. The second leucine heptad repeat is followed by a small region rich in acidic residues, also present in a 35 number of defined transcriptional activation domains (Johnson et al., 1993, J. Nutr Biochem 4:386-398). While each of these domains has been found within proteins that do not act

as transcriptional regulators, the fact that all of them are found within the deduced SCR protein sequence indicates that SCR is a transcriptional regulatory protein.

5 6.2.5. SCR IS A MEMBER OF A NOVEL PROTEIN FAMILY

The Arabidopsis SCR protein sequence was compared with the sequences in the available databases. Eleven expressed sequence tags (ESTs), nine from Arabidopsis, one from rice and one from maize, showed significant similarity
10 to residues 394 to 435 of the SCR sequence, a region immediately amino-terminal to the second leucine heptad repeat (FIGS. 15K-L). This region is designated the VHIID domain. Subsequent analysis of these EST sequences has revealed that the sequence similarity extends beyond this
15 region; in fact, the similarity extends throughout the entire known gene products. The combination and order of the motifs found in these sequences do not show significant similarity to the general structures of other established regulatory protein families (i.e., bZIP, zinc finger, MADS-domain, and
20 homeodomain), indicating that the SCR proteins comprise a novel family.

6.2.6. SCR IS EXPRESSED IN THE CORTEX/ENDODERMAL INITIALS AND IN THE ENDODERMIS

25 RNA blot analysis revealed expression of SCR in Arabidopsis siliques, leaves and roots of wild-type plants (FIG. 6A). No hybridization was detected to RNA from *scr-1* plants (FIG. 6B, lane 2). This indicates that *scr-1* has a reduced level of RNA expression and may represent the null
30 phenotype. Hybridization to RNA species larger than the normal size were detected in *scr-2*. This indicates that abnormal SCR transcripts are made in this allele, suggesting that functional but possibly altered proteins may be produced.

35 To determine if expression was localized to any particular cell type, RNA *in situ* was hybridization performed on sections of root tissue. In mature roots, expression was

localized primarily to the endodermis (FIGS. 7A and 7B). Expression appeared to start very close to or within the cortex/endodermal initials and continue up the endodermal cell file as far as the section extended. Expression was
5 also detected in late-torpedo stage embryos in the endodermis throughout the embryonic axis (FIG. 7C). Sense strand controls showed only background hybridization (FIG. 7D).

To determine whether the localization of *SCR* RNA was regulated at the transcriptional or post-transcriptional
10 level, enhancer trap (ET) lines were prepared and examined in which the β -glucuronidase (*uid-A* or *GUS*) coding sequence with a minimal promoter was expressed in the root endodermis. (See Section 7, *infra*). Restriction fragment length polymorphisms were observed when DNA from one of these lines,
15 ET199 and wild-type were probed with *SCR*. PCR and sequence analysis confirmed that the enhancer-trap construct had inserted approximately 1 kb upstream of the *SCR* start site and in the same orientation as that of *SCR* transcription.

In mature roots, expression in ET199 whole mounts
20 showed a similar pattern to that of the *in situ* hybridizations, with the strongest staining present in endodermal cells (FIG. 7E). Transverse sections indicated that expression was primarily in endodermal cells in the elongation zone (FIG. 7F). Longitudinal sections through the
25 meristematic zone revealed that expression could be detected in the cortex/endodermal initial (FIG. 7G). Of particular interest was the restriction of expression to the endodermal daughter cell after the periclinal division (FIG. 7G). This indicated that the expression pattern observed in the *in situ*
30 analysis was not due to post-transcriptional partitioning of *SCR* RNA. Rather, it suggests that after the periclinal division of the cortex/endodermis initial only one of the two cells is able to transcribe *SCR* RNA.

6.3. DISCUSSION

6.3.1. THE SCR GENE REGULATES AN ASYMMETRIC DIVISION REQUIRED FOR ROOT RADIAL ORGANIZATION

5 The formation of the cortex and endodermal layers
in the Arabidopsis root requires two asymmetric divisions.
In the first, an anticlinal division of the cortex/endodermal
initial generates two cells with different developmental
potentials. One will continue to function as an initial,
10 while the other undergoes a periclinal division to generate
the first cells in the endodermal and cortex cell files.
This second asymmetric division is eliminated in the
scarecrow mutant, resulting in a single cell layer instead of
two. The *scr* mutation appears to have little effect on any
15 other cell divisions in the root indicating that it is
involved in regulating a single asymmetric division in this
organ. Several other mutations have been characterized that
appear to affect specific cell division pathways in
Arabidopsis. These include *knolle* (*kn*) in which formation of
20 the epidermis is impaired (Lukowitz et al., 1996, Cell 84:61-
71), *wooden leg* (*wol*) in which vascular cell division is
defective (Scheres et al., 1995, Development 121:53-62) and
fass (*fs*) in which there are supernumerary cortex and
vascular cells (Scheres et al., 1995, Development 121:53-62);
25 Torres Ruiz & Jurgens, 1994, Development 120:2967-2978).
Only in the case of *scr* and *short-root* (*shr*) mutants has it
been shown that the defect is in a specific asymmetric
division.

 Mutational analyses in several organisms have
30 revealed that the genes that regulate asymmetric divisions
can be specific to a single type of division or can affect
divisions that are not clonally related (Horvitz &
Herskowitz, 1992, Cell 68:237-255). In most cases, these
mutations result in the formation of two identical daughter
35 cells with similar developmental potentials (Horvitz &
Herskowitz, 1992, Cell 68:237-255). Both resulting cells
have the identity of one or the other of the normal daughter

cells, an example of which is the *swi* mutation in *S. cerevisiae* (Nasmyth et al., 1987, Cell 48:579-587). However, there are also examples of mutations that result in the formation of chimeric cell types such as the *ham-1* mutation in *C. elegans* (Desai et al., 1988, Nature 336:638-646).

6.3.2. SCR INVOLVEMENT IN CELL SPECIFICATION OR CELL DIVISION

Genes that regulate asymmetric cell divisions can be divided into those that specify the differentiated fates of the daughter cells and those that function to effect the division of the mother cell (Horvitz & Herskowitz, 1992, Cell, 68:237-255). The aberrant cell layer formed in the *scr* mutant has differentiated features of both endodermal and cortex cells. Thus, *scr* is in the rare class of asymmetric division mutants in which a chimeric cell type is created. The ability to express differentiated characteristics of cortex and endodermal cells implies that the differentiation pathways for both these cell types are intact and do not require the functional *SCR* gene. This indicates that *SCR* is involved primarily in regulating a specific cell division, and that the correct occurrence of this division can be unlinked from cell specification. This is in contrast to the *shr* mutant, in which the periclinal division of the cortex/endodermal initial also fails to occur and the resulting cell lacks endodermal markers (Benfey et al., 1993, Development 119:57-70) and has cortex attributes. A genetic analysis was used to address the function of *SHR* and *SCR* in the asymmetric division of the cortex/endodermal initial. Placing mutants of each of these genes in a *fs* mutant background asked whether the supernumerary cell divisions characteristic of *fs* were sufficient to restore normal cell identities (Scheres et al., 1995, Development 121:53-62). In the *shr,fs* double mutant there were additional cell layers but no endodermal, indicating that the *SHR* gene has a role in specifying cell identity. In the *scr,fs* double mutant no alteration in cell identity was observed as compared to *fs*

(Scheres et al., 1995, Development 121:53-62). Taken together with the cell marker analysis presented herein, these results are consistent with a role for *SCR* in generating the division of the mother cell while the *SHR* gene may be involved in specifying the fate of the endodermal daughter.

6.3.3. A ROLE FOR SCR IN EMBRYONIC DEVELOPMENT

At least one additional cell division appears to be affected in the *scr* mutant. During embryonic development, the ground tissue does not divide to form the endodermal and cortex layers of the embryonic root and hypocotyl. As shown herein, expression of *SCR* was detected in the endodermal tissue throughout the embryonic axis shortly after this division occurs. Thus, *SCR* may play a direct role in regulating both this division and the division of the cortex/endodermal initial in the root apical meristem. Alternatively, the radial organization established in the embryo may somehow act as a template that directs the division of the cortex/endodermal initial, thus perpetuating the pattern. This is consistent with the finding in the *scr* mutant that the aberrant pattern established in the embryo is perpetuated in the primary root. It is also consistent with a recent study in which the daughter cells of the cortex/endodermal initial were laser ablated (van den Berg et al., 1995, Nature 378:62-65). When a single daughter cell was ablated, it was replaced by a cell that followed the normal asymmetric division pattern. When three adjacent daughter cells were ablated, the central initial divided anticlinally but failed to perform the periclinal division (van den Berg et al., 1995, Nature 378:62-65). This provided evidence that information from mature cells is required for the correct division pattern of cortex/endodermal initials suggesting a "top down" transfer of information. However, the absence of a cell layer in lateral roots and callus-derived roots of the *scr* mutant suggests that embryo events are not unique in their ability to establish radial

organization. Rather, these observations implicate SCR in regulating both embryonic and post-embryonic root radial organization.

5 6.3.4. **TISSUE-SPECIFIC EXPRESSION OF SCR IS
REGULATED AT THE TRANSCRIPTIONAL LEVEL**

 Although not intending to be limited to any theory or explanation regarding the mechanism of SCR action, the cloning of the gene and the expression pattern provide some
10 clues as to the role of SCR in the regulation of a specific asymmetric division. The SCR gene is expressed in the cortex/endodermal initial, but immediately after division is restricted to the endodermal lineage. A similar pattern is seen in the ET199 enhancer trap line in which SCR regulatory
15 elements are in proximity to a GUS gene, indicating that SCR restriction to the endodermal cell file is due to differential regulation of expression of the SCR gene in this cell and the first cell in the cortex file. Another marker line in which expression of GUS is detected only in the
20 cortex daughter cell provides a control for differential degradation of GUS RNA or protein. Thus, partitioning of SCR RNA as a means of achieving this segregation of expression can be ruled out. What remains to be determined is whether this difference in transcriptional activity of the two
25 daughter cells is due to internal polarity of the mother cell prior to division such that cytoplasmic determinants are unequally distributed, or to external polarity that influences cell fate after division. Since SCR is expressed prior to cell division, an attractive hypothesis is that it
30 is involved in establishing polarity in the cortex/endodermal initial. The sequence of the SCR protein strongly suggests that it acts as a transcription factor. Hence, it may act to regulate the expression of other genes essential for the establishment of unequal division. Alternatively, it is
35 conceivable that it could play a role in creating an external polarity that provides a signal to divide asymmetrically.

Its expression in more mature endodermal cells is consistent with a role in "top-down" signaling.

6.3.5. A NEW FAMILY OF TRANSCRIPTIONAL REGULATORS

5 Analysis of eighteen EST clones found in the GenBank database reveals that the proteins they encode share a high degree of homology with Arabidopsis SCR protein. See Table 1 and FIGS. 15A-S. Further sequence analysis of the encoded proteins indicate that a high degree of sequence
10 similarity extends from at least the highly conserved VHIID domain to the carboxy-terminus of the gene products. Comparison of the amino termini of these proteins is precluded by the fact that the ESTs are incomplete. The high degree of similarity among these proteins, in combination
15 with the motifs observed in the SCR protein (homopolymeric motifs, two leucine heptad repeats and a bZIP-like basic domain that may also function as a nuclear localization sequence) indicates that these proteins form a novel class of regulatory proteins.

20 The insertion sites of the T-DNA in the two *scr* mutant alleles raised the possibility that the mutant phenotype was due to the production of truncated proteins. Northern blot analysis indicated SCR RNA is undetectable in *scr-1*. This suggests that the phenotype is either the null,
25 or due to highly reduced RNA expression. In *scr-2*, an alteration in RNA size was detected which would be consistent with the presence of a functional and possibly truncated protein. This could provide an explanation for the observation that *scr-2* appears to be the weaker allele.

30

7. EXAMPLE 2: ENHANCER TRAP ANALYSIS OF ROOT DEVELOPMENT

An enhancer trap system was used in order to provide a more detailed molecular analysis of gene expression in lateral root patterning and development in Arabidopsis
35 *thaliana*. A new collection of marker lines that express β -glucuronidase (GUS) activity in a cell-type specific manner in each of the cells of the root was generated. These lines

allow differentiation of cells to be monitored based on molecular characteristics. One of these marker lines, ET199, resulted from the integration of the GUS cassette in proximity to an SCR enhancer. The results described below demonstrate that transcriptional activation of the SCR gene plays an important role in root development in Arabidopsis, and that SCR gene transcriptional regulatory elements can express a transgene in a developmentally and tissue specific manner.

10

7.1. MATERIALS AND METHODS

7.1.1. PLANT GROWTH CONDITIONS:

Arabidopsis seeds from NO-O and Columbia ecotypes were sterilized and sown on MS plates containing 4.5% sucrose. Plates were oriented vertically and maintained under 18 hours light, 6 hours dark cycle.

7.1.2. HISTOLOGY AND GUS STAINING:

For observation of lateral roots, roots were removed from plates and infiltrated in 25% glycerol for several hours to overnight. Roots were then mounted in 50% glycerol. Whole seedlings were stained for GUS activity for up to three days in the following solution: 1X GUS buffer, 20% methanol, 0.5 mg/ml X-Glu. Addition of methanol greatly improves the specificity and reproducibility of staining. Staining solution was made fresh from a 10X buffer (1 M Tris pH7.5, 290 mg NaCl, 66 mg $K_3Fe(CN)_6$) that was stored for no more than one week. Stained roots were cleared in glycerol and mounted as above. All samples were observed using Nomarski optics on a Leitz Laborlux S microscope. Photographs were taken using a Leitz MPS52 camera, and images were scanned into Adobe Photoshop to create figures. In some cases the intensity of the blue color was increased.

35

7.1.3. CONSTRUCTION OF ENHANCER TRAP LINES:

Plant Cloning Vector (PCV) (Koncz et al., 1994, Specialized vectors for gene tagging and expression studies, in Plant Molecular Biology Manual, Gelvin & Schilperoort, 5 eds., Vol. B2, pp. 1-2, Kluwer Academic Press, Dordrecht, The Netherlands) contains a Bam HI site immediately adjacent to the T-DNA right border sequence. The β -glucuronidase gene fused to the TATA region (-46 to 78) of the CaMV 35S promoter was introduced into this site (Benfey et al., 1990, EMBO J. 10 9:1677-1684). 350 transgenic lines were generated by *Agrobacterium* mediated root transformation (Marton & Browse, 1991, Plant Cell Reports 10:235-239), and 4 independent lines from each transformant were screened for GUS activity in the root.

15

7.2. RESULTS

7.2.1. DIFFERENTIATION IN THE LRP

The marker lines described above reflect patterns of gene expression that are specific to individual root cell 20 types. There are no readily apparent mutant phenotypes in any of these lines. Therefore, they can be used to analyze the differentiation state of the cells during normal development of the lateral root primordial (LRP). If there are stages at which the pericycle cells proliferate in the 25 absence of patterning, it can be expected that all cells would be identical with none expressing differentiated characteristics. In contrast, organization of the LRP would be reflected in differential patterns of GUS gene expression, with certain cells beginning to turn on transcription from 30 differentiated cell-type specific promoters (i.e., those that drive GUS expression in the enhancer trap lines).

The process of lateral root formation is divided into the following seven stages:

35

Stage I: The LRP is first visible as a set of pericycle cells that are clearly shorter in length than their neighbors, having undergone a series of anticlinal divisions. Laskowski et al., 1995, Dev. 121:3303-3310 predict that there are approximately 4 founder pericycle cells involved. In the longitudinal plane, these divisions result in the formation of 8-10 small cells, which enlarge in a radial direction.

Stage II: A periclinal division occurs that divides the LRP into two layers (Upper Layer (UL) and Lower Layer (LL)). Not all the small pericycle-derived cells appear to participate in this division -- typically the most peripheral cells do not divide. Hence, as the UL and LL cells expand radially the domed shape of the LRP begins to appear.

15

Stage III: The UL divides periclinally, generating a three layer primordium comprised of UL1, UL2 and LL. Again, some peripheral cells do not divide, creating peripheral regions that are one and two cell layers thick. This further emphasizes the domed shape of the LRP.

20

Stage IV: The LL divides periclinally, creating a total of four cell layers (UL1, UL2, LL1, LL2). At this stage the LRP has penetrated the parent endodermal layer.

25

Stage V: The central cells in LL2 undergo a number of divisions that push the overlying layers up and distort the cells in LL1. These divisions are difficult to visualize at this stage, but clearly form a knot of mitotic activity. The LRP at this stage is midway through the parent cortex. The outer layer contains 10-12 cells.

30

Stage VI: This stage is characterized by several events. The four central cells of UL1 divide periclinally. This division is particularly useful in identifying the median longitudinal plane in the enlarging LRP. At this point there are a total of twelve cells in UL1, four in the middle

that have undergone the periclinal division and four on either side. In addition, all but the most central cells of UL2 undergo a periclinal division. At this point the LRP has passed through the parent cortex layer and has penetrated the epidermis. The central cells apparently derived from LL2 have a distinct elongated shape characteristic of vascular elements.

Stage VII: As the primordium enlarges it becomes difficult to characterize the divisions in the internal layers. However, the cells in the outermost layer can still be seen very clearly. All of these cells undergo a anticlinal division, resulting in 16 central cells (8 cells in each of two layers) flanked by 8-10 cells on each side. We refer to this as the 8-8-8 cell pattern. The LRP appears to be just about to emerge from the parent root.

7.2.2. MARKER LINES

An enhancer trapping cassette was generated by fusing the GUS coding sequence to the minimal promoter of the 35S promoter from CaMV. This minimal promoter does not produce a detectable level of GUS expression. However, its presence allows other upstream elements to direct GUS expression in a developmental and/or cell-specific manner (Benfey et al., 1990, EMBO J. 9:1677-1684). The use of a minimal promoter instead of a promoterless construct allows GUS expression to occur even if the enhancer trap cassette inserts at a distance from the coding region. Since the insert does not have to be within the structural gene, there are often no mutations generated in the enhancer trap lines. The minimal promoter:GUS construct was cloned immediately adjacent to the T-DNA right border sequence of PCV (Koncz et al., *supra*) and introduced into Arabidopsis. 350 independent lines were generated and analyzed for GUS activity in the root. The following lines most clearly define each cell type. All of the lines were generated through enhancer trapping, as described herein, below, except for CorAX92

(Dietrich et al., 1992, Plant Cell 4:1371-1382) and EpiGL2:GUS (Masucci et al., Dev. 122:1253-1260) which are transgenic plants that contain cell-type specific promoters fused to the GUS gene.

5

Ste05 - expresses GUS in the stele including the pericycle layer throughout primary and lateral roots. At the root tip, staining becomes weaker in the elongation zone; therefore, it is likely that only differentiated stele cells express GUS activity. Stelar GUS expression is also seen in aerial parts of the plant.

End195 - expresses GUS in the endodermis of primary and lateral roots. Staining can be seen most clearly in the cells in the meristematic region of the root, although overstaining shows that more mature cells also express some GUS activity. It appears that there is no staining in the cortex/endodermal initial, but staining is evident in the first daughter cell of this initial. GUS expression is also seen at the base of young leaves and in the stipules.

ET199 - expresses GUS in the endodermis of primary and lateral roots, again most clearly in cells in the meristematic region. Unlike End195, staining in ET199 appears to continue down to the cortex/endodermal initial and, in younger roots, even into the cells of the quiescent center. Expression in the aerial parts of the plant is detectable in the young leaf primordia.

30 CorAX92 - This line was generated by fusing the 5' and 3' sequences from a cortex specific gene isolated from oilseed rape to the GUS reporter gene (Dietrich et al., Plant Cell 4:1371-1382). Expression is limited to the cortex layer, extending to but not including the cortex/endodermal initial. Staining is also apparent in the petioles and leaf blades of expanded leaves.

EpiGL2:GUS - This line was generated by fusing the GL2 promoter to the GUS gene (Masucci et al., Dev. 122:1253-1260). Expression is seen in the non-hair forming epidermal cells (atrachoblasts). Staining is seen near the root tip, but it is difficult to determine if it includes the epidermal initial. Staining is also seen in the trichomes, leaf primordia, and the epidermis of the hypocotyl and leaf petioles.

10 CRC219 - This line shows staining in the columella root cap only.

LRC244 - This line shows staining in the lateral root cap only.

15

RC162 - This line shows staining in both the lateral and columella root caps.

Two marker lines show differential staining at very early stages of LRP development. One of these, ET199, presents a complex and dynamic pattern of expression. Staining is first apparent at stage II in only the four central cells of the UL. At stage III staining is strongest in the central cells of UL2. As the LRP reaches stage V the staining remains strongest in the central 2-4 cells of UL2. By stage VI staining also begins to extend into the newly formed endodermal layer, and staining in both the central cells and endodermis persists beyond emergence of the lateral root.

30 Another line, LRB10 (lateral root base), does not express GUS in the primary root tip. Staining in the LRP is seen at stage I, and at stage II all the cells of the UL and LL are stained. However, by stage IV and V only the cells at the periphery of the LRP are still expressing GUS. As the LRP develops, these cells continue to stain, although less intensely, resulting in a ring of GUS expressing cells at the base of the LR.

LRB10 and ET199 clearly demonstrate non-identity between the cells at very early stages, stage IV in the case of LRB10 and within the UL at stage II in ET199. In addition, although it is difficult to identify the nature of the cells that correspond to the observed staining pattern in LRB10 and the early staining cells of ET199, post-emergent lateral roots show analogous staining in these lines, suggesting that the stained cells are already expressing markers that reflect their differentiated cell fates. Hence, these observations suggest a very early onset of differentiation in the cells of the LRP.

7.2.3. ET199 PROVIDES EVIDENCE FOR THE ROLE OF SCR IN PLANT DEVELOPMENT

Fortuitously, it was discovered that the GUS cassette in ET199 described Section 7.2.2, above, is situated approximately 1 kb upstream from the *SCR* gene. The *SCR* cDNA was labelled and used to probe genomic DNA from WT and ET199 plants. The band pattern seen in the Southern was completely consistent with a T-DNA inserted 1 kb upstream of the putative *SCARECROW* start site. Subsequently, a DNA fragment was PCR amplified using a primer within the T-DNA and a primer within *SCARECROW*. The size of this fragment was also consistent with the predicted insertion site. Partial sequencing of the PCR fragment confirmed the presence of *SCARECROW* sequence. Mutants in the *SCR* gene are completely lacking one of the radial layers between the epidermis and pericycle in both primary and lateral roots, due to the absence of specific cell division during embryogenesis and of the cortex/endodermal initial during post-embryonic growth. The expression pattern (described in Section 7.2.2., above) that was observed in the central cells of the developing LRP of ET199 provide strong evidence that the cells in this region are involved in the establishment of the meristematic initials. More importantly, these results demonstrate that transcriptional activation of the *SCR* gene plays a major role in the development of the Arabidopsis LRP. Furthermore,

these results demonstrate that a transgene can be expressed under the control of *SCR* gene transcriptional regulatory elements in a developmental and tissue-specific manner.

5 8. EXAMPLE 3: ACTIVITY OF ARABIDOPSIS *SCR*
PROMOTER IN TRANSGENIC ROOTS

 The expression pattern of Arabidopsis *SCR* has been determined by analysis of an enhancer trap line, ET199, in which a GUS coding region with a minimal promoter was fortuitously inserted 1 kb upstream of the *SCR* coding region (see *supra*). In ET199 plants, GUS expression is detected in the endodermis, endodermal initials and sometimes in the quiescent center (QC) of the root. See *supra* and Malamy and Benfey, 1997, Dev. 124:33-44. This expression pattern of *SCR* in the primary root has been confirmed by *in situ* analysis (See *supra* and Di Laurenzio et al., 1996, Cell 86:423-433).

 The following experiments demonstrate that 2.5 kb of 5' sequence upstream of the Arabidopsis *SCR* coding region is sufficient to confer *SCR* expression pattern to a heterologous gene. The 5' sequence used in these studies starts from the Hind III site approximately 2.5 kb upstream of the ATG initiation site and extends 3' downstream to the base pair immediately upstream of the ATG initiation site (see FIG. 14). This 5' sequence was fused to a GUS coding sequence. The resulting *SCR* promoter::GUS construct was incorporate into an *Agrobacterium* vector, which was used to transform and generate transgenic roots using standard procedures.

 A large number of roots were regenerated. They show GUS staining pattern that is similar to the *SCR* expression pattern in ET199 plants (Figure 19, Panel f). Since organs regenerated from callus often have an abnormal morphology, transgenic roots were transferred to liquid culture. Roots grown in liquid culture appeared morphologically normal and showed GUS expression in the endodermis, endodermal initial and QC (Figure 19, Panel g), similar to the expression pattern of *SCR* seen in the

enhancer trap line ET199. These results indicate that the 2.5 kb region upstream of the *SCR* start site is sufficient to confer the *SCR* expression pattern in the root.

The expression of the *SCR* promoter::*GUS* construct 5 was also examined in *scr* mutant background. The *scr* mutant has an altered root organization (see, *supra*). Whereas the wild-type root of *Arabidopsis* has four distinct cell layers surrounding the vascular tissue, the roots of *scr* mutant have only three.

10 Transgenic roots of the *scr* mutant were generated that contained a *SCR* promoter::*GUS* construct. As in the wild-type, a large number of transgenic roots were formed that had detectable *GUS* expression (Figure 20, Panel a). These roots were shorter than wild-type regenerated roots, 15 consistent with the shorter root phenotype of the *scr* mutant.

Additional transgenic root experiments demonstrated that the *SCR* gene under control of its own promoter can rescue the *scr* mutant phenotype. Transgenic *scr* roots were generated that contained the full length *SCR* gene 20 under the control of its own promoter. The length of transgenic roots containing the construct were longer than those of the *scr* mutant, indicating that the introduced *SCR* gene partially rescued the mutant. Whereas *scr* regenerated roots that carried the *SCR* promoter::*GUS* construct were very 25 short (Figure 21, Panel a; and Figure 20, Panel a), roots transformed with the *SCR* promoter and coding region were noticeably longer (Figure 21, Panel b). The difference was even more obvious in liquid culture, in which *scr* mutant roots remained short (Figure 21, Panel c), while *SCR* gene 30 complemented *scr* mutant roots were long and resembled wild-type roots (Figure 21, Panel d).

Anatomical studies of the regenerated roots confirmed the ability of the *SCR* promoter::*SCR* gene construct to rescue the *scr* mutant phenotype. Whereas regenerated 35 roots of *scr* mutant were missing an internal layer (Figure 21, Panel e), the *scr* mutant roots that were transformed with the *SCR* promoter::*SCR* gene construct had a radial

organization that resembled wild-type root (Figure 21, Panel f).

9. EXAMPLE 4: ISOLATION SCR SEQUENCES USING PCR-CLONING STRATEGY

5 Based on the comparison of the sequences of SCR paralogs in Arabidopsis, degenerate primers SCR3AII, SCR5AII and SCR5B were designed and used in PCR amplification of SCR sequences from genomic DNA of various plant species. The
10 amplification was performed according to condition described in Section 5.1.1., supra, using DNA isolated from maize plants grown from a commercial seed mixture. Amplification products (104 bp fragment for the SCR5B+SCR3AII primer combination; 146 bp fragment for the SCR5AII+SCR3AII primer
15 combination) were obtained, and each cloned into a T/A vector (Invitrogen, San Diego, CA) and sequenced. Two of the three different types of clones obtained had deduced amino acid sequences that were very similar to a part of the Arabidopsis SCR protein (i.e., approximately 90% identity), suggesting
20 that they represent parts from two different alleles of the maize SCR gene (i.e., ZCR gene). The two clones each had only two conservative changes in their nucleotide sequence.

 The 146 bp amplification product, ZmSc11, was subsequently used as a probe for screening of a genomic
25 library generated in lambda BlueSTAR vector (NOVAGEN) from maize (HiII line) genomic DNA. The screening was performed according to the standard procedures described in Genius™ System User's Guide For Membrane Hybridization (Boehringer-Mannheim): The probe was a single-strand DNA molecule
30 corresponding to the ZmSc11 fragment produced by PCR (Genius, Boehringer-Mannheim). Hybridization was performed according to recommendations of the manufacturer's manual (Boehringer-Mannheim). Prehybridization was for 2 hr in 50% formamide hybridization solution at 42°C. Hybridization was
35 overnight at 42°C with 200 ng/ml probe concentration. Filters were washed twice at room temperature in 2xSSC, 0.1%

SDS for 5 min, and for stringent washing at 65°C in 0.5xSSC, 0.1% SDS twice for 15 min.

A positive clone was identified. The clone contained a 13 kb insert, which was subcloned into a plasmid vector. The resulting plasmid was designated pZCR. A 5 kb Eco RI fragment containing the maize *SCR* (*ZCR*) sequence was subcloned and sequenced. The nucleotide sequence of the region containing a partial *ZCR* coding sequence is shown in FIG. 17A and the corresponding deduced amino acid sequence is shown in FIG. 17B. The *ZCR* protein contain a segment that is highly homologous to a corresponding segment in the Arabidopsis *SCR* protein (FIG. 17B). This segment is flanked by segments of low homology. Thus, it is possible that the genomic clone of *ZCR* is a composite clone, containing sequences that are not *ZCR* sequences.

The deduced *ZCR* protein sequence was aligned with that of Arabidopsis *SCR* protein. The comparison revealed new conserved sites in the *SCR* coding sequence which were used to design new, more specific PCR primers (i.e., 1F, 1R, and 4R) for use in amplification of *SCR* sequences from yet other plant species.

Using combinations of primers 1F+1R and 1F+4R, PCR amplification were performed as described in section 5.1.1.. Two DNA of expected size were obtain from soybean: a 247 bp DNA from the 1F+1R primer combination and a 379 bp DNA from the 1F+4R primer combination. A DNA of expected size (247 kb) was obtained from carrot and spruce when their genomic DNA was amplified using 1F+4R primer combination. The nucleotide sequences of the 379 kb soybean DNA (*SRPg1*), the 247 kb DNA from carrot (*SRPd1*) and spruce (*SRPp1*) are shown in FIGS. 16K-M. The corresponding deduced amino acid sequences of these amplified sequences are shown in FIG. 18. Comparison of these partial *SCR* coding sequences indicate this approach isolated DNA sequences that encode *SCR* proteins with amino acid sequences that are very similar but not identical to a segment of Arabidopsis *SCR* protein (see FIG. 18).

10. EXAMPLE 5. EXPRESSION PATTERN OF MAIZE ZCR GENE
 IN ROOT TISSUE

 These experiments examined the expression pattern
of ZCR in the primary root and quiescent centers of maize
5 root. The expression pattern was determined by *in situ*
hybridization using a ZCR RNA probe, corresponding to an
amino acid segment region that is highly homologous to a
corresponding segment of the Arabidopsis SCR protein. The
experiment was carried out as follows. Restriction fragments
10 containing the maize ZCR sequence were isolated from pZCR and
subcloned into a pBluescript vector for *in vitro*
transcription. The probe was synthesized using conditions
described in the Genius Dig RNA labeling kit. The
pBluescript plasmid was linearized, and 1 μ g was used as a
15 template to synthesize digoxigenin-labeled RNA using the T7
polymerase. The RNA probe was subjected to mild alkali
hydrolysis by heated at 60°C for 1 hr in 100 mM carbonate
buffer (pH 10.2) to yield a probe size of approximately 0.15
kb. Probe concentration for hybridization was optimized at 1
20 μ g/ml/kb. *In situ* hybridization of root tips from 48 to 72
hr-old maize seedlings or excised quiescent centers (QCs) of
roots were carried out following procedures described in
Section 6.1.6., *supra*.

 The results show that ZCR expression in maize
25 primary roots is localized to a file of cells that is
identified as the endodermal layer. The expression pattern
continues in a single uninterrupted file through the QC which
consists of approximately 1000-1500 cells (FIG. 22).

 In two-week old regenerating QCs, ZCR expression
30 is found in a file of cells extending through the newly
formed apex. Thus, the regenerated roots exhibits a ZCR
expression pattern that is similar to that seen in the
primary root, even though the root apex does not contain the
normal arrangement of cell files at this stage.

 ZCR expression during regeneration of the root
35 apex was also examined. In the initial stages of
regeneration, cell proliferation occurs to fill in the

removed tissue and begins to regenerate the basic shape of the root tip. All cells on the blunt edge of the root appears to contribute to the new population of cells. The ZCR expression pattern indicates that molecular signals are differentially present in these cells at an early stage in regeneration. The gene appears to be diagnostic of cells that are preparing to undergo asymmetrical division in order to re-establish the normal organization of the root apex from the large undifferentiated cells. The results indicate that ZCR expression is required for pattern formation since it is expressed prior to the generation of any specific anatomical pattern in the newly formed not tissue.

11. EXAMPLE 6. EXPRESSION PATTERN OF ZCR GENE IN SOYBEAN ROOTS AND ROOT NODULES

SCR expression in soybean roots and nodules was examined using *in situ* hybridization with a SCR probe. The procedure used were as described in Sections 6.1.6. and 11.

In primary roots, SCR is expressed in the endodermis. Expression was also found in cells at the root tip that are located at the distal end of the endodermal cell files. In soybean nodules, expression of SCR was detected in the peripheral tissue at the site of developing vascular strands. At later stages of vascular development within the nodule, SCR expression was found flanking the vascular tissue. These results indicate that SCR is involved in regulating vascularization in the nodule by contributing to the radial organization that is required to generate endodermis. These findings indicate that SCR promoter may be used to express proteins in a highly tissue-specific manner in soybean nodules. One application is to use SCR promoter to engineer nodules through production of components in a tissue-specific manner. Another application is that modification of the expression of SCR could enhance nodule activity by improving vascularization and/or the number of endodermal layers.

12. EXAMPLE 7. SCR EXPRESSION AFFECTS GRAVITROPISM OF AERIAL STRUCTURES

In addition to being defective in specific embryonic and postembryonic meristematic divisions, both the *scr* and the *shr* mutants have shoots that exhibit severely defective gravitropism. Complementation analysis showed that *scr* is allelic to a *sgr* (shoot gravitropism) mutant, *sgr1*. Four mutant alleles of *SCR* (i.e., *scr1*, *scr2*, *sgr1-1* and *sgr1-2*) have been identified. All four of these mutants have normal root gravitropism and defective shoot gravitropism.

Etiolated hypocotyls of *scr* mutants placed on their sides do not respond to gravity even after 3 hr. Similar behaviors were observed with the inflorescence stems of *sgr1-1* mutant, which do not curve upwards even after two days on their sides. In contrast, the roots of these plants respond rapidly to the change in orientation with the same kinetics as the wild type. Thus, mutations in the *SCR* gene lead to a radial pattern deficiency in the root but have no effect on root gravitropism.

Comparable results were also obtained for *shr* roots and for hypocotyls and inflorescence stems, i.e., data indicate that *shr* shows normal root gravitropism but almost no stem gravitropism.

13. DEPOSIT OF MICROORGANISMS

The following microorganisms have been deposited in accordance with the terms of the Budapest Treaty with the American Type Culture Collection; 12301 Parklawn Drive, Rockville, MD 20852, U.S.A., on the dates indicated:

30	<u>Microorganism</u>	<u>Clone</u>	<u>Accession</u>	<u>Date</u>
			<u>No.</u>	
	DH5 α	pGEX-2TK ⁺ (pLIG 1-3/Sac+MOB1Sac)	98031	April 26, 1996
	DH5 α	pNYH1 (Zm-scl1b)	98032	April 26, 1996
	DH5 α	pNYH2 (Zm-scl1)	98033	April 26, 1996
35	DH5 α	pNYH3 (Zm-scl2)	98034	April 26, 1996
	DH5 α	pZCR		April 18, 1997

Although the invention is described in detail with reference to specific embodiments thereof, it will be understood that variations which are functionally equivalent are within the scope of this invention. Indeed, various
5 modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description and accompanying drawings such modifications are intended to fall within the scope of
10 the appended claims.

Various publications are cited herein, each of the disclosures of which is incorporated by reference in its entirety.

15

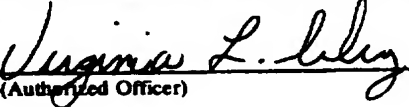
20

25

30

35

International Application No: PCT/

MICROORGANISMS	
Optional Sheet in connection with the microorganism referred to on page <u>86</u> , lines <u>25-37</u> of the description *	
A. IDENTIFICATION OF DEPOSIT * Further deposits are identified on an additional sheet *	
Name of depositary institution * American Type Culture Collection	
Address of depositary institution (including postal code and country) * 12301 Parklawn Drive Rockville, MD 20852 US	
Date of deposit * <u>April 26, 1998</u> Accession Number * <u>98031</u>	
B. ADDITIONAL INDICATIONS * (leave blank if not applicable). This information is continued on a separate attached sheet	
C. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE * (If the indications are on all designated States)	
D. SEPARATE FURNISHING OF INDICATIONS * (leave blank if not applicable) The indications listed below will be submitted to the International Bureau later * (Specify the general nature of the indications e.g., "Accession Number of Deposit")	
E. <input checked="" type="checkbox"/> This sheet was received with the International application when filed (to be checked by the receiving Office) <div style="text-align: right;"> (Authorized Officer)</div> <input type="checkbox"/> The date of receipt (from the applicant) by the International Bureau * <div style="display: flex; justify-content: space-between;"><div>was</div><div>_____ (Authorized Officer)</div></div>	

Form PCT/RO/134 (January 1981)

International Application No: PCT/ /

Form PCT/RO/134 (cont.)

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Rockville, MD 20852
US

<u>Accession No.</u>	<u>Date of Deposit</u>
98032	April 26, 1996
98033	April 26, 1996
98034	April 26, 1996
	April 18, 1997

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Banfey, Phillip N.
Di Laurenzio, Laura
Wysocka-Diller, Joanna
Malamy, Jocelyn E.
Pysh, Leonard
Helaruitta, Yrjo
- (ii) TITLE OF INVENTION: SCARECROW GENE, PROMOTER AND USES THEREOF
- (iii) NUMBER OF SEQUENCES: 67
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: Pennie & Edmonds LLP
 - (B) STREET: 1155 Avenue of the Americas
 - (C) CITY: New York
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 - (E) COUNTRY: USA
 - (F) ZIP: 10036-2711
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: US
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 08/638,617
 - (B) FILING DATE: 26-APR-1996
- (viii) ATTORNEY/AGENT INFORMATION:
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 - (C) TELEX: 66141 PENNIE

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2163 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown
- (ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CCTTATTTAT AACCATGCAA TCTCAGACC AACAAACCCTT CAATCTCCAT GCGGGAATCC

60

GGCGATTTCA	ACGGTGGTCA	ACCTCCTCCT	CATAGTCCTC	TGAGAACAAC	TTCTTCCGGT	120
AGTAGCAGCA	GCAACAACCG	TGGTCCTCCT	CCTCCTCCTC	CTCCTCCTTT	AGTGATGGTG	180
AGAAAAAGAT	TAGCTTCCGA	GATGTCTTCT	AACCCTGACT	ACAACAACCTC	CTCTCGTCCCT	240
CCTCGCCGTG	TCTCTCACCT	TCTTGACTCC	AACTACAATA	CTGTACACACC	ACAACAACCA	300
CCGTCTCTTA	CGGCGGGCGC	TACTGTATCT	TCTCAACCAA	ACCCACCACT	CTCTGTTTGT	360
GGCTTCTCTG	GTCTTCCCGT	TTTTCCTTCA	GACCGTGGTG	GTCGGAATGT	TATGATGTCC	420
GTACAACCAA	TGGATCAAGA	CTCTTCATCT	TCTTCTGCTT	CACCTACTGT	ATGGGTTGAC	480
GCCATTATCA	GAGACCTTAT	CCATTCCCTCA	ACTTCAGTCT	CTATTCCCTCA	ACTTATCCAA	540
AACGTTAGAG	ACATTATCTT	CCCTTGTAAC	CCAAATCTCG	GTGCTCTTCT	TGAATACAGG	600
CTCCGATCTC	TCATGCTCCT	TGATCCTTCC	TCTTCCTCTG	ACCCTTCTCC	TCAAACCTTC	660
GAACCTCTCT	ATCAGATCTC	CAACAATCCT	TCTCCTCCAC	AACAGCAACA	GCAGCACCAA	720
CAACAACAAC	AACAGCATAA	GCCTCCTCCT	CCTCCGATTC	AGCAGCAAGA	AAGAGAAAAT	780
TCTTCTACCG	ATGCACCACC	GCAACCAGAG	ACAGTGACGG	CCACTGTTCC	CGCCGTCCAA	840
ACAAATACGG	CGGAGGCTTT	AAGAGAGAGG	AAGGAAGAGA	TTAAGAGGCA	GAAGCAAGAC	900
GAAGAAGGAT	TACACCTTCT	CACATTGCTG	CTACAGTGTG	CTGAAGCTGT	CTCTGCTGAT	960
AATCTCGAAG	AAGCAAACAA	GCTTCTTCTT	GAGATCTCTC	AGTTATCAAC	TCCTTACGGG	1020
ACCTCAGCGC	AGAGAGTAGC	TGCTTACTTC	TCGGAAGCTA	TGTCAGCGAG	ATTACTCAAC	1080
TCGTGTCTCG	GAATTTACGC	GGCTTTGCCT	TCACGGTGGA	TGCCTCAAAC	GCATAGCTTG	1140
AAAATGGTCT	CTGCGTTTCA	GGTCTTTAAT	GGGATAAGCC	CTTTAGTGAA	ATTCTCACAC	1200
TTTACAGCGA	ATCAGGCGAT	TCAAGAAGCA	TTTGAGAAAG	AAGACAGTGT	ACACATCATT	1260
GACTTGGACA	TCATGCAGGG	ACTTCAATGG	CCTGGTTTAT	TCCACATTCT	TGCTTCTAGA	1320
CCTGGAGGAC	CTCCACACGT	GCGACTCACG	GGACTTGGTA	CTTCCATGGA	AGCTCTTCAG	1380
GCTACAGGGA	AACGTCTTTC	GGATTTTACA	GATAAGCTTG	GCCTGCCTTT	TGAGTTCTGC	1440
CCTTTAGCTG	AGAAAGTTGG	AAACTTGGAC	ACTGAGAGAC	TCAATGTGAG	GAAAAGGGAA	1500
GCTGTGGCTG	TTCACTGGCT	TCAACATTCT	CTTTATGATG	TCCTGGCTC	TGATGCACAC	1560
ACTCTCTGGT	TACTCCAAAG	GTAATAATAA	CATTACCTTT	TAATCACTCT	TTATCTATAA	1620
ATTATTTTAA	GATTATATAG	GAAAGATATG	TTCTAAAAAG	CTGGCTTTTT	TGGTTAATGA	1680
TTGGGGAATG	AACAGATTAG	CTCCTAAAGT	TGTGACAGTA	GTGGAGCAAG	ATTTGAGCCA	1740
CGCTGGTTCT	TTCTTAGGAA	GATTTGTAGA	GGCAATACAT	TACTACTCTG	CACTCTTTGA	1800
CTCACTGGGA	GCAAGCTACG	GCGAAGAGAG	TGAAGAGAGA	CATGTCGTGG	AACAGCAGCT	1860
ATTATCGAAA	GAGATACGGA	ATGTATTAGC	GGTTGGAGGA	CCATCGAGAA	GCGGTGAAGT	1920
GAAGTTTGAG	AGCTGGAGGG	AGAAAATGCA	ACAATGTGGG	TTTAAAGGTA	TATCTTTAGC	1980
TGGAAATGCA	GCTACACAAG	CGACTCTACT	GTTGGGAATG	TTTCCTTCGG	ATGGTTACAC	2040
TTTGTTTGAT	GATAATGGTA	CACCTAAGCT	TGGATGGAAA	GATCTTTTCGT	TACTCACTGC	2100

TTCAGCTTGG ACGCCTCGTT CTTAGTTTTT TTCTCCTTTT TCACAAACAA TGTGCCCATATA 2160

AAT 2163

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 653 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met	Ala	Glu	Ser	Gly	Asp	Phe	Asn	Gly	Gly	Gln	Pro	Pro	Pro	His	Ser	1	5	10	15
Pro	Leu	Arg	Thr	Thr	Ser	Ser	Gly	Ser	Ser	Ser	Ser	Asn	Asn	Arg	Gly	20	25	30	
Pro	Pro	Pro	Pro	Pro	Pro	Pro	Pro	Leu	Val	Met	Val	Arg	Lys	Arg	Leu	35	40	45	
Ala	Ser	Glu	Met	Ser	Ser	Asn	Pro	Asp	Tyr	Asn	Asn	Ser	Ser	Arg	Pro	50	55	60	
Pro	Arg	Arg	Val	Ser	His	Leu	Leu	Asp	Ser	Asn	Tyr	Asn	Thr	Val	Thr	65	70	75	80
Pro	Gln	Gln	Pro	Pro	Ser	Leu	Thr	Ala	Ala	Ala	Thr	Val	Ser	Ser	Gln	85	90	95	
Pro	Asn	Pro	Pro	Leu	Ser	Val	Cys	Gly	Phe	Ser	Gly	Leu	Pro	Val	Phe	100	105	110	
Pro	Ser	Asp	Arg	Gly	Gly	Arg	Asn	Val	Met	Met	Ser	Val	Gln	Pro	Met	115	120	125	
Asp	Gln	Asp	Ser	Ser	Ser	Ser	Ser	Ala	Ser	Pro	Thr	Val	Trp	Val	Asp	130	135	140	
Ala	Ile	Ile	Arg	Asp	Leu	Ile	His	Ser	Ser	Thr	Ser	Val	Ser	Ile	Pro	145	150	155	160
Gln	Leu	Ile	Gln	Asn	Val	Arg	Asp	Ile	Ile	Phe	Pro	Cys	Asn	Pro	Asn	165	170	175	
Leu	Gly	Ala	Leu	Leu	Glu	Tyr	Arg	Leu	Arg	Ser	Leu	Met	Leu	Leu	Asp	180	185	190	
Pro	Ser	Ser	Ser	Ser	Asp	Pro	Ser	Pro	Gln	Thr	Phe	Glu	Pro	Leu	Tyr	195	200	205	
Gln	Ile	Ser	Asn	Asn	Pro	Ser	Pro	Pro	Gln	Gln	Gln	Gln	Gln	His	Gln	210	215	220	
Gln	Gln	Gln	Gln	Gln	His	Lys	Pro	Pro	Pro	Pro	Pro	Ile	Gln	Gln	Gln	225	230	235	240
Glu	Arg	Glu	Asn	Ser	Ser	Thr	Asp	Ala	Pro	Pro	Gln	Pro	Glu	Thr	Val	245	250	255	

Thr Ala Thr Val Pro Ala Val Gln Thr Asn Thr Ala Glu Ala Leu Arg
 260 265 270
 Glu Arg Lys Glu Glu Ile Lys Arg Gln Lys Gln Asp Glu Glu Gly Leu
 275 280 285
 His Leu Leu Thr Leu Leu Leu Gln Cys Ala Glu Ala Val Ser Ala Asp
 290 295 300
 Asn Leu Glu Glu Ala Asn Lys Leu Leu Leu Glu Ile Ser Gln Leu Ser
 305 310 315 320
 Thr Pro Tyr Gly Thr Ser Ala Gln Arg Val Ala Ala Tyr Phe Ser Glu
 325 330 335
 Ala Met Ser Ala Arg Leu Leu Asn Ser Cys Leu Gly Ile Tyr Ala Ala
 340 345 350
 Leu Pro Ser Arg Trp Met Pro Gln Thr His Ser Leu Lys Met Val Ser
 355 360 365
 Ala Phe Gln Val Phe Asn Gly Ile Ser Pro Leu Val Lys Phe Ser His
 370 375 380
 Phe Thr Ala Asn Gln Ala Ile Gln Glu Ala Phe Glu Lys Glu Asp Ser
 385 390 395 400
 Val His Ile Ile Asp Leu Asp Ile Met Gln Gly Leu Gln Trp Pro Gly
 405 410 415
 Leu Phe His Ile Leu Ala Ser Arg Pro Gly Gly Pro Pro His Val Arg
 420 425 430
 Leu Thr Gly Leu Gly Thr Ser Met Glu Ala Leu Gln Ala Thr Gly Lys
 435 440 445
 Arg Leu Ser Asp Phe Thr Asp Lys Leu Gly Leu Pro Phe Glu Phe Cys
 450 455 460
 Pro Leu Ala Glu Lys Val Gly Asn Leu Asp Thr Glu Arg Leu Asn Val
 465 470 475 480
 Arg Lys Arg Glu Ala Val Ala Val His Trp Leu Gln His Ser Leu Tyr
 485 490 495
 Asp Val Thr Gly Ser Asp Ala His Thr Leu Trp Leu Leu Gln Arg Leu
 500 505 510
 Ala Pro Lys Val Val Thr Val Val Glu Gln Asp Leu Ser His Ala Gly
 515 520 525
 Ser Phe Leu Gly Arg Phe Val Glu Ala Ile His Tyr Tyr Ser Ala Leu
 530 535 540
 Phe Asp Ser Leu Gly Ala Ser Tyr Gly Glu Glu Ser Glu Glu Arg His
 545 550 555 560
 Val Val Glu Gln Gln Leu Leu Ser Lys Glu Ile Arg Asn Val Leu Ala
 565 570 575
 Val Gly Gly Pro Ser Arg Ser Gly Glu Val Lys Phe Glu Ser Trp Arg
 580 585 590
 Glu Lys Met Gln Gln Cys Gly Phe Lys Gly Ile Ser Leu Ala Gly Asn
 595 600 605
 Ala Ala Thr Gln Ala Thr Leu Leu Leu Gly Met Phe Pro Ser Asp Gly

610	615	620
Tyr Thr Leu Val Asp Asp Asn Gly Thr Leu Lys Leu Gly Trp Lys Asp		
625	630	635 640
Leu Ser Leu Leu Thr Ala Ser Ala Trp Thr Pro Arg Ser		
	645	650

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 23 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Pro Ala Val Gln Thr Asn Thr Ala Glu Ala Leu Arg Glu Arg Lys Glu
1 5 10 15
Glu Ile Lys Arg Gln Lys Gln
20

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 23 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Leu Lys Arg Ala Arg Asn Thr Glu Ala Ala Arg Arg Ser Arg Ala Arg
1 5 10 15
Lys Leu Gln Arg Met Lys Gln
20

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 23 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Arg Arg Leu Ala Gln Asn Arg Glu Ala Ala Arg Lys Ser Arg Leu Arg
1 5 10 15

Lys Lys Ala Tyr Val Gln Gln
20

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 23 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: unknown
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Ile Arg Arg Glu Arg Asn Lys Met Ala Ala Ala Lys Cys Arg Asn Arg
1 5 10 15

Arg Arg Glu Leu Thr Asp Thr
20

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 23 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: unknown
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Arg Lys Arg Met Arg Asn Arg Ile Ala Ala Ser Lys Cys Arg Lys Arg
1 5 10 15

Lys Leu Glu Arg Ile Ala Arg
20

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 23 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: unknown
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Val Arg Leu Met Lys Asn Arg Glu Ala Ala Arg Glu Cys Arg Arg Lys
1 5 10 15

Lys Lys Glu Tyr Val Lys Cys
20

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 23 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: unknown
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Lys Arg Lys Glu Ser Asn Arg Glu Ser Ala Arg Arg Ser Arg Tyr Arg
 1 5 10 15

Lys Ala Ala His Leu Lys Glu
 20

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 23 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: unknown
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Met Arg Gln Ile Arg Asn Arg Asp Ser Ala Met Lys Ser Arg Glu Arg
 1 5 10 15

Lys Lys Ser Tyr Ile Lys Asp
 20

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 23 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: unknown
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Arg Arg Met Val Ser Asn Arg Glu Ser Ala Arg Arg Ser Arg Lys Lys
 1 5 10 15

Lys Gln Ala His Leu Ala Asp
 20

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 43 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: unknown

(D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Ala Phe Glu Lys Glu Asp Ser Val His Ile Ile Asp Leu Asp Ile Met
 1 5 10 15
 Gln Gly Leu Gln Trp Pro Gly Leu Phe His Ile Leu Ala Ser Arg Pro
 20 25 30
 Gly Gly Pro Pro His Val Arg Leu Thr Gly Leu
 35 40

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 43 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: unknown

(D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Ala Val Lys Asn Glu Ser Phe Val His Ile Ile Asp Phe Gln Ile Ser
 1 5 10 15
 Gln Gly Gly Gln Trp Val Ser Leu Ile Arg Ala Leu Gly Ala Arg Pro
 20 25 30
 Gly Gly Pro Pro Asn Val Arg Ile Thr Gly Ile
 35 40

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 43 amino acids

(B) TYPE: amino acid

(C) STRANDEDNESS: unknown

(D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Ala Met Glu Gly Glu Lys Met Val His Val Ile Asp Leu Asp Ala Ser
 1 5 10 15
 Glu Pro Ala Gln Trp Leu Ala Leu Leu Gln Ala Phe Asn Ser Arg Pro
 20 25 30
 Glu Gly Pro Pro His Leu Arg Ile Thr Gly Val
 35 40

(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 29 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: unknown
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Ala Ile Lys Gly Glu Glu Glu Val His Ile Ile Asp Phe Asp Ile Asn
 1 5 10 15
 Gln Gly Asn Gln Tyr Met Thr Leu Ile Arg Ser Ile Ala
 20 25

(2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 26 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: unknown
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Ile His Val Ile Asp Phe Xaa Leu Gly Val Gly Gly Gln Trp Ala Ser
 1 5 10 15
 Phe Leu Gln Glu Leu Ala His Arg Arg Gly
 20 25

(2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 36 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: unknown
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Val His Ile Ile Xaa Phe Xaa Leu Met Gln Gly Leu Gln Trp Pro Ala
 1 5 10 15
 Leu Met Asp Val Phe Ser Ala Arg Lys Gly Gly Pr Pro Lys Leu Arg
 20 25 30
 Ile Thr Gly Ile
 35

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1085 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

GGCACGAGCC CAACGGGTCC TGAGCTTCTT ACTTATATGC ATATCTTGTA TGAAGCCTGC	60
CCTTATTTCA AATTCGGTTA TGAATCTGCT AATGGAGCTA TAGCTGAAGC TGTGAAGAAC	120
GAAAGTTTTG TGCACATTAT CGATTTCCAG ATTTCTCAAG GTGGTCAATG GGTGAGTTG	180
ATCCGTGCTC TTGGTGCTAG ACCTGGTGGA CCTCCGAACG TTAGGATAAC GGGGAATTGAT	240
GATCCGAGAT CATCGTTTGC TCGTCAAGGA GGACTTGAGT TAGTTGGACA AAGACTTGGG	300
AAGCTAGCTG AAATGTGCGG TGTTCCGTTT GAGTTCCATG GAGCTGCTTT ATGCTGCACG	360
GAAGTCGAAA TCGAGAAGCT AGGAGTTAGA AATGGAGAAG CGCTCGCGGT TAACTTCCCG	420
CTTGTTCTTC ACCACATGCC TGATGAGAGT GTAAGTGTGG AGAATCACAG AGATAGATTG	480
TTGAGATTGG TCAAACACTT GTCACCAAAC GTTGTGACTC TGGTTGAGCA AGAAGCGAAT	540
ACAAACACTG CGCCGTTTCT TCCCGGTTT GTCGAGACAA TGAACCATTA CTTGGCAGTT	600
TTCGAATCAA TAGATGTGAA ACTCGCTAGA GATCACAAGG AAAGGATCAA TGTGAGCAG	660
CATTGTTTGG CTAGAGAGGT TGTGAATCTT ATAGCTTGTTG AAGGTGTTGA AAGAGAAGAG	720
AGGCACGAGC CACTAGGGAA ATGGAGGTCT CGGTTTCACA TGGCGGGATT TAAACCGTAT	780
CCTTTGAGCT CGTATGTGAA CGCAACAATC AAAGGATTGC TTGAGAGTTA TTCAGAGAAG	840
TATACACTTG AAGAAAGAGA TGGAGCATTG TATTTAGGAT GGAAGAATCA ACCTCTTATC	900
ACTTCTTGTTG CTTGGAGGTA ACTAATAAAA ACCTTGTTTC GTTTCAGAAG AGATTAGAAA	960
CTTCTTTTAA AGTTTGCAGA ATCTGTTTGT AAAAGTAAAA CTCATGCATG ATCCGNAGGA	1020
ACAAGTTGTC AAATGTTGTA GTAGTAAGTG ATATGTTGAT GACCCAAAAA AAAAAAAAAA	1080
AAAAA	1085

(2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 306 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

Gly	Thr	Ser	Pro	Thr	Gly	Pro	Glu	Leu	Leu	Thr	Tyr	Met	His	Ile	Leu
1				5					10					15	

Tyr Glu Ala Cys Pro Tyr Phe Lys Phe Gly Tyr Glu Ser Ala Asn Gly
 20 25 30
 Ala Ile Ala Glu Ala Val Lys Asn Glu Ser Phe Val His Ile Ile Asp
 35 40 45
 Phe Gln Ile Ser Gln Gly Gly Gln Trp Val Ser Leu Ile Arg Ala Leu
 50 55 60
 Gly Ala Arg Pro Gly Gly Pro Pro Asn Val Arg Ile Thr Gly Ile Asp
 65 70 75 80
 Asp Pro Arg Ser Ser Phe Ala Arg Gln Gly Gly Leu Glu Leu Val Gly
 85 90 95
 Gln Arg Leu Gly Lys Leu Ala Glu Met Cys Gly Val Pro Phe Glu Phe
 100 105 110
 His Gly Ala Ala Leu Phe Cys Thr Glu Val Glu Ile Glu Lys Leu Gly
 115 120 125
 Val Arg Asn Gly Glu Ala Leu Ala Val Asn Phe Pro Leu Val Leu His
 130 135 140
 His Met Pro Asp Glu Ser Val Thr Val Glu Asn His Arg Asp Arg Leu
 145 150 155 160
 Leu Arg Leu Val Lys His Leu Ser Pro Asn Val Val Thr Leu Val Glu
 165 170 175
 Gln Glu Ala Asn Thr Asn Thr Ala Pro Phe Leu Pro Arg Phe Val Glu
 180 185 190
 Thr Met Asn His Tyr Leu Ala Val Phe Glu Ser Ile Asp Val Lys Leu
 195 200 205
 Ala Arg Asp His Lys Glu Arg Ile Asn Val Glu Gln His Cys Leu Ala
 210 215 220
 Arg Glu Val Glu Asn Leu Ile Ala Cys Glu Gly Val Glu Arg Glu Glu
 225 230 235 240
 Arg His Glu Pro Leu Gly Lys Trp Arg Ser Arg Phe His Met Ala Gly
 245 250 255
 Phe Lys Pro Tyr Pro Leu Ser Ser Tyr Val Asn Ala Thr Ile Lys Gly
 260 265 270
 Leu Leu Glu Ser Tyr Ser Glu Lys Tyr Thr Leu Glu Glu Arg Asp Gly
 275 280 285
 Ala Leu Tyr Leu Gly Trp Lys Asn Gln Pro Leu Ile Thr Ser Cys Ala
 290 295 300
 Trp Arg
 305

(2) INFORMATION FOR SEQ ID NO:20:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 1231 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:20:

GCTATGGAAG GAGAGAAGAT GGTTCATGTG ATTGATCTCG ATGCTTCTGA GCCAGCTCAA	60
TGGCTTGCTT TGCTTCAAGC TTTTAACTCT AGGCCTGAAG GTCCACCTCA TTTGAGAATC	120
ACTGGTGTTT ATCACCAGAA GGAAGTGCTT GAACAAATGG CTCATAGACT CATTGAGGAA	180
GCAGAGAAAC TCGATATCCC GTTTCAGTTT AATCCCGTTG TGAGTAGGTT AGACTGTTTA	240
AATGTAGAAC AGTTGCGGGT TAAAACAGGA GAGGCCTTAG CCGTTAGCTC GGTTCCTCAA	300
TTGCATACCT TCTTGGCCTC TGATGATGAT CTCATGAGAA AGAACTGCGC TTTACGGTTT	360
CAGAACAACC CTAGTGGAGT TGAAGTGAG AGAGTTCTAA TGATGAGCCA TGGCTCTGCA	420
GCTGAGGCAC GTGAGAATGA TATGAGTAAC AACAATGGGT ATAGCCCTAG CCGTGACTCG	480
GCCTCATCTT TGCCTTTACC AAGTTCAGGA AGGACTGATA GCTTCCTCAA TGCTATTTGG	540
GGTTTGTCTC CAAAGGTCAT GGTGGTCACT GAGCAAGACT CAGACCACAA CGGCTCCACA	600
CTAATGGAGA GGCTATTAGA ATCACTTTAC ACCTACGCAG CATTGTTTGA TTGCTTGGAA	660
ACAAAAGTTC CAAGAACGTC TCAAGATAGG ATCAAAGTGG AGAAGATGCT CTTGCGGGAG	720
GAGATCAAGA ACATCATATC CTGCGAGGGA TTTGAGAGAA GAGAAAGACA CGAGAAGCTT	780
GAGAAATGGA GCCAGAGGAT CGATTGGGCT GGTTTTGGGA ATGTTCTCTT TAGCTATTAT	840
GGGATGTTGC AGGCTAGGAG ATTGCTTCAA GGGTGCGGTT TTGATGGGTA TAGAATCAAG	900
GAAGAGAGCG GGTGCCGAGT AATTTGCTGG CAAGATCGAC CTCTATACTC GGTATCAGCT	960
TGGAGATGCA GGAAGTGAAT GATATATTAC AGTTTGTCTT CTATTTTGGT TATGAGCAGA	1020
GTCCCTTTCT TTTTGTATA CATGGGGACA CAATCTTAGT TGTTTTGTGA TGGTGACTTT	1080
CTGTCTCTTT ATGCTATTTT GGCTTAAATG CTTCTACTGC CTCTGCATGT AAAGCCTTTG	1140
TGTGTTGGTT CAATTTGGTC TGGTGTGGGT GTAATACCAA ACCAAATCCA ATTTGAGCTG	1200
AAGATAACTA ATTTGATGAT CGGCTCGTGC C	1231

(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 325 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Ala Met Glu Gly Glu Lys Met Val His Val Ile Asp Leu Asp Ala Ser	
1 5 10 15	
Glu Pro Ala Gln Trp Leu Ala Leu Leu Gln Ala Phe Asn Ser Arg Pro	
20 25 30	

Glu Gly Pro Pro His Leu Arg Ile Thr Gly Val His His Gln Lys Glu
 35 40 45
 Val Leu Glu Gln Met Ala His Arg Leu Ile Glu Glu Ala Glu Lys Leu
 50 55 60
 Asp Ile Pro Phe Gln Phe Asn Pro Val Val Ser Arg Leu Asp Cys Leu
 65 70 75 80
 Asn Val Glu Gln Leu Arg Val Lys Thr Gly Glu Ala Leu Ala Val Ser
 85 90 95
 Ser Val Leu Gln Leu His Thr Phe Leu Ala Ser Asp Asp Asp Leu Met
 100 105 110
 Arg Lys Asn Cys Ala Leu Arg Phe His Asn Asn Pro Ser Gly Val Asp
 115 120 125
 Leu Gln Arg Val Leu Met Met Ser His Gly Ser Ala Ala Glu Ala Arg
 130 135 140
 Glu Asn Asp Met Ser Asn Asn Asn Gly Tyr Ser Pro Ser Gly Asp Ser
 145 150 155 160
 Ala Ser Ser Leu Pro Leu Pro Ser Ser Gly Arg Thr Asp Ser Phe Leu
 165 170 175
 Asn Ala Ile Trp Gly Leu Ser Pro Lys Val Met Val Val Thr Glu Gln
 180 185 190
 Asp Ser Asp His Asn Gly Ser Thr Leu Met Glu Arg Leu Leu Glu Ser
 195 200 205
 Leu Tyr Thr Tyr Ala Ala Leu Phe Asp Cys Leu Glu Thr Lys Val Pro
 210 215 220
 Arg Thr Ser Gln Asp Arg Ile Lys Val Glu Lys Met Leu Phe Gly Glu
 225 230 235 240
 Glu Ile Lys Asn Ile Ile Ser Cys Glu Gly Phe Glu Arg Arg Glu Arg
 245 250 255
 His Glu Lys Leu Glu Lys Trp Ser Gln Arg Ile Asp Leu Ala Gly Phe
 260 265 270
 Gly Asn Val Pro Leu Ser Tyr Tyr Ala Met Leu Gln Ala Arg Arg Leu
 275 280 285
 Leu Gln Gly Cys Gly Phe Asp Gly Tyr Arg Ile Lys Glu Glu Ser Gly
 290 295 300
 Cys Ala Val Ile Cys Trp Gln Asp Arg Pro Leu Tyr Ser Val Ser Ala
 305 310 315 320
 Trp Arg Cys Arg Lys
 325

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1368 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

CTTTGTCAAT GGTAAATGAG CTGAGGCAGA TAGTTTCTAT CCAAGGAGAC CCTTCTCAGA	60
GAATCGCAGC TTACATGGTG GAAGGTCTAG CTGCAAGAAT GGCCGCTTCA GGAAAATTCA	120
TCTACAGAGC ATTGAAATGC AAAGAGCCTC CTTCGGATGA GAGGCTTGCA GCTATGCAAG	180
TCCTGTTTGA AGTCTGCCCT TGTTCAGT TCGGGTTTTT AGCAGCTAAT GGTGCGATAC	240
TTGAAGCAAT CAAAGGTGAA GAAGAAGTTC ACATAATCGA TTTCGATATA AACCAAGGGA	300
ACCAATACAT GACACTGATA CGAAGCATTG CTGAGTTGCC TGGTAAACGA CCTCGCCTGA	360
GGTTAACAGG AATTGATGAC CCTGAATCAG TCCAACGCTC CATTGGAGGG CTAAGAATCA	420
TCGGTCTAAG ACTCGAGCAA CTCGCAGAGG ATAATGGAGT ATCCTTCAAA TTCAAAGCAA	480
TGCCTTCAAA GACTTCGATT GTCTCTCCAT CAACACTCGG TTGCAAACCA GGAGAAACCT	540
TAATAGTGAA CTTTGCATTG CAACTTCACC ACATGCCTGA CGAGAGTGTC ACAACAGTAA	600
ACCAGCGGGA CGAGCTACTT CACATGGTCA AAAGCTTAAA CCCAAAGCTT GTCACGGTCG	660
TTGAACAAGA CGTGAACACA AACACTTCAC CGTTCTTTCC CAGATTCATA GAGGCTTACG	720
AATACTACTC AGCAGTTTTT GAGTCTCTAG ACATGACACT TCCAAGAGAA AGCCAAGAGA	780
GGATGAATGT AGAAAGACAG TGTCTCGCTA GAGACATAGT CAACATTGTT GCTTGCGAAG	840
GAGAAGAACG GATAGAGAGA TACGAGGCTG CGGGAAAATG GAGAGCAAGG ATGATGATGG	900
CTGGATTCAA TCCAAAACCA ATGAGTGCTA AAGTAACCAA CAATATACAA AACCTGATAA	960
AGCAACAATA TTGCAATAAG TACAAGCTTA AAGAAGAAAT GGGTGAGCTC CATTTTTGCT	1020
GGGAGGAGAA AAGCTTAATC GTTGCTTCAG CTTGGAGGTA AGATAAGTGA CAAGAGCATA	1080
TAGTCTTTAT GTTTCATAAA ACATAATTAT GTTTTTACTG TAATCTTGGG TTATTGTGTA	1140
ACTGGTTAAA TCATCTCCAT GTATTATTAC CAGAGGTTAG GGGTGATCAC AGGTACTAAA	1200
AGCTAATCTA ACACTTATGG AAGAATTTTT CTTTCTTTTT TTTCCCTATT ATATAAAAT	1260
AATTAGAGTT TTGGTTCTAA ACCTATTTGC TAAGTGCGAA TGAGTCTTTA CATGTTTATA	1320
TTTCAGTTCA AATGGTTAAA TTTGTTAAGG TTCTCACTTA AAAAAAAA	1368

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 351 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

Leu	Ser	Met	Val	Asn	Glu	Leu	Arg	Gln	Ile	Val	Ser	Ile	Gln	Gly	Asp
1				5					10					15	

Pro Ser Gln Arg Ile Ala Ala Tyr Met Val Glu Gly Leu Ala Ala Arg
 20 25 30
 Met Ala Ala Ser Gly Lys Phe Ile Tyr Arg Ala Leu Lys Cys Lys Glu
 35 40 45
 Pro Pro Ser Asp Glu Arg Leu Ala Ala Met Gln Val Leu Phe Glu Val
 50 55 60
 Cys Pro Cys Phe Lys Phe Gly Phe Leu Ala Ala Asn Gly Ala Ile Leu
 65 70 75 80
 Glu Ala Ile Lys Gly Glu Glu Glu Val His Ile Ile Asp Phe Asp Ile
 85 90 95
 Asn Gln Gly Asn Gln Tyr Met Thr Leu Ile Arg Ser Ile Ala Glu Leu
 100 105 110
 Pro Gly Lys Arg Pro Arg Leu Arg Leu Thr Gly Ile Asp Asp Pro Glu
 115 120 125
 Ser Val Gln Arg Ser Ile Gly Gly Leu Arg Ile Ile Asn Leu Arg Leu
 130 135 140
 Glu Gln Leu Ala Glu Asp Asn Gly Val Ser Phe Lys Phe Lys Ala Met
 145 150 155 160
 Pro Ser Lys Thr Ser Ile Val Ser Pro Ser Thr Leu Gly Cys Lys Pro
 165 170 175
 Gly Glu Thr Leu Ile Val Asn Phe Ala Phe Gln Leu His His Met Pro
 180 185 190
 Asp Glu Ser Val Thr Thr Val Asn Gln Arg Asp Glu Leu Leu His Met
 195 200 205
 Val Lys Ser Leu Asn Pro Leu Val Thr Val Val Glu Gln Asp Val Asn
 210 215 220
 Thr Asn Thr Ser Pro Phe Phe Pro Arg Phe Ile Glu Ala Tyr Glu Tyr
 225 230 235 240
 Tyr Ser Ala Val Phe Glu Ser Leu Asp Met Thr Leu Pro Arg Glu Ser
 245 250 255
 Gln Glu Arg Met Asn Val Glu Arg Gln Cys Leu Ala Arg Asp Ile Val
 260 265 270
 Asn Ile Val Ala Cys Glu Gly Glu Glu Arg Ile Glu Arg Tyr Glu Ala
 275 280 285
 Ala Gly Lys Trp Arg Ala Arg Met Met Met Ala Gly Phe Asn Pro Lys
 290 295 300
 Pro Met Ser Ala Lys Val Thr Asn Asn Ile Gln Asn Leu Ile Lys Gln
 305 310 315 320
 Gln Tyr Cys Asn Lys Tyr Lys Leu Lys Glu Glu Met Gly Glu Leu His
 325 330 335
 Phe Cys Trp Glu Glu Lys Ser Leu Ile Val Ala Ser Ala Trp Arg
 340 345 350

(2) INFORMATION FOR SEQ ID NO:24:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 100 base pairs

- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

CCAGGAGGCG TTCGAGCGGG AGGAGCGTGT GCACATCATC GACCTCGACA TCATGCAGGG 60
GCTGCAGTGG CCGGGCCTCC TCCACATCCT TGCCTCCCGC 100

(2) INFORMATION FOR SEQ ID NO:25:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 33 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

Gln Glu Ala Phe Glu Arg Glu Glu Arg Val His Ile Ile Asp Leu Asp
1 5 10 15
Ile Met Gln Gly Leu Gln Trp Pro Gly Leu Phe His Ile Leu Ala Ser
20 25 30
Arg

(2) INFORMATION FOR SEQ ID NO:26:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1094 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

CCACGCGTCC GTCAAAGGAT ACAACCATGT ACACATAATT GACTTTTCCC TGATGCAAGG 60
TCTCCAGTGG CCGGCACTCA TGGATGTCTT CTCCGCCCGT GAGGGTGGGC CACCAAAGCT 120
CCGAATCACA GGCATTGGCC CGAACCCAAT AGGTGGCCGT GACGAGCTCC ATGAAGTGGG 180
AATTCGCCTC GCCAAGTATG CACACTCGGT GGGTATCGAC TTCACTTTCC AGGGAGTCTG 240
TGTGATCAA CTTGATAGGT TGTGCGACTG GATGCTTCTC AAACCAATCA AAGGAGAGGC 300
AGTTGCCATA AACTCCATCC TACAACTCCA TCGCCTCCTC GTTGACCCAG ATGCAAACCC 360
AGTGGTGCCC GCACCAATAG ATATCCTCCT CAAATTGGTC ATCAAGATAA ACCCCATGAT 420

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CTTCACGGTG GTTGAGCATG AGGCAGATCA CAACAGACCA CCACTACTAG AGAGGTTTAC      480
TAATGCCCTC TTCCACTATG CGACCATGTT TGA CTCTTTG GAGGCCATGC ATCGTTGTAC      540
CAGTGGTAGA GACATCACCG ACTCACTCAC AGAGGTGTAC CTTCGAGGTG AGATTTTGA      600
CATTGTCTGC GGCGAGGGCA GTGCACGCAC CGAACGTCAT GAGTTGTTTG GTCACTGGAG      660
GGAGAGGCTC ACCTATGCTG GGCTAACTCA AGTGTGGTTC GACCCCGATG AGGTTGACAC      720
GCTAAAAGAC CAGTTGATCC ATGTGACATC CTTATCTGGC TCTGGGTTCA ACATCCTAGT      780
GTGTGATGGC AGCCTTGAC TAGCGTGGCA TAATCGCCCG TTATATGTGG CAACAGCTTG      840
GTGTGTGACA GGAGGAAATG CTGCCAGTTC CATGGTTGGC AACATCTGTA AGGGTACAAA      900
TGATAGTAGA AGAAAGGAAA ACCGTAATGG ACCCATGGAG TAGCAGGAAG AATAACCATG      960
TCATGAGCAA ATCGATCAAG TAATAAATG CACTGATGAC ATGCATGGTG ATCTAAAGTT     1020
TTTTTGCGTG AATGTGCAAT GACGAATTGT TCAATTTGAA TAACCTAATC ATGAGACTCA     1080
AAAAAAAAAA AAAA                                     1094

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(2) INFORMATION FOR SEQ ID NO:27:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 313 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

```

His Ala Ser Val Lys Gly Tyr Asn His Val His Ile Ile Asp Phe Ser
1           5           10           15
Leu Met Gln Gly Leu Gln Trp Pro Ala Leu Met Asp Val Phe Ser Ala
20        25        30
Arg Glu Gly Gly Pro Pro Lys Leu Arg Ile Thr Gly Ile Gly Pro Asn
35        40        45
Pro Ile Gly Gly Arg Asp Glu Leu His Glu Val Gly Ile Arg Leu Ala
50        55        60
Lys Tyr Ala His Ser Val Gly Ile Asp Phe Thr Phe Gln Gly Val Cys
65        70        75        80
Val Asp Gln Leu Asp Arg Leu Cys Asp Trp Met Leu Leu Lys Pro Ile
85        90        95
Lys Gly Glu Ala Val Ala Ile Asn Ser Ile Leu Gln Leu His Arg Leu
100       105       110
Leu Val Asp Pro Asp Ala Asn Pro Val Val Pro Ala Pro Ile Asp Ile
115      120      125
Leu Leu Lys Leu Val Ile Lys Ile Asn Pro Met Ile Phe Thr Val Val
130      135      140
Glu His Glu Ala Asp His Asn Arg Pro Pro Leu Leu Glu Arg Phe Thr

```


145	150	155	160
Asn Ala Leu Phe	His Tyr Ala Thr Met	Phe Asp Ser Leu Glu Ala Met	
	165	170	175
His Arg Cys Thr	Ser Gly Arg Asp Ile	Thr Asp Ser Leu Thr Glu Val	
	180	185	190
Tyr Leu Arg Gly Glu Ile Phe	Asp Ile Val Cys Gly Glu Gly Ser Ala		
	195	200	205
Arg Thr Glu Arg His Glu	Leu Phe Gly His Trp Arg Glu Arg Leu Thr		
	210	215	220
Tyr Ala Gly Leu Thr Gln Val Trp Phe	Asp Pro Asp Glu Val Asp Thr		
	225	230	235
Leu Lys Asp Gln	Leu Ile His Val Thr Ser Leu Ser Gly Ser Gly Phe		
	245	250	255
Asn Ile Leu Val Cys Asp Gly Ser	Leu Ala Leu Ala Trp His Asn Arg		
	260	265	270
Pro Leu Tyr Val Ala Thr Ala	Trp Cys Val Thr Gly Gly Asn Ala Ala		
	275	280	285
Ser Ser Met Val Gly Asn Ile	Cys Lys Gly Thr Asn Asp Ser Arg Arg		
	290	295	300
Lys Glu Asn Arg Asn Gly Pro Met Glu			
	305	310	

(2) INFORMATION FOR SEQ ID NO:28:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 611 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

CCCAACTTGG GAAGCCCTTC CTCCGCTCCG CCTCCTACCT CAAGGAGGCC CTCCTCCTCG	60
CACTCGCCGA CAGCCACCAT GGCTCCTCCG GCGTCACCTC GCGCTCGAC GTTGCCCTCA	120
AGCTTGCAGC ATACAAGTCT TTCTCTGACC TGTCACCTGT GCTCCAGTTC ACTAACTTTA	180
CCGCAACAAG GCGCTTCTTG ATGAGATTGG TGGCATGGCA ACTTCCTGCA TCCATGTCAT	240
TGACTTTGAT CTCGGTGTG GTGGTCAGTG GGCTTCCTTC TTGCAGGAGC TTGCCACCG	300
CCGGGGAGCT GGAGGTATGG CCTTGCCGTT GTTGAAGCTC ACGGCTTTCA TGTCGACTGC	360
TTCTCACCAT CCACTGGAGC TGCACCTTAC CCAGGATAAC CTCTCTCAGT TTGCCGCAGA	420
GCTCAGAATT CCTTTGAAT TCAATGCCGT CAGTCTTGAT GCATTCAATC CTGCGGAATC	480
TATTTCTTCC TCTGGTGATG AAGTTGTTGC TGTTAGCCTC CCTGTTGGCT GCTCTGCTCG	540
TGCACCACCG CTGCCAGCGA TTCTTCGGTT GGTGAAACAG CTTTGTCTTA AGGTTGTCGT	600

GGCTATTGAT C

611

(2) INFORMATION FOR SEQ ID NO:29:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 502 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

```

TTTTTTTTTT TTTTTTTTTT TTTTTTTTTT TACAGAGCAA CAGCAGTATA ATATTAATTC      60
TGTACCACAC AACCATTGTA TAGGTTAAAT TACCCTCTAG TCTCTACTCA TAAGCAGTGT      120
TTCCAATGAG ATGATCATGG CTAATTGAGC AGAGCATGGC AACAACCTAA AGCAACATCA      180
TTAGCTATAG AGACTGACAC CAATATTCCT AAATCCACTA GGCTAGCTAA TAAGCTGCAA      240
CGAAAAGCAA TATGAAGAGT TCAACAGCTC AAGACAACAA TTTCATTGTC AACATTTAAT      300
TGCAAGAATA AATGGACATT ACTGGAGTGG TCGATGCTTG CAAACGGTGG TGGAACCTTG      360
GTGGAGTGAA GCTTATGGCT GATCAGCACC GCCAAGATGA TATGGATACA AGCTCCCCAC      420
GCTGCCAGTA GAGCGTAAGA GCAGCTCCGC GTTCTCCAC ATGGAATCCT CGGACCTGCA      480
CCCGCTTCAG GAGGCAGTCT GC                                          502

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(2) INFORMATION FOR SEQ ID NO:30:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 298 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

```

Pro Gln Gln Gln Gln Gln His Gln Gln Gln Gln Gln Gln His Lys Pro
1           5           10           15
Pro Pro Pro Pro Ile Gln Gln Gln Glu Arg Glu Asn Ser Ser Thr Asp
20          25          30
Ala Pro Pro Gln Pro Glu Thr Val Thr Ala Thr Val Pro Ala Val Gln
35          40          45
Thr Asn Thr Ala Glu Ala Leu Arg Glu Arg Lys Glu Glu Ile Lys Arg
50          55          60
Gln Lys Gln Asp Glu Glu Gly Leu His Leu Leu Thr Leu Leu Leu Gln
65          70          75          80
Cys Ala Glu Ala Val Ser Ala Asp Asn Leu Glu Glu Ala Asn Lys Leu
85          90          95

```

Leu Leu Glu Ile Ser Gln Leu Ser Thr Pro Tyr Gly Thr Ser Ala Gln
 100 105 110
 Arg Val Ala Ala Tyr Phe Ser Glu Ala Met Ser Ala Arg Leu Leu Asn
 115 120 125
 Ser Cys Leu Gly Ile Tyr Ala Ala Leu Pro Ser Arg Trp Met Pro Gln
 130 135 140
 Thr His Ser Leu Lys Met Val Ser Ala Phe Gln Val Phe Asn Gly Ile
 145 150 155 160
 Ser Pro Leu Val Lys Phe Ser His Phe Thr Ala Asn Gln Ala Ile Gln
 165 170 175
 Glu Ala Phe Glu Lys Glu Asp Ser Val His Ile Ile Asp Leu Asp Ile
 180 185 190
 Met Gln Gly Leu Gln Trp Pro Gly Leu Phe His Ile Leu Ala Ser Arg
 195 200 205
 Pro Gly Gly Pro Pro His Val Arg Leu Thr Gly Leu Gly Thr Ser Met
 210 215 220
 Glu Ala Leu Gln Ala Thr Gly Lys Arg Leu Ser Asp Phe Thr Asp Lys
 225 230 235 240
 Leu Gly Leu Pro Phe Glu Phe Cys Pro Leu Ala Glu Lys Val Gly Asn
 245 250 255
 Asp Leu Thr Glu Arg Leu Asn Val Arg Lys Arg Glu Ala Ala Val His
 260 265 270
 Trp Leu Gln His Ser Leu Tyr Asp Val Thr Gly Ser Asp Ala His Thr
 275 280 285
 Leu Trp Leu Leu Gln Arg Leu Ala Pro Lys
 290 295

(2) INFORMATION FOR SEQ ID NO:31:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 307 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

Gly Thr Ser Pro Thr Gly Pro Glu Leu Leu Thr Tyr Met His Ile Leu
 1 5 10 15
 Tyr Glu Ala Cys Pro Tyr Phe Lys Phe Gly Tyr Glu Ser Ala Asn Gly
 20 25 30
 Ala Ile Ala Glu Ala Val Lys Asn Glu Ser Phe Val His Ile Ile Asp
 35 40 45
 Phe Gln Ile Ser Gln Gly Gly Gln Trp Val Ser Leu Ile Arg Ala Leu
 50 55 60
 Gly Ala Arg Pro Gly Gly Pro Pro Asn Val Arg Ile Thr Gly Ile Asp

65	70					75					80				
Asp	Pro	Arg	Ser	Ser	Phe	Ala	Arg	Gln	Gly	Gly	Leu	Glu	Leu	Val	Gly
				85					90					95	
Gln	Arg	Leu	Gly	Lys	Leu	Ala	Glu	Met	Cys	Gly	Val	Pro	Phe	Glu	Phe
			100					105					110		
His	Gly	Ala	Ala	Leu	Cys	Cys	Thr	Glu	Val	Glu	Ile	Glu	Lys	Leu	Gly
		115					120					125			
Val	Arg	Asn	Gly	Glu	Ala	Leu	Ala	Val	Asn	Phe	Pro	Leu	Val	Leu	His
	130					135					140				
His	Met	Pro	Asp	Glu	Ser	Val	Thr	Val	Glu	Asn	His	Arg	Asp	Arg	Leu
145					150					155					160
Leu	Arg	Leu	Val	Lys	His	Leu	Ser	Pro	Asn	Val	Val	Thr	Leu	Val	Glu
				165					170					175	
Gln	Glu	Ala	Asn	Thr	Asn	Thr	Ala	Pro	Phe	Leu	Pro	Arg	Phe	Val	Glu
			180					185					190		
Thr	Met	Asn	His	Tyr	Leu	Ala	Val	Phe	Glu	Ser	Ile	Asp	Val	Lys	Leu
	195						200					205			
Ala	Arg	Asp	His	Lys	Glu	Arg	Ile	Asn	Val	Glu	Gln	His	Cys	Leu	Ala
	210					215					220				
Arg	Glu	Val	Val	Asn	Leu	Ile	Ala	Cys	Glu	Gly	Val	Glu	Arg	Glu	Glu
225				230						235				240	
Arg	His	Glu	Pro	Leu	Gly	Lys	Trp	Arg	Ser	Arg	Phe	His	Met	Ala	Gly
				245					250					255	
Phe	Lys	Pro	Tyr	Pro	Leu	Ser	Ser	Tyr	Val	Asn	Ala	Thr	Ile	Lys	Gly
			260					265					270		
Leu	Leu	Glu	Ser	Tyr	Ser	Glu	Lys	Tyr	Thr	Leu	Glu	Glu	Arg	Asp	Gly
	275						280					285			
Ala	Leu	Tyr	Leu	Gly	Trp	Lys	Asn	Gln	Pro	Leu	Ile	Thr	Ser	Cys	Ala
	290					295					300				
Trp	Arg	Xaa													
305															

(2) INFORMATION FOR SEQ ID NO:32:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 353 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

Leu	Ser	Met	Val	Asn	Glu	Leu	Arg	Gln	Ile	Val	Ser	Ile	Gln	Gly	Asp
1				5				10					15		
Pro	Ser	Gln	Arg	Ile	Ala	Ala	Tyr	Met	Val	Glu	Gly	Leu	Ala	Ala	Arg
			20				25					30			

Met Ala Ala Ser Gly Lys Phe Ile Tyr Arg Ala Leu Lys Cys Lys Glu
 35 40 45
 Pro Pro Ser Asp Glu Arg Leu Ala Ala Met Gln Val Leu Phe Glu Val
 50 55 60
 Cys Pro Cys Phe Lys Phe Gly Phe Leu Ala Ala Asn Gly Ala Ile Leu
 65 70 75 80
 Glu Ala Ile Lys Gly Glu Glu Glu Val His Ile Ile Asp Phe Asp Ile
 85 90 95
 Asn Gln Gly Asn Gln Tyr Met Thr Leu Ile Arg Ser Ile Ala Glu Leu
 100 105 110
 Pro Gly Lys Arg Pro Arg Leu Arg Leu Thr Gly Ile Asp Asp Pro Glu
 115 120 125
 Ser Val Gln Arg Ser Ile Gly Gly Leu Arg Ile Ile Gly Leu Arg Leu
 130 135 140
 Glu Gln Leu Ala Glu Asp Asn Gly Val Ser Phe Lys Phe Lys Ala Met
 145 150 155 160
 Pro Ser Lys Thr Ser Ile Val Ser Pro Ser Thr Leu Gly Cys Lys Pro
 165 170 175
 Gly Glu Thr Leu Ile Val Asn Phe Ala Phe Gln Leu His His Met Pro
 180 185 190
 Asp Glu Ser Val Thr Thr Val Asn Gln Arg Asp Glu Leu Leu His Met
 195 200 205
 Val Lys Ser Leu Asn Pro Lys Leu Val Thr Val Val Glu Gln Asp Val
 210 215 220
 Asn Thr Asn Thr Ser Pro Phe Phe Pro Arg Phe Ile Glu Ala Tyr Glu
 225 230 235 240
 Tyr Tyr Ser Ala Val Phe Glu Ser Leu Asp Met Thr Leu Pro Arg Glu
 245 250 255
 Ser Gln Glu Arg Met Asn Val Glu Arg Gln Cys Leu Ala Arg Asp Ile
 260 265 270
 Val Asn Ile Val Ala Cys Glu Gly Glu Glu Arg Ile Glu Arg Tyr Glu
 275 280 285
 Ala Ala Gly Lys Trp Arg Ala Arg Met Met Met Ala Gly Phe Asn Pro
 290 295 300
 Lys Pro Met Ser Ala Lys Val Thr Asn Asn Ile Gln Asn Leu Ile Lys
 305 310 315 320
 Gln Gln Tyr Cys Asn Lys Tyr Lys Leu Lys Glu Glu Met Gly Glu Leu
 325 330 335
 His Phe Cys Trp Glu Glu Lys Ser Leu Ile Val Ala Ser Ala Trp Arg
 340 345 350
 Xaa

(2) INFORMATION FOR SEQ ID NO:33:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 326 amino acids

(B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

Ala	Met	Glu	Gly	Glu	Lys	Met	Val	His	Val	Ile	Asp	Leu	Asp	Ala	Ser	1	5	10	15
Glu	Pro	Ala	Gln	Trp	Leu	Ala	Leu	Leu	Gln	Ala	Phe	Asn	Ser	Arg	Pro	20	25	30	
Glu	Gly	Pro	Pro	His	Leu	Arg	Ile	Thr	Gly	Val	His	His	Gln	Lys	Glu	35	40	45	
Val	Leu	Glu	Gln	Met	Ala	His	Arg	Leu	Ile	Glu	Glu	Ala	Glu	Lys	Leu	50	55	60	
Asp	Ile	Pro	Phe	Gln	Phe	Asn	Pro	Val	Val	Ser	Arg	Leu	Asp	Cys	Leu	65	70	75	80
Asn	Val	Glu	Gln	Leu	Arg	Val	Lys	Thr	Gly	Glu	Ala	Leu	Ala	Val	Ser	85	90	95	
Ser	Val	Leu	Gln	Leu	His	Thr	Phe	Leu	Ala	Ser	Asp	Asp	Asp	Leu	Met	100	105	110	
Arg	Lys	Asn	Cys	Ala	Leu	Arg	Phe	Gln	Asn	Asn	Pro	Ser	Gly	Val	Asp	115	120	125	
Leu	Gln	Arg	Val	Leu	Met	Met	Ser	His	Gly	Ser	Ala	Ala	Glu	Ala	Arg	130	135	140	
Glu	Asn	Asp	Met	Ser	Asn	Asn	Asn	Gly	Tyr	Ser	Pro	Ser	Gly	Asp	Ser	145	150	155	160
Ala	Ser	Ser	Leu	Pro	Leu	Pro	Ser	Ser	Gly	Arg	Thr	Asp	Ser	Phe	Leu	165	170	175	
Asn	Ala	Ile	Trp	Gly	Leu	Ser	Pro	Lys	Val	Met	Val	Val	Thr	Glu	Gln	180	185	190	
Asp	Ser	Asp	His	Asn	Gly	Ser	Thr	Leu	Met	Glu	Arg	Leu	Leu	Glu	Ser	195	200	205	
Leu	Tyr	Thr	Tyr	Ala	Ala	Leu	Phe	Asp	Cys	Leu	Glu	Thr	Lys	Val	Pro	210	215	220	
Arg	Thr	Ser	Gln	Asp	Arg	Ile	Lys	Val	Glu	Lys	Met	Leu	Phe	Gly	Glu	225	230	235	240
Glu	Ile	Lys	Asn	Ile	Ile	Ser	Cys	Glu	Gly	Phe	Glu	Arg	Arg	Glu	Arg	245	250	255	
His	Glu	Lys	Leu	Glu	Lys	Trp	Ser	Gln	Arg	Ile	Asp	Leu	Ala	Gly	Phe	260	265	270	
Gly	Asn	Val	Pro	Leu	Ser	Tyr	Tyr	Ala	Met	Leu	Gln	Ala	Arg	Arg	Leu	275	280	285	
Leu	Gln	Gly	Cys	Gly	Phe	Asp	Gly	Tyr	Arg	Ile	Lys	Glu	Glu	Ser	Gly	290	295	300	

Cys Ala Val Ile Cys Trp Gln Asp Arg Pro Leu Tyr Ser Val Ser Ala
 305 310 315 320
 Trp Arg Cys Arg Lys Xaa
 325

(2) INFORMATION FOR SEQ ID NO:34:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 277 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

Asn Lys Arg Leu Lys Ser Cys Ser Ser Pro Asp Ser Met Val Thr Ser
 1 5 10 15
 Thr Ser Thr Gly Thr Gln Ile Gly Gly Val Ile Gly Thr Thr Val Thr
 20 25 30
 Thr Thr Thr Thr Thr Thr Thr Ala Ala Ala Glu Ser Thr Arg Ser Val
 35 40 45
 Ile Leu Val Asp Ser Gln Glu Asn Gly Val Arg Leu Val His Ala Leu
 50 55 60
 Met Ala Cys Ala Glu Ala Ile Gln Gln Asn Asn Leu Thr Leu Ala Glu
 65 70 75 80
 Ala Leu Val Lys Gln Ile Gly Cys Leu Ala Val Ser Gln Ala Gly Ala
 85 90 95
 Met Arg Lys Val Ala Thr Tyr Phe Ala Glu Ala Leu Ala Arg Arg Ile
 100 105 110
 Tyr Arg Leu Ser Pro Pro Gln Asn Gln Ile Asp His Cys Leu Ser Asp
 115 120 125
 Thr Leu Gln Met His Phe Tyr Glu Thr Cys Pro Tyr Leu Lys Phe Ala
 130 135 140
 His Phe Thr Ala Asn Gln Ala Ile Leu Glu Ala Phe Glu Gly Lys Lys
 145 150 155 160
 Arg Val His Val Ile Asp Phe Ser Met Asn Gln Gly Leu Gln Trp Pro
 165 170 175
 Ala Leu Met Gln Ala Leu Ala Leu Arg Glu Gly Gly Pro Pro Thr Phe
 180 185 190
 Arg Leu Thr Gly Ile Gly Pro Pro Ala Pro Asp Asn Ser Asp His Leu
 195 200 205
 His Glu Val Gly Cys Lys Leu Ala Gln Leu Ala Glu Ala Ile His Val
 210 215 220
 Glu Phe Glu Tyr Arg Gly Phe Val Ala Asn Ser Leu Ala Asp Leu Asp
 225 230 235 240
 Ala Ser Met Leu Glu Leu Arg Pro Ser Asp Thr Glu Ala Val Ala Val

	245		250		255
Asn Ser Val Phe Glu Leu His Lys Leu Leu Gly Arg Xaa Gly Gly Ile					
	260		265		270
Glu Lys Val Leu Gly					
	275				

(2) INFORMATION FOR SEQ ID NO:35:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 262 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

Gly	Gly	Gly	Gly	Asp	Thr	Tyr	Thr	Thr	Asn	Lys	Arg	Leu	Lys	Cys	Ser	1	5	10	15
Asn	Gly	Val	Val	Glu	Thr	Thr	Thr	Ala	Thr	Ala	Glu	Ser	Thr	Arg	His	20	25	30	
Val	Val	Leu	Val	Asp	Ser	Gln	Glu	Asn	Gly	Val	Arg	Leu	Val	His	Ala	35	40	45	
Leu	Leu	Ala	Cys	Ala	Glu	Ala	Val	Gln	Lys	Glu	Asn	Leu	Thr	Val	Ala	50	55	60	
Glu	Ala	Leu	Val	Lys	Gln	Ile	Gly	Phe	Leu	Ala	Val	Ser	Gln	Ile	Gly	65	70	75	80
Ala	Met	Arg	Gln	Val	Ala	Thr	Tyr	Phe	Ala	Glu	Ala	Leu	Ala	Arg	Arg	85	90	95	
Ile	Tyr	Arg	Leu	Ser	Pro	Ser	Gln	Ser	Pro	Ile	Asp	His	Ser	Leu	Ser	100	105	110	
Asp	Thr	Leu	Gln	Met	His	Phe	Tyr	Glu	Thr	Cys	Pro	Tyr	Leu	Lys	Phe	115	120	125	
Ala	His	Phe	Thr	Ala	Asn	Gln	Ala	Ile	Leu	Glu	Ala	Phe	Gln	Gly	Lys	130	135	140	
Lys	Arg	Val	His	Val	Ile	Asp	Phe	Ser	Met	Ser	Gln	Gly	Leu	Gln	Trp	145	150	155	160
Pro	Ala	Leu	Met	Gln	Ala	Leu	Ala	Leu	Arg	Pro	Gly	Gly	Pro	Pro	Val	165	170	175	
Phe	Arg	Leu	Thr	Gly	Ile	Gly	Pro	Pro	Ala	Pro	Asp	Asn	Phe	Asp	Tyr	180	185	190	
Leu	His	Glu	Val	Gly	Cys	Lys	Leu	Ala	His	Leu	Ala	Glu	Ala	Ile	His	195	200	205	
Val	Glu	Phe	Glu	Tyr	Arg	Gly	Phe	Val	Ala	Asn	Thr	Leu	Ala	Asp	Leu	210	215	220	
Asp	Ala	Ser	Met	Leu	Glu	Leu	Arg	Pro	Ser	Glu	Ile	Glu	Ser	Val	Ala	225	230	235	240


```
Val Asn Ser Val Phe Glu Leu His Lys Leu Leu Gly Arg Pro Gly Ala
                245                      250                      255

Ile Asp Lys Val Leu Gly
                260
```

(2) INFORMATION FOR SEQ ID NO:36:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 203 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(x1) SEQUENCE DESCRIPTION: SEQ ID NO:36:

Gln 1	Leu	Gly	Lys 5	Pro	Phe	Leu	Arg	Ser	Ala 10	Ser	Tyr	Leu	Lys	Glu 15	Ala
Leu	Leu	Leu	Ala 20	Leu	Ala	Asp	Ser	His 25	His	Gly	Ser	Ser	Gly 30	Val	Thr
Ser	Pro	Leu 35	Asp	Val	Ala	Leu	Lys 40	Leu	Ala	Ala	Tyr	Lys 45	Ser	Phe	Ser
Asp	Leu 50	Ser	Pro	Val	Leu	Gln 55	Phe	Thr	Asn	Phe	Thr 60	Ala	Asn	Lys	Ala
Leu 65	Leu	Asp	Glu	Ile	Gly 70	Gly	Met	Ala	Thr	Ser 75	Cys	Ile	His	Val	Ile 80
Asp	Phe	Asn	Leu	Gly 85	Val	Gly	Gly	Gln	Trp 90	Ala	Ser	Phe	Leu	Gln 95	Glu
Leu	Ala	His	Arg 100	Arg	Gly	Ala	Gly	Gly 105	Met	Ala	Leu	Pro	Leu 110	Leu	Lys
Leu	Thr	Ala 115	Phe	Met	Ser	Thr	Ala 120	Ser	His	His	Pro	Leu 125	Glu	Leu	His
Leu	Thr 130	Gln	Asp	Asn	Leu	Ser 135	Gln	Phe	Ala	Ala	Glu 140	Leu	Arg	Ile	Pro
Phe 145	Glu	Phe	Asn	Ala	Val 150	Ser	Leu	Asp	Ala	Phe 155	Asn	Pro	Ala	Glu	Ser 160
Ile	Ser	Ser	Ser	Gly 165	Asp	Glu	Val	Val	Ala 170	Val	Ser	Leu	Pro	Val 175	Gly
Cys	Ser	Ala	Arg 180	Ala	Pro	Pro	Leu	Pro 185	Ala	Ile	Leu	Arg	Leu 190	Val	Lys
Gln	Leu	Cys 195	Pro	Lys	Val	Val	Val	Ala 200	Ile	Asp					

(2) INFORMATION FOR SEQ ID NO:37:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 131 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

```

His Ala Ser Val Lys Gly Tyr Asn His Val His Ile Ile Asp Phe Ser
1          5          10          15
Leu Met Gln Gly Leu Gln Trp Pro Ala Leu Met Asp Val Phe Ser Ala
20          25          30
Arg Glu Gly Gly Pro Pro Lys Leu Arg Ile Thr Gly Ile Gly Pro Asn
35          40          45
Pro Ile Gly Gly Arg Asp Glu Leu His Glu Val Gly Ile Arg Leu Ala
50          55          60
Lys Tyr Ala His Ser Val Gly Ile Asp Phe Thr Phe Gln Gly Val Cys
65          70          75          80
Val Asp Gln Leu Asp Arg Leu Cys Asp Trp Met Leu Leu Lys Pro Ile
85          90          95
Lys Gly Glu Ala Val Ala Ile Asn Ser Ile Leu Gln Leu His Arg Leu
100         105         110
Leu Val Asp Pro Asp Ala Asn Pro Val Val Pro Ala Pro Ile Asp Ile
115         120         125
Leu Leu Lys
130

```

(2) INFORMATION FOR SEQ ID NO:38:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

```

Gln Glu Ala Phe Glu Arg Glu Glu Arg Val His Ile Ile Asp Leu Asp
1          5          10          15
Ile Met Gln Gly Leu Gln Trp Pro Gly Leu Phe His Ile Leu Ala Ser
20          25          30
Arg

```

(2) INFORMATION FOR SEQ ID NO:39:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 29 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

Phe Ala Gly Cys Arg Arg Val His Val Val Asp Phe Gly Ile Lys Gln
 1 5 10 15
 Gly Met Gln Trp Pro Ala Leu Leu Xaa Asp Leu Ala Leu
 20 25

(2) INFORMATION FOR SEQ ID NO:40:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 73 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

Gly Arg Asn Gly Arg Thr Leu Trp Leu Gly Glu Gly His Ile Asp Leu
 1 5 10 15
 Trp Pro Leu Gln Gly Leu Leu Ser Gln Gly Leu Gln Arg Ala Leu Cys
 20 25 30
 Ala Arg Pro Leu Gly Ala Pro His Val Phe Leu Pro Gly Leu His Thr
 35 40 45
 Leu Ser Leu Gly Leu Gln Xaa Arg His Leu Leu Val His Met Met Ala
 50 55 60
 Leu Ser Tyr Ser Tyr Gly Arg Xaa Pro
 65 70

(2) INFORMATION FOR SEQ ID NO:41:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 59 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

Thr Ser Asp Ser Ala Ser Ser Phe Asn Ile Pro Thr Ser Ala Gln Asn
 1 5 10 15
 His Tyr Ala Thr Gly Ser Phe Ser Thr Asn Ser Arg Thr Thr Asn Val
 20 25 30
 Ala Thr Ala Thr Thr Asn Ser Ala Thr Ala His Trp Val Ala Thr Asp
 35 40 45
 Ala Glu His Thr Asp Thr Ile Ile Ala Gln Pro
 50 55

(2) INFORMATION FOR SEQ ID NO:42:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 110 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

```

Arg Xaa Phe Asp Ser Leu Glu His Asp Ala Ser Lys Gly Glu Pro Arg
1           5           10           15
Glu Asp Glu Arg Gly Arg Xaa Cys Leu Ala Arg Asn Ile Val Asn Ile
20           25           30
Val Xaa Cys Lys Xaa Glu Glu Arg Ile Glu Arg Tyr Glu Val Thr Gly
35           40           45
Lys Trp Arg Ala Arg Met Met Met Ala Gly Phe Ser Pro Arg Pro Met
50           55           60
Ser Gly Arg Val Thr Ser Asn Ile Glu Ser Leu Ile Lys Arg Asp Tyr
65           70           75           80
Cys Ser Lys Tyr Lys Val Lys Glu Glu Met Gly Glu Leu His Phe Ser
85           90           95
Trp Glu Glu Lys Ser Leu Ile Val Ala Ser Ala Trp Ser Xaa
100          105          110

```

(2) INFORMATION FOR SEQ ID NO:43:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 137 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

```

Asn Gly Ser Tyr Asn Ala Pro Phe Phe Val Thr Arg Phe Arg Glu Ala
1           5           10           15
Leu Phe His Tyr Ser Ala Ile Phe Asp Met Leu Glu Thr Asn Ile Pro
20           25           30
Lys Asp Asn Glu Gln Arg Leu Leu Ile Glu Ser Ala Leu Phe Ser Arg
35           40           45
Glu Xaa Asn Val Ile Ser Cys Glu Gly Leu Glu Arg Met Glu Arg Pro
50           55           60
Glu Thr Tyr Lys Gln Trp Gln Val Arg Asn Gln Arg Val Gly Phe Lys
65           70           75           80
Gln Leu Pro Leu Asn Gln Asp Met Met Lys Arg Ala Arg Xaa Glu Gly

```

85	90	95
Gln Val Leu Pro Thr Arg Thr Phe Ile Ile Asp Glu Asp Asn Arg Trp		
100	105	110
Leu Leu Gln Gly Trp Lys Gly Arg Ile Leu Phe Ala Leu Ser Thr Trp		
115	120	125
Lys Pro Asp Asn Arg Ser Ser Ser Xaa		
130	135	

(2) INFORMATION FOR SEQ ID NO:44:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 41 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

Asn Gly Gly Ala Phe Ala Pro Ser Thr Trp Thr Ala Arg Ser Leu Asn	
1	15
Gly Gly Ala Phe Ala Pro Ser Thr Trp Thr Ala Arg Ser Leu Pro Val	
20	30
Pro Ser Ser Pro Ser Thr Asp Ser Phe	
35	40

(2) INFORMATION FOR SEQ ID NO:45:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1279 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

GCGGCTATCT TCTACGGCCA CCACCACCAT ACACCTCCGC CGGCAAAGCG GCTCAACCCT	60
GGTCCCCTGG GGATAACAGA GCAGCTGGTT AAGGCAGCAG AGGTCATAGA GAGCGACACG	120
TGTCTAGCTC AGGGGATATT GCGCGGGCTC AATCAACAGC TCTCTTCTCC CGTCGGGAAG	180
CCATTAGAAA GAGCAGCTTT TTA CTTC AAA GAAGCTCTCA ATAATCTCCT TCACAACGTC	240
TCCCAAACCC TAAACCCTTA TTCCCTCATC TTCAAGATCG CTGCTTACAA ATCCTTCTCA	300
GAGATCTCTC CCGTTCTTCA GTTCGCCAAC TTTACCTCCA ACCAAGCCCT CTTAGAGTCC	360
TTCCATGGCT TCCACCGTCT CCACATCATC GACTTCGATA TCGGCTACGG TGGCCAATGG	420
GCTTCCCTCA TGCAAGAGCT TGTCTCCGC GACAAAGCCG CTCCTCTCTC CCTCAAGATC	480
ACCGTTTTCG CTTCTCCGGC GAACCACGAC CAGCTCGAAC TTGGCTTCAC TCAAGACAAC	540

```

CTCAAGCACT TCGCCTCTGA GATCAACATC TCCCTTGACA TCCAAGTTTT GAGCTTAGAC      600
CTCCTCGGCT CCATCTCGTG GCCTAACTCG TCGGAGAAAG AAGCTGTCCG CGTTAACATC      660
TCCGCCGCGT CCTTCTCGCA CCTCCCTTTG GTCCTCOGTT TCGTGAAGCA TCTATCTCCG      720
ACGATCATCG TCTGCTCCGA CAGAGGATGC GAGAGGACGG ATCTGCCCTT CTCTCAACAG      780
CTCGCCCACT CGCTGCACTC ACACACCGCT CTCTTCGAAT CCCTCGACGC CGTCAACGCC      840
AACCTCGACG CAATGCAGAA GATCGAGAGG TTTCTTATAC AGCCGGAGAT AGAGAAGCTG      900
GTGTTGGATC GTAGCCGTCC GATAGAAAGG CCGATGATGA CGTGGCAAGC GATGTTTCTA      960
CAGATGGGTT TCTACCCGGT GACGCACAGT AACTTCACGG AGTCTCAAGC CGAGTGTTTA     1020
GTCCAACGGA CGCCAGTGAG AGGCTTTCAC GTCGAGAAGA AACATAACTC ACTTCTCCTA     1080
TGTTGGCAAA GGACAGAACT CGTCGGAGTT TCAGCATGGA GATGTCGCTC CTCCTGATTT     1140
CCACCGGAGT TTCAATTATT AAAAAAATAT TTTCCCTAAT TCAATTTATC TTAAATGACA     1200
AATTTTTAGT TTCTGATTTT ATTTTGCTCA GTGCGATGGA TTTTAAATT TAAGTTTCAC     1260
ACAAATATAT AAATTTTTG                                     1279

```

(2) INFORMATION FOR SEQ ID NO:46:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 379 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:

```

Ala Ala Ile Phe Tyr Gly His His His His Thr Pro Pro Pro Ala Lys
 1              5              10              15
Arg Leu Asn Pro Gly Pro Val Gly Ile Thr Glu Gln Leu Val Lys Ala
 20              25              30
Ala Glu Val Ile Glu Ser Asp Thr Cys Leu Ala Gln Gly Ile Leu Ala
 35              40              45
Arg Leu Asn Gln Gln Leu Ser Ser Pro Val Gly Lys Pro Leu Glu Arg
 50              55              60
Ala Ala Phe Tyr Phe Lys Glu Ala Leu Asn Asn Leu Leu His Asn Val
 65              70              75              80
Ser Gln Thr Leu Asn Pro Tyr Ser Leu Ile Phe Lys Ile Ala Ala Tyr
 85              90              95
Lys Ser Phe Ser Glu Ile Ser Pro Val Leu Gln Phe Ala Asn Phe Thr
 100             105             110
Ser Asn Gln Ala Leu Leu Glu Ser Phe His Gly Phe His Arg Leu His
 115             120             125
Ile Ile Asp Phe Asp Ile Gly Tyr Gly Gly Gln Trp Ala Ser Leu Met
 130             135             140

```

Gln Glu Leu Val Leu Arg Asp Asn Ala Ala Pro Leu Ser Leu Lys Ile
 145 150 155 160
 Thr Val Phe Ala Ser Pro Ala Asn His Asp Gln Leu Glu Leu Gly Phe
 165 170 175
 Thr Gln Asp Asn Leu Lys His Phe Ala Ser Glu Ile Asn Ile Ser Leu
 180 185 190
 Asp Ile Gln Val Leu Ser Leu Asp Leu Leu Gly Ser Ile Ser Trp Pro
 195 200 205
 Asn Ser Ser Glu Lys Glu Ala Val Ala Val Asn Ile Ser Ala Ala Ser
 210 215 220
 Phe Ser His Leu Pro Leu Val Leu Arg Phe Val Lys His Leu Ser Pro
 225 230 235 240
 Thr Ile Ile Val Cys Ser Asp Arg Gly Cys Glu Arg Thr Asp Leu Pro
 245 250 255
 Phe Ser Gln Gln Leu Ala His Ser Leu His Ser His Thr Ala Leu Phe
 260 265 270
 Glu Ser Leu Asp Ala Val Asn Ala Asn Leu Asp Ala Met Gln Lys Ile
 275 280 285
 Glu Arg Phe Leu Ile Gln Pro Glu Ile Glu Lys Leu Val Leu Asp Arg
 290 295 300
 Ser Arg Pro Ile Glu Arg Pro Met Met Thr Trp Gln Ala Met Phe Leu
 305 310 315 320
 Gln Met Gly Phe Ser Pro Val Thr His Ser Asn Phe Thr Glu Ser Gln
 325 330 335
 Ala Glu Cys Leu Val Gln Arg Thr Pro Val Arg Gly Phe His Val Glu
 340 345 350
 Lys Lys His Asn Ser Leu Leu Leu Cys Trp Gln Arg Thr Glu Leu Val
 355 360 365
 Gly Val Ser Ala Trp Arg Cys Arg Ser Ser Xaa
 370 375

(2) INFORMATION FOR SEQ ID NO:47:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 745 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:

TGCATACAAC GCACCGTTTT TCGTAACACG GTTTCGCGAA GCTCTATTTC ATTTCTCCTC	60
GATTTTGTGAC ATGCTTGAGA CAATTGTGCC ACGAGAAGAC GAAGAGAGGA TGTTCCCTGA	120
GATGGAGGTC TTTGGGAGAG AGGCACTGAA TGTGATTGCT TGCGAAGGTT GGGAAAGAGT	180
GGAGAGGCCT GAGACATACA AGCAGTGGCA CGTACGGGCT ATGAGGTCAG GCTTGGTGCA	240

```

GGTTCATTT GACCCAAGCA TTATGAAGAC ATCGCTGCAT AAGGTCCACA CATTCTACCA      300
CAAGGATTTT GTGATCGATC AAGATAACCG GTGGCTCTTG CAAGGCTGGA AGGGAAGAAC      360
TGTCATGGCT CTTTCTGTTT GGAAACCAGA GTCCAAGGCT TGACCGAGAA ATCCTCGTTG      420
GCATATGAGA GACCATCTCT TGATTTTCTT CCTGTGTAAT TCCCAGAGAC AGAATTACAG      480
ATGTAAGAAG AGAATGCTGC ACAAAGAACT TGTTCAAAGA TAATATTGAT GTAAGTCCTG      540
TTTTATAACT TTCTAGCTGT GTTTTGTGTT TTTCTCAGCT AGATTCTCCT AACGGTATTC      600
TTGTAGCTAG GGTGATCAGA TTGTTTGTAT ATTGCTAGCA GAGTTAGTTT GTCTAGATTG      660
TAACACATAT AAGAGGAAGC TTAGAGTTTC TATGGTTTAA AGAGAAGTTT TTTCTTCTC      720
CAATGTAAAA AAAAAAAAAA AAAAAA      745

```

(2) INFORMATION FOR SEQ ID NO:48:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 134 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:

```

Ala Tyr Asn Ala Pro Phe Phe Val Thr Arg Phe Arg Glu Ala Leu Phe
1          5          10          15
His Phe Ser Ser Ile Phe Asp Met Leu Glu Thr Ile Val Pro Arg Glu
20          25          30
Asp Glu Glu Arg Met Phe Leu Glu Met Glu Val Phe Gly Arg Glu Ala
35          40          45
Leu Asn Val Ile Ala Cys Glu Gly Trp Glu Arg Val Glu Arg Pro Glu
50          55          60
Thr Tyr Lys Gln Trp His Val Arg Ala Met Arg Ser Gly Leu Val Gln
65          70          75          80
Val Pro Phe Asp Pro Ser Ile Met Lys Thr Ser Leu His Lys Val His
85          90          95
Thr Phe Tyr His Lys Asp Phe Val Ile Asp Gln Asp Asn Arg Trp Leu
100          105          110
Leu Gln Gly Trp Lys Gly Arg Thr Val Met Ala Leu Ser Val Trp Lys
115          120          125
Pro Glu Ser Lys Ala Xaa
130

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(2) INFORMATION FOR SEQ ID NO:49:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 775 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:

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AAAAAATGGG AAACCATCAC TCTTGATGAA CTTATGATCA ATCCAGGAGA GACAACGGTC      60
GTCAACTGCA TTCATCGGTT ACAATACACT CCTGATGAAA CTGTGTCATT AGACTCTCCA      120
AGAGACACGG TTCTGAAGCT ATTCAGAGAT ATCAATCCTG ACCTCTTTGT GTTTGCAGAG      180
ATTAACGGAA TGTACAACTC TCCTTTCTTC ATGACGAGGT TCCGAGAAGC GCTTTTTCAT      240
TACTCTTCAC TCTTTGACAT GTTTGACACC ACAATACACG CAGAGGATGA GTACAAAAAC      300
AGGTCACGTG TGGAGAGAGA GTTACTTGTG AGAGACGCCA TGAGCGTGAT TTCCTGCGAG      360
GGTGCAGAGC GGTTTGCGAG GCCTGAAACC TACAAGCAAT GGCGAGTTAG GATTTTGAGA      420
GCCGGGTTTA AGCCAGCAAC TATTAGCAAA CAGATCATGA AGGAGGCTAA GGAAATTGTG      480
AGGAAACGTT ACCATAGAGA TTTTGTGATC GATAGCGATA ACAATTGGAT GCTTCAAGGA      540
TGGAAGGAA GAGTCATCTA TGCTTTTTCT TGCTGGAAAC CTGCTGAGAA GTTCACAAAC      600
AATAATTTAA ACATCTGAAA AATGTTACTT CTCAATTACA TCATTTTGTG TTCCCAATGG      660
TTTTGTAGAA TATGTTTGAT CCCGTGAGTG GATGCAACTC TTTTTCCTG CAAGTACATA      720
TTGTATTCAA ATCCTTGTGG AAATGATAAA TTGTTTAATC AAAAAAAAAA AAAAA      775

```

(2) INFORMATION FOR SEQ ID NO:50:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 206 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:50:

```

Lys Lys Trp Glu Thr Ile Thr Leu Asp Glu Leu Met Ile Asn Pro Gly
 1           5           10           15
Glu Thr Thr Val Val Asn Cys Ile His Arg Leu Gln Tyr Thr Pro Asp
          20           25           30
Glu Thr Val Ser Leu Asp Ser Pro Arg Asp Thr Val Leu Lys Leu Phe
          35           40           45
Arg Asp Ile Asn Pro Asp Leu Phe Val Phe Ala Glu Ile Asn Gly Met
          50           55           60
Tyr Asn Ser Pro Phe Phe Met Thr Arg Phe Arg Glu Ala Leu Phe His
          65           70           75           80
Tyr Ser Ser Leu Phe Asp Met Phe Asp Thr Thr Ile His Ala Glu Asp
          85           90           95
Glu Tyr Lys Asn Arg Ser Leu Leu Glu Arg Glu Leu Leu Val Arg Asp

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100	105	110
Ala Met Ser Val Ile Ser Cys	Glu Gly Ala Glu Arg Phe	Ala Arg Pro
115	120	125
Glu Thr Tyr Lys Gln Trp Arg	Val Arg Ile Leu Arg Ala	Gly Phe Lys
130	135	140
Pro Ala Thr Ile Ser Lys Gln Ile Met Lys	Glu Ala Lys Glu Ile Val	
145	150	155
Arg Lys Arg Tyr His Arg Asp Phe Val Ile Asp Ser Asp Asn Asn Trp		
165	170	175
Met Leu Gln Gly Trp Lys Gly Arg Val Ile Tyr Ala Phe Ser Cys Trp		
180	185	190
Lys Pro Ala Glu Lys Phe Thr Asn Asn Asn Leu Asn Ile Xaa		
195	200	205

(2) INFORMATION FOR SEQ ID NO:51:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 548 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: unknown
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:51:

AATCGCTTGA ACCGAATTTG GATCGAGATT CGAAAGAAAG GCTGAGAGTG GAGAGAGTGC	60
TGTTCCGGTAG GAGGATTATG GATTTGGTCC GATCAGATGA TGATAATAAT AAACCGGGAA	120
CCCGGTTTGG GTTAATCGAG GAGAAAGAAC AATGGAGAGT GTTGATGGAG AAAGCTGGAT	180
TTGAGCCGGT TAAACCGAGT AATTACGCGG TTAGCCAAGC GAAGCTGCTA CTATGGAAct	240
ACAATTATAG TACATTGTAT TCACTTGTTG AATCGGAGCC AGGTTTCATC TCCTTGGCTT	300
GGAACAATGT GCCTCTCCTC ACCGTTTCCT CTTGGCGTTG ACTACTTGGT CCGATAAGTT	360
AATCTAGTAT TTTGAGTTAG CTTTtagAAT TGAATTGTTT GCGGTTAGAT TTGGATGTTT	420
AATTAGTCTC TAGCCTATTC TCTTACTCTT TTTTGTCTAG TGCTTGGAGT GATGATGGTT	480
TGTCGTTTAT GTTCATTTGT AATATATATT GTATGTAACA TTTGACTAAA AAAAAAAAAA	540
AAAAAAAA	548

(2) INFORMATION FOR SEQ ID NO:52:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 113 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:

Ser Leu Glu Pro Asn Leu Asp Arg Asp Ser Lys Glu Arg Leu Arg Val
 1 5 10 15
 Glu Arg Val Leu Phe Gly Arg Arg Ile Met Asp Leu Val Arg Ser Asp
 20 25 30
 Asp Asp Asn Asn Lys Pro Gly Thr Arg Phe Gly Leu Met Glu Glu Lys
 35 40 45
 Glu Gln Trp Arg Val Leu Met Glu Lys Ala Gly Phe Glu Pro Val Lys
 50 55 60
 Pro Ser Asn Tyr Ala Val Ser Gln Ala Lys Leu Leu Leu Trp Asn Tyr
 65 70 75 80
 Asn Tyr Ser Thr Leu Tyr Ser Leu Val Glu Ser Glu Pro Gly Phe Ile
 85 90 95
 Ser Leu Ala Trp Asn Asn Val Pro Leu Leu Thr Val Ser Ser Trp Arg
 100 105 110
 Xaa

(2) INFORMATION FOR SEQ ID NO:53:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1093 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:53:

CGCAATGTTG AGATCTTGGA AGCAATAGCT GGGGAAACCA GAGTCCACAT TATCGATTTT 60
 CAGATTGCAC AGGGATCACA ATACATGTTT TTGATTTCAGG AGCTTGCGAA ACGCCCTGGT 120
 GGGCCGCCGT TGCTGCGTGT GACGGGTGTG GATGATTCAC AGTCCACCTA TGCTCGTGGG 180
 GGAGGACTCA GCTTGGTAGG TGAGAGGCTT GCAACTTTGG CGCAGTCATG TGGTGTCCCCG 240
 TTTGAGTTTC ACGATGCCAT CATGTCTGGG TGCAAGGTGC AGCGGGAACA TCTCGGGTTG 300
 GAACCTGGCT TTGCTGTTGT TGTGAACTTC CCATATGTAT TACACCACAT GCCAGACGAG 360
 AGCGTAAGTG TTGAAAATA CAGAGACAGG CTGCTGCATC TGATCAAGAG CCTCTCCCCA 420
 AAACCTGGTTA CTCTAGTAGA GCAAGAATCC AACACAAACA CCTCGCCATT GGTGTCACGG 480
 TTTGTGGAAA CACTGGATTA CTACACAGCG ATGTTTGAGT CGATAGATGC AGCACGGCCA 540
 CGGGATGATA AGCAGAGAAT CAGCGCAGAA CAACACTGTG TAGCAAGAGA CATAGTGAAC 600
 ATGATAGCAT GTGAGGAGTC AGAGAGAGTA GAGAGACACG AGGTACTGGG GAAATGGAGG 660
 GTCAGAATGA TGATGGCTCG GTTCACGGGT TGGCCGGTCA GCACATCTGC AGCGTTTGCA 720
 GCGAGTGAGA TGCTGAAAGC TTATGACAAA AACTACAAAC TGGGAGGCCA TGAAGGAGCG 780

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CTCTACCTCT TCTGGAAGAG ACGACCCATG GCTACATGTT CCGTGTGGAA GCCAAACCCA      840
AACTATATTG GGTAAGTTAT AGTGATGATG GTTACTTGAG TGGATAAAGA AGAGCACAAAC      900
AAAAACACAT CTGTCGCTGT AAATTTTTTA GGATGTGCAA TGATGTTTTA AGTTGTAACA      960
CAACCTAAGT TATATATGTA TACAAACCAA ACCTGGTGGT TGTTCCTCTC TTGTAAATTG     1020
TCATGTGGTT GTGGGTGGGA AGCTAGTAAT GAAATATAAC CAAACATTG ATTAGGTCAA     1080
AAAAAAAAAA AAA                                                              1093

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(2) INFORMATION FOR SEQ ID NO:54:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 285 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:54:

```

Ala Asn Val Glu Ile Leu Glu Ala Ile Ala Gly Glu Thr Arg Val His
1           5           10
Ile Ile Asp Phe Gln Ile Ala Gln Gly Ser Gln Tyr Met Phe Leu Ile
20          25          30
Gln Glu Leu Ala Lys Arg Pro Gly Gly Pro Pro Leu Leu Arg Val Thr
35          40          45
Gly Val Asp Asp Ser Gln Ser Thr Tyr Ala Arg Gly Gly Gly Leu Ser
50          55          60
Leu Val Gly Glu Arg Leu Ala Thr Leu Ala Gln Ser Cys Gly Val Pro
65          70          75          80
Phe Glu Phe His Asp Ala Ile Met Ser Gly Cys Lys Val Gln Arg Glu
85          90          95
His Leu Gly Leu Glu Pro Gly Phe Ala Val Val Val Asn Phe Pro Tyr
100         105         110
Val Leu His His Met Pro Asp Glu Ser Val Ser Val Glu Lys Tyr Arg
115        120        125
Asp Arg Leu Leu His Leu Ile Lys Ser Leu Ser Pro Lys Leu Val Thr
130        135        140
Leu Val Glu Gln Glu Ser Asn Thr Asn Thr Ser Pro Leu Val Ser Arg
145        150        155        160
Phe Val Glu Thr Leu Asp Tyr Tyr Thr Ala Met Phe Glu Ser Ile Asp
165        170        175
Ala Ala Arg Pro Arg Asp Asp Lys Gln Arg Ile Ser Ala Glu Gln His
180        185        190
Cys Val Ala Arg Asp Ile Val Asn Met Ile Ala Cys Glu Glu Ser Glu
195        200        205
Arg Val Glu Arg His Glu Val Leu Gly Lys Trp Arg Val Arg Met Met

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210	215	220
Met Ala Gly Phe Thr Gly Trp Pro Val Ser Thr Ser Ala Ala Phe Ala		
225	230	235 240
Ala Ser Glu Met Leu Lys Ala Tyr Asp Lys Asn Tyr Lys Leu Gly Gly		
	245	250 255
His Glu Gly Ala Leu Tyr Leu Phe Trp Lys Arg Arg Pro Met Ala Thr		
	260	265 270
Cys Ser Val Trp Lys Pro Asn Pro Asn Tyr Ile Gly Xaa		
	275	280 285

(2) INFORMATION FOR SEQ ID NO:55:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1928 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:55:

AAAGACTTTA GCAGATTTTC AAGCGGCTCA GAACATCAAC AACACAACA ACAACAACCG	60
TTTATAGTC AAGCAGCTCT CAACGCTTTT CTTTCAAGGT CTGTGAAGCC TCGAAATTAT	120
CAGAATTTTC AATCTCCGTC GGCCGATGAT TGATCTCAGC TCGGTGAATG ATATGAGTTT	180
GTTTGGTGGT TCTGGTTCAT CTCAGCGTTA CGGTTTACCG GTTCCCAGGT CTCAGACGCA	240
ACAGCAACAA TCGGATTACG GTTTATTTGG TGGGATCCGA ATGGGAATCG GGTCCGGTAT	300
TAATAATTAT CCAACATTAA CCGGCGTTCC GTGTATTGAA CCGGTTCAAA ACCGGGTTCA	360
TGAATCGGAG AACATGTTGA ATAGTTTAAG AGAGCTTGAG AAACAGCTTT TAGATGATGA	420
CGATGAGAGT GGTGGTGATG ATGACGTGTC AGTTATAACA AATTCAAATT CCGATTGGAT	480
TCAAAATCTC GTGACTCCGA ACCCGAACCC GAACCCGGTT TTGTCTTTTT CACCGAGCTC	540
TTCTTCTTCG TCTTCTTCGC CTTCTACAGC TTCGACGACG ACATCGGTAT GTTCTAGGCA	600
AACGGTTATG GAAATCGCGA CGGCGATCGC GGAAGGGAAA ACAGAGATAG CGACGGAGAT	660
TTTGCGCGGT GTTCTCAAA CGCCTAATCT TGAGAGGAAT TCAGAGGAGA AGCTTGTTGA	720
TTTCATGGTG GCTGCGCTTC GATCGAGGAT AGCTTCTCCA GTGACGGAAT TGTATGGAA	780
GGAGCATTTA ATCTCGACTC AATTGCTCTA CGAGCTCTCT CCTTGTTTCA AACTCGGTTT	840
CGAGGCCGCG AATCTCGCCA TTCTCGACGC CGCCGATAAC AACGACGGTG GAATGATGAT	900
ACCGCACGTT ATCGATTTCT ATATCGGAGA AGGTGGACAA TACGTTAACC TTCTCCGTAC	960
ATTATCCACG CGCCGGAATG GTAAAAGTCA GAGTCAGAAT TCTCCGCTGG TTAAGATCAC	1020
CGCCGTGGCG AACACGTTT ACGGATGTTT AGTCGATGAC GGTGGAGAAG AGAGGTTAAA	1080
AGCCGTCGGA GATTTGTTGA GCCAACTCGG TGATCGACTC GGTATCTCCG TAAGTTTCAA	1140

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CGTGGTGACG AGTTTACGAC TCGGTGATCT GAATCGTGAA TCTCTCGGGT GTGATCCCGA      1200
CGAGACTTTG GCTGTGAACT TAGCTTTCAA GCTTTATCGT GTTCCCGACG AAAGCGTATG      1260
CACGGAGAAT CCAAGAGACG AACTTCTCCG GCGCGTGAAG GGACTTAAAC CGCGCGTGGT      1320
TACTCTAGTG GAGCAAGAAA TGAATTCGAA TACGGCGCCG TTTTATAGGA GAGTGAGTGA      1380
GTCATGCGCG TGTTACGGTG CGTTGCTTGA GTCGGTCGAG TCTACGGTTC CTAGTACGAA      1440
TTCCGACCGT GCCAAAGTTG AGGAAGGAAT TGGCCGGAAG CTAGTAAACG CCGTGCCGTG      1500
CGAAGGAATC GATCGTATAG AGCGGTGCGA GGTGTTCCGG AAATGGCGAA TCCGGATGAG      1560
CATGGCTGGG TTTGAGTTAA TGCCATTGAG TGAGAAGATA GCGGAGTCGA TGAAGAGTCG      1620
TGGAAACCGA GTCCACCCGG GCTTTACCGT TAAAGAAGAT AACGGAGGTG TGTGCTTTGG      1680
TTGGATGGGA CGGGCACTCA CTGTCGCATC CGCTTGGCGT TAACTTCACA CACTCTTTTT      1740
TTTCTTCTTA TTATTACCAT ATTATTATTA ATTTTCGAGA TTATTCTGAT ATTATTATCA      1800
TTGTGATTTT CCGTTTCGAA AAGTGTAAGA ATCTTATGTA ACAAAGAAAA AAAAAAGACT      1860
TTTATGTTTT TCTAATAATA AAAGAAAGAG TGATTGGGTT CAAAAAAAAA AAAAAAAAAA      1920
AAAAAAAAA                                     1928

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(2) INFORMATION FOR SEQ ID NO:56:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 524 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:56:

```

Asp Leu Thr Ser Val Asn Asp Met Ser Leu Phe Gly Gly Ser Gly Ser
1           5           10           15
Ser Gln Arg Tyr Gly Leu Pro Val Pro Arg Ser Gln Thr Gln Gln Gln
20          25          30
Gln Ser Asp Tyr Gly Leu Phe Gly Gly Ile Arg Met Gly Ile Gly Ser
35          40          45
Gly Ile Asn Asn Tyr Pro Thr Leu Thr Gly Val Pro Cys Ile Glu Pro
50          55          60
Val Gln Asn Arg Val His Glu Ser Glu Asn Met Leu Asn Ser Leu Arg
65          70          75          80
Glu Leu Glu Lys Gln Leu Leu Asp Asp Asp Asp Glu Ser Gly Gly Asp
85          90          95
Asp Asp Val Ser Val Ile Thr Asn Ser Asn Ser Asp Trp Ile Gln Asn
100         105         110
Leu Val Thr Pro Asn Pro Asn Pro Asn Pro Val Leu Ser Phe Ser Pro
115         120         125

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Ser Ser Ser Ser Ser Ser Ser Ser Pro Ser Thr Ala Ser Thr Thr Thr
 130 135 140
 Ser Val Cys Ser Arg Gln Thr Val Met Glu Ile Ala Thr Ala Ile Ala
 145 150 155 160
 Glu Gly Lys Thr Glu Ile Ala Thr Glu Ile Leu Ala Arg Val Ser Gln
 165 170 175
 Thr Pro Asn Leu Glu Arg Asn Ser Glu Glu Lys Leu Val Asp Phe Met
 180 185 190
 Val Ala Ala Leu Arg Ser Arg Ile Ala Ser Pro Val Thr Glu Leu Tyr
 195 200 205
 Gly Lys Glu His Leu Ile Ser Thr Gln Leu Leu Tyr Glu Leu Ser Pro
 210 215 220
 Cys Phe Lys Leu Gly Phe Glu Ala Ala Asn Leu Ala Ile Leu Asp Ala
 225 230 235 240
 Ala Asp Asn Asn Asp Gly Gly Met Met Ile Pro His Val Ile Asp Phe
 245 250 255
 Asp Ile Gly Glu Gly Gly Gln Tyr Val Asn Leu Leu Arg Thr Leu Ser
 260 265 270
 Thr Arg Arg Asn Gly Lys Ser Gln Ser Gln Asn Ser Pro Val Val Lys
 275 280 285
 Ile Thr Ala Val Ala Asn Asn Val Tyr Gly Cys Leu Val Asp Asp Gly
 290 295 300
 Gly Glu Glu Arg Leu Lys Ala Val Gly Asp Leu Leu Ser Gln Leu Gly
 305 310 315 320
 Asp Arg Leu Gly Ile Ser Val Ser Phe Asn Val Val Thr Ser Leu Arg
 325 330 335
 Leu Gly Asp Leu Asn Arg Glu Ser Leu Gly Cys Asp Pro Asp Glu Thr
 340 345 350
 Leu Ala Val Asn Leu Ala Phe Lys Leu Tyr Arg Val Pro Asp Glu Ser
 355 360 365
 Val Cys Thr Glu Asn Pro Arg Asp Glu Leu Leu Arg Arg Val Lys Gly
 370 375 380
 Leu Lys Pro Arg Val Val Thr Leu Val Glu Gln Glu Met Asn Ser Asn
 385 390 395 400
 Thr Ala Pro Phe Leu Gly Arg Val Ser Glu Ser Cys Ala Cys Tyr Gly
 405 410 415
 Ala Leu Leu Glu Ser Val Glu Ser Thr Val Pro Ser Thr Asn Ser Asp
 420 425 430
 Arg Ala Lys Val Glu Glu Gly Ile Gly Arg Lys Leu Val Asn Ala Val
 435 440 445
 Ala Cys Glu Gly Ile Asp Arg Ile Glu Arg Cys Glu Val Phe Gly Lys
 450 455 460
 Trp Arg Met Arg Met Ser Met Ala Gly Phe Glu Leu Met Pro Leu Ser
 465 470 475 480
 Glu Lys Ile Ala Glu Ser Met Lys Ser Arg Gly Asn Arg Val His Pro

	485	490	495
Gly Phe Thr Val Lys Glu Asp Asn Gly Gly Val Cys Phe Gly Trp Met			
500	505	510	
Gly Arg Ala Leu Thr Val Ala Ser Ala Trp Arg Xaa			
515	520		

(2) INFORMATION FOR SEQ ID NO:57:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2635 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: unknown
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:57:

TCCTACTCAA GGTTCCTTCTT TGTCATCTTG TTGCCGAATC CACAAAGAGG AGAATAAAGA	60
TTCGACCTTT ATTAGATATT AACGACTCTG GATTTTTGGG TTTTGGAGT TGGATCCACA	120
TGGGTTCTTA TCCGGATGGA TTCCCTGGAT CCATGGACGA GTTGGATTTC AATAAGGACT	180
TTGATTTGCC TCCCTCCTCA AACCAAACCT TAGGTTTAGC TAATGGGTTT TATTTAGATG	240
ACTTAGATTT CTCATCCTTG GATCCTCCAG AGGCATATCC CTCCCAGAAC AACACAACA	300
ACAACATCAA CAACAAAGCT GTAGCAGGAG ATCTGTTATC ATCTTCATCT GATGACGCTG	360
ATTTCTCTGA TTCTGTTTTG AAGTATATAA GCCAAGTTCT TATGGAAGAG GATATGGAAG	420
AGAAGCCTTG TATGTTTCAT GATGCTTTGG CTCTTCAAGC TGCTGAGAAA TCTCTCTATG	480
AGGCTCTTGG TGAGAAAGAC CCTTCTTCGT CTTCTGCTTC TTCTGTGGAT CATCCTGAGA	540
GATTGGCTAG TCATAGCCCT GACGGTTCTT GTTCAGGTGG TGCTTTTAGT GATTACGCTA	600
GCACCACTAC CACTACTTCC TCTGATTCTC ACTGGAGTGT TGATGGTTTG GAGAATAGAC	660
CTTCTTGGTT ACATACACCT ATGCCGAGTA ATTTTGT TTTT CCAGTCTACT TCTAGGTCCA	720
ACAGTGTCAC CGGTGGTGGT GGTGGTGGTA ATAGTGCGGT TTACGGTTCA GGTTTTGGCG	780
ATGATTTGGT TTCGAATATG TTTAAAGATG ATGAATTGGC TATGCAGTTC AAGAAAGGGG	840
TTGAGGAAGC TAGTAAGTTC CTTCTTAAGT CTTCTCAGCT CTTTATTGAT GTGGATAGTT	900
ACATCCCTAT GAATTCTGGT TCCAAGGAAA ATGGTTCTGA GGTTTTTGTT AAGACGGAGA	960
AGAAAGATGA GACAGAGCAT CATCATCATC ATAGCTATGC ACCACCACCC AACAGATTAA	1020
CTGGTAAGAA AAGCCATTGG CGCGACGAAG ATGAAGATTT CGTTGAAGAA AGAAGTAACA	1080
AGCAATCAGC TGTTTATGTT GAGGAAAGCG AGCTTTCTGA AATGTTTGAT AACATGTTCC	1140
TATGTGGCCC TGGGAAACCT GTATGCATTC TTAACCAGAA CTTTCCTACA GAATCCGCTA	1200
AAGTCGTGAC CGCACAGTCA AATGGAGCAA AGATTCTGGT GAAGAAATCA ACTTCTACTA	1260
GTCATAGTAA CGATTCTAAG AAAGAAACTG CTGATTTGAG GACTCTTTTG GTGTTATGTG	1320

CACAAGCTGT ATCAGTGGAT GATCGTAGAA CCGCCAACGT TTAGCTAAGG CAGATACGAG 1380
 AGCATTCTTC GCCTCTAGGC AATGGTTCAG AGCGGTTGGC TCATTATTTT GCAAATAGTC 1440
 TTGAAGCACG CTTAGCTGGG ACCGGTACAC AGATCTACAC CGCTTTATCT TCGAAGAAAA 1500
 CGTCTGCAGC AGACATGTTG AAGGCTTACC AGACATACAT GTCGGTCTGC CCTTTCAGA 1560
 AAGCTGCTAT CATATTTGCT AACCACAGCA TGATGCGTTT CACTGCAAAC GCCAACACGA 1620
 TCCACATAAT AGATTTTCGA ATATCTTACG GTTTTCAGTG GCCTGCTCTG ATTCATCGCC 1680
 TCTCGCTCAG CAGACCTGGT GGTTCGCCTA AGCTTCGAAT TACCGGTNNN NNNNNNNNNN 1740
 NNNNNNNNNN NNNNNNNNNN NNNGAGTTCA GGAGACAGGT CATCGCTTGG CTCGATACTG 1800
 TCAGCGACAC AATGTTCCGT TTGAGTACAA CGCAATTGCT CAGAAATGGG GAAACGATCC 1860
 AAGTCGAAGA CTTAAAGCTT CGACAAGGAG AGTATGTGGT TGTGAACTCT TTGTTCCGTT 1920
 TCAGGAACCT TCTAGATGAG ACCGTTCTGG TAAACAGCCC GAGAGATGCA GTTTTGAAGC 1980
 TGATAAGAAA AATAAACCCG AATGTCTTCA TTCCAGCGAT CTTAAGCGGG AATTACAACG 2040
 CGCCATTCTT TGTACGAGG TTCAGAGAAG CGTTGTTTCA TTA CTGGCT GTGTTTGATA 2100
 TGTGTGACTC GAAGCTAGCT AGGGAAGACG AGATGAGGCT GATGTATGTG TTTGAGTTTT 2160
 ATGGGAGAGA GATTGTGAAT GTTGTGGCTT CTGAAGGAAC AGAGAGAGTG GAGAGCCGAG 2220
 AGACATATAA GCAGTGGCAG GCGAGACTGA TCCGAGCCGG ATTTAGACAG CTTCCGCTTG 2280
 AGAAGGAACT GATGCAGAAT CTGAAGTTGA AAATCGAAAA CGGGTACGAT AAAA ACTTCG 2340
 ATGTTGATCA AAACGGTAAC TGTTACTTTC AAGGGTGGAA AGGTAGAATC GTGTATGCTT 2400
 CATCTCTATG GGTTCCTTCG TCTTCATAGA TGTTGTTTCT TACGTTCTAA GCGACTGGGA 2460
 TTTATGTAGG GCTTTTCTGT TGATAGTCTC TCGCCAACAC GAGTGGATTA AGTTCAGAGT 2520
 TAGGGTTCTT GAACACTAGA ATGTTGTTAT ATTATGCTTG TGACATAGCG TGTGTAAGAG 2580
 TGTAGCCTAA GAGATATAGT ACTCATTGCA TGATCTTTTG CTATATGTTN CATGT 2635

(2) INFORMATION FOR SEQ ID NO:58:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 809 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:58:

Leu Leu Lys Val Leu Leu Cys His Leu Val Ala Glu Ser Thr Lys Arg
 1 5 10 15
 Arg Ile Lys Ile Arg Pro Leu Leu Asp Ile Asn Asp Ser Gly Phe Leu
 20 25 30
 Gly Phe Trp Ser Trp Ile His Met Gly Ser Tyr Pro Asp Gly Phe Pro
 35 40 45

Gly Ser Met Asp Glu Leu Asp Phe Asn Lys Asp Phe Asp Leu Pro Pro
 50 55 60
 Ser Ser Asn Gln Thr Leu Gly Leu Ala Asn Gly Phe Tyr Leu Asp Asp
 65 70 75 80
 Leu Asp Phe Ser Ser Leu Asp Pro Pro Glu Ala Tyr Pro Ser Gln Asn
 85 90 95
 Asn Asn Asn Asn Asn Ile Asn Asn Lys Ala Val Ala Gly Asp Leu Leu
 100 105 110
 Ser Ser Ser Ser Asp Asp Ala Asp Phe Ser Asp Ser Val Leu Lys Tyr
 115 120 125
 Ile Ser Gln Val Leu Met Glu Glu Asp Met Glu Glu Lys Pro Cys Met
 130 135 140
 Phe His Asp Ala Leu Ala Leu Gln Ala Ala Glu Lys Ser Leu Tyr Glu
 145 150 155 160
 Ala Leu Gly Glu Lys Asp Pro Ser Ser Ser Ser Ala Ser Ser Val Asp
 165 170 175
 His Pro Glu Arg Leu Ala Ser His Ser Pro Asp Gly Ser Cys Ser Gly
 180 185 190
 Gly Ala Phe Ser Asp Tyr Ala Ser Thr Thr Thr Thr Thr Ser Ser Asp
 195 200 205
 Ser His Trp Ser Val Asp Gly Leu Glu Asn Arg Pro Ser Trp Leu His
 210 215 220
 Thr Pro Met Pro Ser Asn Phe Val Phe Gln Ser Thr Ser Arg Ser Asn
 225 230 235 240
 Ser Val Thr Gly Gly Gly Gly Gly Gly Asn Ser Ala Val Tyr Gly Ser
 245 250 255
 Gly Phe Gly Asp Asp Leu Val Ser Asn Met Phe Lys Asp Asp Glu Leu
 260 265 270
 Ala Met Gln Phe Lys Lys Gly Val Glu Glu Ala Ser Lys Phe Leu Pro
 275 280 285
 Lys Ser Ser Gln Leu Phe Ile Asp Val Asp Ser Tyr Ile Pro Met Asn
 290 295 300
 Ser Gly Ser Lys Glu Asn Gly Ser Glu Val Phe Val Lys Thr Glu Lys
 305 310 315 320
 Lys Asp Glu Thr Glu His His His His His Ser Tyr Ala Pro Pro Pro
 325 330 335
 Asn Arg Leu Thr Gly Lys Lys Ser His Trp Arg Asp Glu Asp Glu Asp
 340 345 350
 Phe Val Glu Glu Arg Ser Asn Lys Gln Ser Ala Val Tyr Val Glu Glu
 355 360 365
 Ser Glu Leu Ser Glu Met Phe Asp Asn Met Phe Leu Cys Gly Pro Gly
 370 375 380
 Lys Pro Val Cys Ile Leu Asn Gln Asn Phe Pro Thr Glu Ser Ala Lys
 385 390 395 400
 Val Val Thr Ala Gln Ser Asn Gly Ala Lys Ile Arg Gly Lys Lys Ser

- 133 -

Leu Lys Ile Glu Asn Gly Tyr Asp Lys Asn Phe Asp Val Asp Gln Asn
 770 775 780
 Gly Asn Trp Leu Leu Gln Gly Trp Lys Gly Arg Ile Val Tyr Ala Ser
 785 790 795 800
 Ser Leu Trp Val Pro Ser Ser Ser Xaa
 805

(2) INFORMATION FOR SEQ ID NO:59:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 90 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:59:

Gln Glu Ala Asp His Asn Lys Thr Gly Phe Leu Asp Arg Phe Thr Glu
 1 5 10 15
 Ala Leu Phe Tyr Tyr Ser Ala Val Phe Asp Ser Leu Asp Ala Ala Asn
 20 25 30
 Asn Asn Asn Asn Asn Asn Gln Arg Met Glu Ala Glu Tyr Leu Gln
 35 40 45
 Arg Glu Ile Cys Asp Ile Val Cys Gly Glu Gly Ala Ala Arg Xaa Glu
 50 55 60
 Arg His Glu Pro Leu Ser Arg Trp Arg Asp Arg Leu Thr Arg Ala Gly
 65 70 75 80
 Leu Ser Ala Val Pro Leu Gly Ser Asn Ala
 85 90

(2) INFORMATION FOR SEQ ID NO:60:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 199 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: unknown
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: DNA (genomic)

(vi) ORIGINAL SOURCE:
 (A) ORGANISM: Daucus carota

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:60:

TCTGCAGACA ATTTNAGGA GGCCAATACC ATGCTATTGG AAATTCAGA ACTGTCCACA 60
 CCTNNNNNNN NNNNNNNNNN NNNNNNNNNN NNGTACTTC TCAGAGGNAA TGTCGGNNAG 120
 ATTAGTTAGC TCCTGCTTAG GAATCTATGC TTCTCTCCN GCAACAGTGG TGCCTCCTCA 180
 TGGTCAGAAA GTGGCCTCA 199

(2) INFORMATION FOR SEQ ID NO:61:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 66 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

- (vi) ORIGINAL SOURCE:
 (A) ORGANISM: *Daucus carota*

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:61:

```

Ser Ala Asp Asn Phe Xaa Glu Ala Asn Thr Met Leu Leu Glu Ile Ser
1          5          10          15
Glu Leu Ser Thr Pro Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Tyr
20          25          30
Phe Ser Glu Xaa Met Ser Xaa Arg Leu Val Ser Ser Cys Leu Gly Ile
35          40          45
Tyr Ala Ser Leu Pro Ala Thr Val Val Pro Pro His Gly Gln Lys Val
50          55          60
Ala Ser
65

```

(2) INFORMATION FOR SEQ ID NO:62:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 321 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: unknown
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: DNA (genomic)

- (vi) ORIGINAL SOURCE:
 (A) ORGANISM: *Glycine max*

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:62:

```

TCAACTGAGA ATCTAGAAGA TGCCAACAAG ATGCTTCTGG AGATTTCTCA GTTATCAACA      60
CCGTTCNNCA CTTCAGCACA GCGTGTGGCA GCATATTTCT CAGAAGCCAT ATCAGCAAGG      120
TTGGTGAGTT CATGTCTAGG GATATACGCA ACTTTGCCAC ACACACACCA AAGCCACAAG      180
GTAGCTTCAG CTTTTCAAGT GTTCAATGGT ATTAGTCCTT TAGTGGAGTT CTCACACTTC      240
ACAGCAAACC AAGCAATTCA AGAAGCCTTC GAAAGAGAAG AGAGGGTGCA CATCATAGAT      300
CTTGATATAA TGCAAGGGTT G                                     321

```

(2) INFORMATION FOR SEQ ID NO:63:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 107 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single

(D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Glycine max

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:63:

```

Ser Thr Glu Asn Leu Glu Asp Ala Asn Lys Met Leu Leu Glu Ile Ser
1          5          10          15
Gln Leu Ser Thr Pro Phe Xaa Thr Ser Ala Gln Arg Val Ala Ala Tyr
          20          25          30
Phe Ser Glu Ala Ile Ser Ala Arg Leu Val Ser Ser Cys Leu Gly Ile
          35          40          45
Tyr Ala Thr Leu Pro His Thr His Gln Ser His Lys Val Ala Ser Ala
          50          55          60
Phe Gln Val Phe Asn Gly Ile Ser Pro Leu Val Glu Phe Ser His Phe
          65          70          75          80
Thr Ala Asn Gln Ala Ile Gln Glu Ala Phe Glu Arg Glu Glu Arg Val
          85          90          95
His Ile Ile Asp Leu Asp Ile Met Gln Gly Leu
          100          105

```

(2) INFORMATION FOR SEQ ID NO:64:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 195 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: unknown
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: DNA (genomic)

(vi) ORIGINAL SOURCE:

(A) ORGANISM: Picea

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:64:

```

TCTGCAGACA ACTTTGAAGA AGCCAATACA ATACTGCCTC AGATCACAGA ACTCTCCACC      60
CCCTATNGCA ACTCGGTGCA ACGAGTGGCT GCCTATNNNN NNNNNNNNNN NNNNNNNNNN      120
NNNNNNNNNN NNTGCATAGG AATGTATTCT CCTCTCCCTC CTATTCACAT GTCCCAGAGC      180
CAGAAAATTG TGAAT                                          195

```

(2) INFORMATION FOR SEQ ID NO:65:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 65 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

- (vi) ORIGINAL SOURCE:
(A) ORGANISM: Picea

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:65:

```

Ser Ala Asp Asn Phe Glu Glu Ala Asn Thr Ile Leu Pro Gln Ile Thr
 1           5           10           15
Glu Leu Ser Thr Pro Tyr Xaa Asn Ser Val Gln Arg Val Ala Ala Tyr
          20           25           30
Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Cys Ile Gly Met
          35           40           45
Tyr Ser Pro Leu Pro Pro Ile His Met Ser Gln Ser Gln Lys Ile Val
 50           55           60
Asn
65

```

(2) INFORMATION FOR SEQ ID NO:66:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 2151 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: unknown
 (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:66:

```

GATATCAGCA TCATCAATTT TAAATGTAAG TTGGCAAAAG ATCATGAGGG TTCTCATAGT      60
AATTTGGCCA CAAGGTATGA CACTGTCTCA ATTGAGCAAT CTAGTAGAGA AACTGATCCA      120
TCATATATTG CTCATATTGA AAGTGAAAAA GATATGCTCA AGAACCTAGT AGAGAAGCTA      180
AAAATTGAAA AATCTAGCTC TACTAGAAAA ATATGATAGG TTGCCTGTTT CTCATGAAAA      240
TTTATTAGAT AATCATATCA TGGCTAGATG TCGCTCATGA GGTGTGTTCTT GCTAGTTTAG      300
ATTCCTGTGG GCATTCATCT CTTTATAGATG CACTAACATG ATAGGAAGTT TCTAATCTGG      360
TGCTTCACAA TTCTGGTGAT TCATGCTTCC TTCATTGCAA TTGATATTGA TGCTTGATTC      420
ATGCTTCAGT CACTTTGTGC GTTTAATTGG TATTGTATGT ATCACTAGAT TGTAGCGTGT      480
CTGCAACTAG TGTTTCACCA TGTGGTTTTT TAGTATCATT CGTATTAGTT TCTAACTTTC      540
TATTGATATA TTAAAGTGAT AACTAGTTTT AGAAATATTC TCTTGTGCCA TTAATGCTAC      600
AACTTGTTTT TAGCGTGTAC GTTAGCATT TAATATTTCC TTATTATGAA AGCGGAAGAG      660
AAACGCGCCC AACCAGAGCA TCCACGTCGT CTCATTTTCC CTTTCATCGTT GGATCATAGA      720
TGAGCGGTCC ACGGTGAACT CCGTTTGCCT GCAAACCAC GTCCTCTACG CGCTGTTAAG      780
TAGCTTCTAG AACATCACG ATGTGTCCCG TCCATTCCCT TAGGAGGAGC CGGATCCGGC      840
GCCGCAGTCG CCCAAGGTCC CGACGCGCGC GGCCTCGGCC GCCGCCGCCA AGGAGCGGAA      900

```

```

GGAGGTGCAG CGGCGGAAGC AGCGCGACGA GGAGGGCCTC CACCTGCTGA GTGCTGACGC      960
TGCTGCTGCA GTGCGCGGAG GCCGTGAACG CGGACAACTT CGACGACGCG CACCAGACGC      1020
TGCTGGAGAT CGCGGAGCTG GCCACGCCGT TCGGCACCTC GACCCAGCGC GTGGCCGCCT      1080
ACTTCGCGGA GGCCATGTCG GCGCGCGTCG TCAGCTCCTG CCTAGGCCTG TACGCGCGCG      1140
TGCCGCGCGG CTCCCCCGCC GCGGCGCGCC TCCACGGCCG CGTGGCCGCC GCGTTCCAGG      1200
TGTTCAACGG CATCAGCCCC TTCGTCAAGT TCTCGCACTT CACCGCCAAC CAGGCCATCC      1260
AGGAGGCGTT CGAGCGGGAG GAGCGTGTGC ACATCATCGA CCTCGACATC ATGCAGGGGC      1320
TGCAGTGGCC GGGCCTCTTC CACATCCTTG TCTCCCGCCC CGGCGGCCCG CCCAGGGTCA      1380
GGCTCACCGG CCTGGGGGGC TCCATGGACG CGCTCGAGGC GACGGGGAAG CGCCTCTCCG      1440
ACTTCGCGGA CACGCTCGGC CTGCCCTTCG AGTTCTGCGC CGTCGCCGAG AAGGCCGGCA      1500
ACGTTGACCC GCAGAAGCTG GCGGTCACGC GCGGGGAGGC CGTCGCCGTC CACTGGCCGC      1560
ACCACTCGCT TTACGACGTC ATCGGCTCCG ACTCCAACAC GCTCTGGCTC ATCCAAAGGT      1620
CCTCCATTTT CTTTCTCTGC CTTTCTTCCA TGTCAAATCT TGATGCAATC ATGACCACTT      1680
TTCAGCTGCT GACATTGGAT AATGTGAGCT TTACGGCAAG CATCAAGTCG TGGTAGTACA      1740
TCCATTACAG CTATTTCTAA AATATTCTTC GGAGGTTTCC TGCTCATAGT AAAAAAAAAAT      1800
CGCGTTTTGA AGCTCAAAAG GCGATTCTT CCGAGGTTTG CTGTTGAGCG CTATTTTGA      1860
AACCCCATTT TCTCAATTGA TTTTATTTT TTAAGAAAAA ATTAGTTCAT TTTTCTCTG      1920
TGAAATGGAG TCCCAAACTA ACCCTAATAT TAAAAAAAC GCGCTTTGGA GCTCAAAACG      1980
CTCGTTGTTA TGACCAACCA GCTTTATAGG TTTAAAAAGG TTGAATCTTG ACAATGCTTT      2040
TGAAAAGGTT GAATCTTGAC AATGCTTTTG AGATGATACT GTAGTGTAGT CTGTAGTGGA      2100
GCATCCTCCA TGGTCTTTGG TGATCGAGAA TTCCTGCAGC CCGGGGGATC C      2151

```

(2) INFORMATION FOR SEQ ID NO:67:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 716 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: unknown

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:67:

```

Tyr Gln His His Gln Phe Xaa Met Xaa Val Gly Lys Arg Ser Xaa Gly
1           5           10           15
Phe Ser Xaa Xaa Phe Gly His Lys Val Xaa His Cys Leu Asn Xaa Ala
20           25           30
Ile Xaa Xaa Arg Asn Xaa Ser Ile Ile Tyr Cys Ser Tyr Xaa Lys Xaa
35           40           45
Lys Arg Tyr Ala Gln Glu Pro Ser Arg Glu Ala Lys Asn Xaa Lys Ile

```


50	55	60
Xaa Leu Tyr Xaa Lys Asn Met Ile Gly Cys Leu Phe Leu Met Lys Ile 65 70 75 80		
Tyr Xaa Ile Ile Ile Ser Trp Leu Asp Val Ala His Glu Val Val Leu 85 90 95		
Ala Ser Leu Asp Ser Cys Gly His Ser Ser Leu Leu Asp Ala Leu Thr 100 105 110		
Xaa Xaa Glu Val Ser Asn Leu Val Leu His Asn Ser Gly Asp Ser Cys 115 120 125		
Phe Leu His Cys Asn Xaa Tyr Xaa Cys Leu Ile His Ala Ser Val Thr 130 135 140		
Leu Cys Val Xaa Leu Val Leu Tyr Val Ser Leu Asp Cys Arg Val Ser 145 150 155 160		
Ala Thr Ser Val Ser Pro Cys Gly Phe Leu Val Ser Phe Val Leu Val 165 170 175		
Ser Asn Phe Leu Leu Ile Tyr Xaa Ser Asp Asn Xaa Phe Xaa Lys Tyr 180 185 190		
Ser Leu Val Pro Leu Met Leu Gln Leu Val Phe Ser Val Tyr Val Ser 195 200 205		
Ile Ile Ile Phe Pro Tyr Tyr Glu Ser Gly Arg Glu Thr Arg Pro Thr 210 215 220		
Arg Ala Ser Thr Ser Ser His Phe Thr Phe Ile Val Gly Ser Xaa Met 225 230 235 240		
Ser Gly Pro Arg Xaa Thr Pro Phe Ala Cys Lys Thr Thr Ser Ser Thr 245 250 255		
Arg Cys Xaa Val Ala Ser Arg Asn Ile Thr Met Cys Pro Val His Ser 260 265 270		
Phe Arg Arg Ser Arg Ile Arg Arg Arg Ser Arg Pro Arg Ser Arg Pro 275 280 285		
Pro Arg Pro Arg Pro Pro Pro Pro Arg Ser Gly Arg Arg Cys Ser Gly 290 295 300		
Gly Ser Ser Ala Thr Arg Arg Ala Ser Thr Cys Xaa Val Leu Thr Leu 305 310 315 320		
Leu Leu Gln Cys Ala Glu Ala Val Asn Ala Asp Asn Leu Asp Asp Ala 325 330 335		
His Gln Thr Leu Leu Glu Ile Ala Glu Leu Ala Thr Pro Phe Gly Thr 340 345 350		
Ser Thr Gln Arg Val Ala Ala Tyr Phe Ala Glu Ala Met Ser Ala Arg 355 360 365		
Val Val Ser Ser Cys Leu Gly Leu Tyr Ala Pro Leu Pro Pro Gly Ser 370 375 380		
Pro Ala Ala Ala Arg Leu His Gly Arg Val Ala Ala Ala Phe Gln Val 385 390 395 400		
Phe Asn Gly Ile Ser Pro Phe Val Lys Phe Ser His Phe Thr Ala Asn 405 410 415		

Gln Ala Ile Gln Glu Ala Phe Glu Arg Glu Glu Arg Val His Ile Ile
 420 425 430
 Asp Leu Asp Ile Met Gln Gly Leu Gln Trp Pro Gly Leu Phe His Ile
 435 440 445
 Leu Val Ser Arg Pro Gly Gly Pro Pro Arg Val Arg Leu Thr Gly Leu
 450 455 460
 Gly Ala Ser Met Asp Ala Leu Glu Ala Thr Gly Lys Arg Leu Ser Asp
 465 470 475 480
 Phe Ala Asp Thr Leu Gly Leu Pro Phe Glu Phe Cys Ala Val Ala Glu
 485 490 495
 Lys Ala Gly Asn Val Asp Pro Gln Lys Leu Gly Val Thr Arg Arg Glu
 500 505 510
 Ala Val Ala Val His Trp Pro His His Ser Leu Tyr Asp Val Ile Gly
 515 520 525
 Ser Asp Ser Asn Thr Leu Trp Leu Ile Gln Arg Ser Ser Ile Phe Leu
 530 535 540
 Leu Cys Leu Ser Ser Met Ser Asn Leu Asp Ala Ile Met Thr Thr Phe
 545 550 555 560
 Gln Leu Leu Thr Leu Asp Asn Val Ser Phe Thr Ala Ser Ile Lys Ser
 565 570 575
 Trp Xaa Tyr Ile His Tyr Ser Tyr Phe Xaa Asn Ile Leu Arg Arg Phe
 580 585 590
 Pro Ala His Ser Lys Lys Lys Ser Arg Phe Glu Ala Gln Lys Ala Ile
 595 600 605
 Ser Ser Glu Val Cys Cys Xaa Ala Leu Phe Trp Lys Pro His Phe Leu
 610 615 620
 Asn Xaa Phe Leu Phe Phe Lys Glu Lys Leu Val His Phe Ser Leu Val
 625 630 635 640
 Lys Trp Ser Pro Lys Leu Thr Leu Ile Leu Lys Lys Thr Arg Phe Gly
 645 650 655
 Ala Gln Asn Ala Arg Cys Tyr Asp Gln Pro Ala Leu Xaa Val Xaa Lys
 660 665 670
 Gly Xaa Ile Leu Thr Met Leu Leu Lys Arg Leu Asn Leu Asp Asn Ala
 675 680 685
 Phe Glu Met Ile Leu Xaa Cys Ser Leu Xaa Trp Ser Ile Leu His Gly
 690 695 700
 Leu Trp Xaa Ser Arg Ile Pro Ala Ala Arg Gly Ile
 705 710 715

WHAT IS CLAIMED IS:

1. An isolated nucleic acid molecule comprising a nucleotide sequence that encodes a SCARECROW protein
5 containing an amino acid sequence substantially similar to the sequence of MOTIF III (VHIID) of Arabidopsis SCR protein shown in FIGS. 13A-F.
2. An isolated nucleic acid molecule comprising a
10 nucleotide sequence that (a) encodes a scarecrow protein having the amino acid sequence shown of any one of SEQ ID NO:2, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:34, SEQ ID NO:35, SEQ ID NO:36, SEQ ID NO:37, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO:46, SEQ ID
15 NO:48, SEQ ID NO:50, SEQ ID NO:51, SEQ ID NO:52, SEQ ID NO:54, SEQ ID NO:56, SEQ ID NO:58, SEQ ID NO:59, SEQ ID NO:61, SEQ ID NO:63, SEQ ID NO:65 or SEQ ID NO:67; or (b) is the complement of the nucleotide sequence of (a).
- 20 3. An isolated nucleic acid molecule comprising a nucleotide sequence that hybridizes to the nucleic acid of Claim 2 and encodes a naturally occurring SCR gene product.
4. A nucleic acid molecule comprising a nucleotide sequence
25 that (a) encodes a SCR protein lacking one to four of the following motifs delineated in FIGS. 13A-F: MOTIF I, MOTIF II, MOTIF III, MOTIF IV, MOTIF V, or MOTIF VI; or (b) is the complement of the nucleotide sequence of (a).
- 30 5. A nucleic acid molecule comprising a nucleotide sequence that (a) encodes a polypeptide corresponding to MOTIF I, MOTIF II, MOTIF IV, MOTIF V or MOTIF VI of the SCARECROW protein delineated in FIGS. 13A-F; or (b) is the complement of the nucleotide sequence of (a).
35
6. The isolated nucleic acid molecule of Claim 1 comprising the nucleic acid sequence of any one of SEQ ID NO:1, SEQ ID

NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO:51, SEQ ID NO:53, SEQ ID NO:55, SEQ ID NO:57, SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:64 or SEQ ID NO:66.

5

7. A DNA vector containing the nucleotide sequence of Claim 1, 2, 3, 4, 5, or 6.

10 8. An expression vector containing the nucleotide sequence of Claim 1, 2, 3, 4, 5, or 6 operatively associated with a regulatory nucleotide sequence containing transcriptional and translational regulatory information that controls expression of the nucleotide sequence in a host cell.

15

9. A genetically engineered host cell containing the nucleotide sequence of Claim 1, 2, 3, 4, 5, or 6.

10. A genetically engineered host cell containing the
20 nucleotide sequence of Claim 1, 2, 3, 4, 5, or 6 operatively associated with a regulatory nucleotide sequence containing transcriptional and translational regulatory information that controls expression of the nucleotide sequence in a host cell.

25

11. An isolated SCARECROW protein.

12. The protein of Claim 11 having the amino acid sequence shown in FIG. 5E (SEQ ID NO:2).

30

13. A SCARECROW protein lacking one to four of the following motifs delineated in FIGS. 13A-F: MOTIF I, MOTIF II, MOTIF III, MOTIF VI, MOTIF V, or MOTIF VI.

35 14. A polypeptide corresponding to MOTIF I, MOTIF II, MOTIF IV, MOTIF V or MOTIF VI of the SCARECROW protein as delineated in FIGS. 13A-F.

15. An antibody that immunospecifically binds the protein or polypeptide of Claim 11, 12, 13 or 14.
16. An anti-idiotypic antibody that mimics an epitope of the SCARECROW protein.
17. A plant engineered to overexpress or underexpress the SCARECROW protein, so that cell division is modified and root development is altered
- 10 18. A plant engineered to overexpress the SCARECROW protein, so that cell division is increased in roots, resulting in thicker root development.
- 15 19. A transgenic plant containing a transgene having the nucleotide sequence of Claim 1, 2, 3, 4, 5, or 6.
- 20 20. A transgenic plant containing a transgene having the nucleotide sequence of Claim 1, 2, 3, 4, 5, or 6 operatively associated with a regulatory nucleotide sequence containing transcriptional and translational regulatory information that controls expression of the nucleotide sequence in a transgenic plant cell.
- 25 21. The transgenic plant of Claim 19, in which the transgene encodes an antisense molecule that suppresses expression of endogenous SCARECROW gene product, so that cell division is decreased in roots, resulting in thinner root development.
- 30 22. A genetically engineered plant in which the endogenous SCARECROW gene is disrupted or inactivated so that cell division is decreased in roots, resulting in thinner root development.
- 35 23. A transgenic plant containing a transgene encoding a gene of interest operatively associated with a SCARECROW promoter, so that the gene of interest is expressed in roots.

24. The transgenic plant of Claim 23, in which the gene of interest encodes a gene product that confers herbicide, salt, pathogen, or insect resistance.

5 25. A transgenic plant containing a transgene encoding a gene of interest operatively associated with a *SCARECROW* promoter, so that the gene of interest is expressed in stems.

26. The transgenic plant of Claim 25, in which the gene of
10 interest encodes a gene product that increases starch, lignin or cellulose biosynthesis.

27. A plant engineered to overexpress or underexpress the *SCARECROW* protein so that the stem or hypocotyl gravitropism
15 is altered.

28. The plant of Claim 28, which is less susceptible to lodging than a wild-type plant.

20

25

30

35

1/74

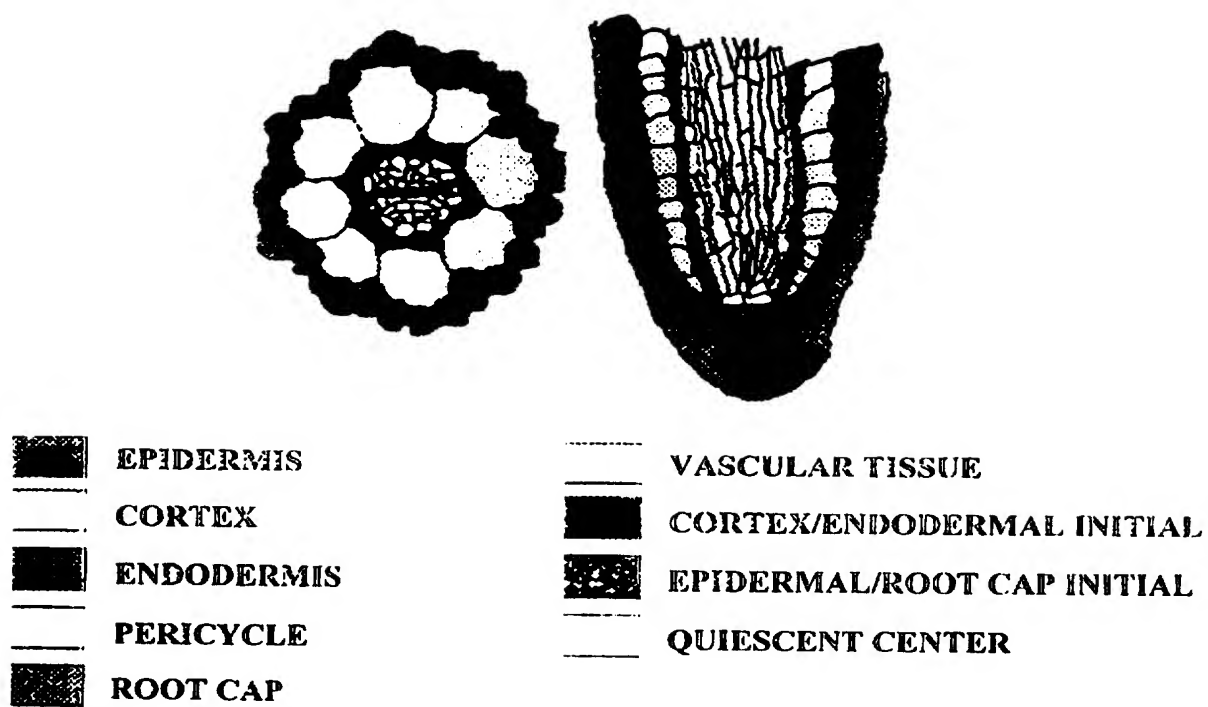


FIG. 1A

2/74

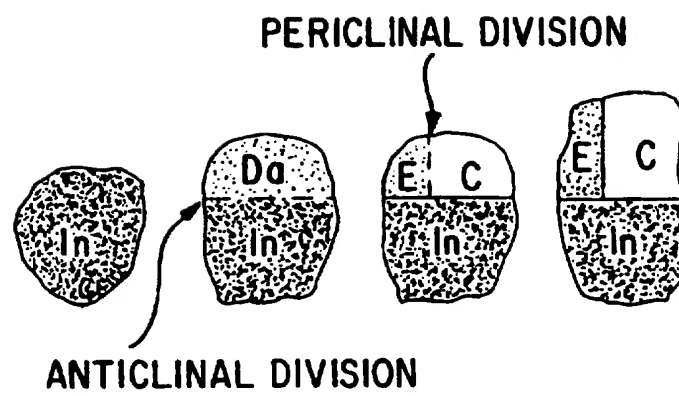


FIG. 1B

3/74

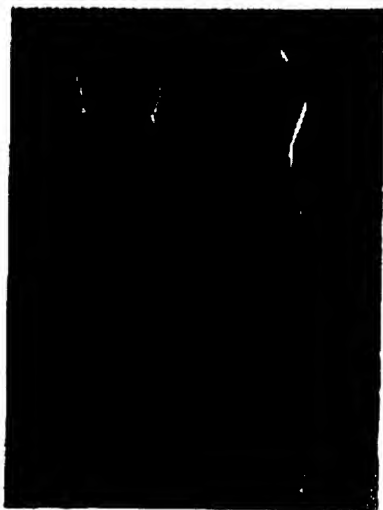


FIG. 2A

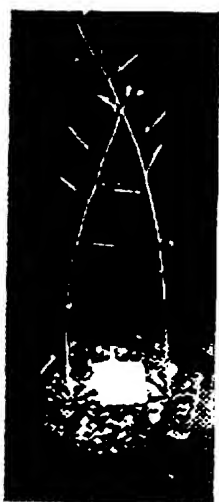


FIG. 2B

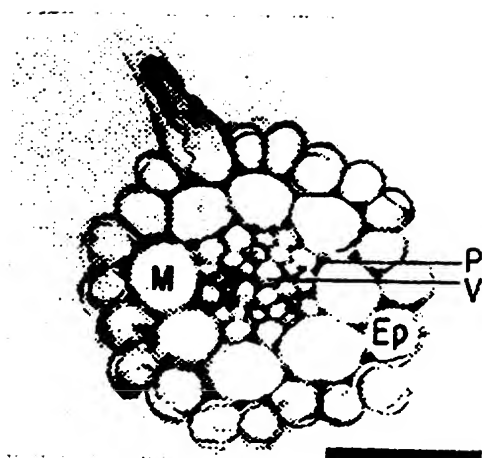


FIG. 2C

4/74

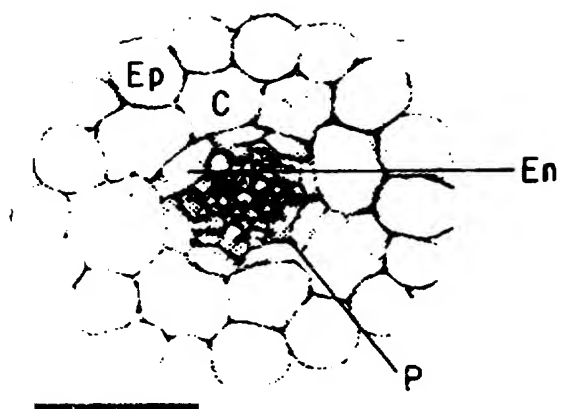


FIG.2D

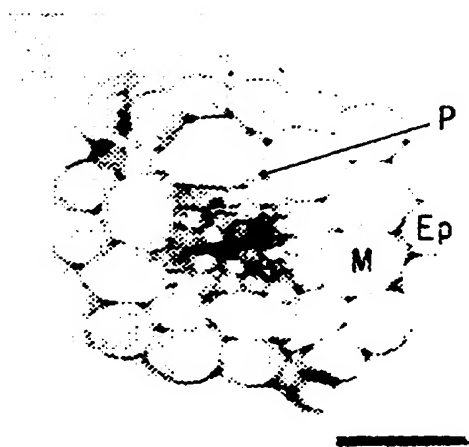


FIG.2E

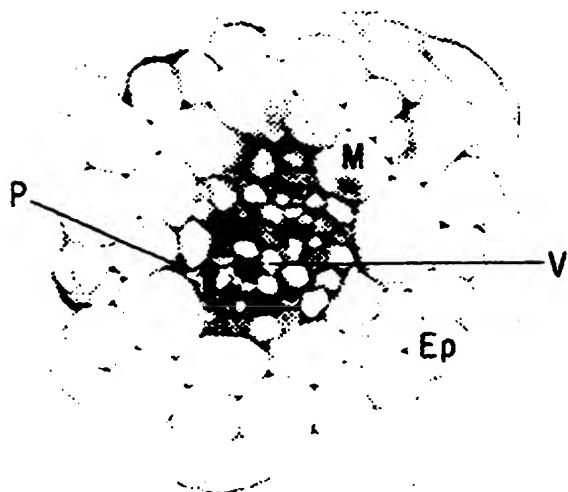


FIG.2F

5/74

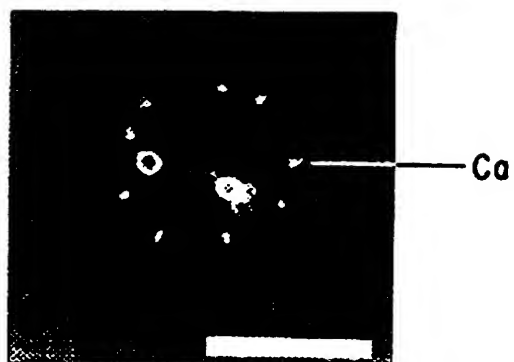


FIG. 3A



FIG. 3D

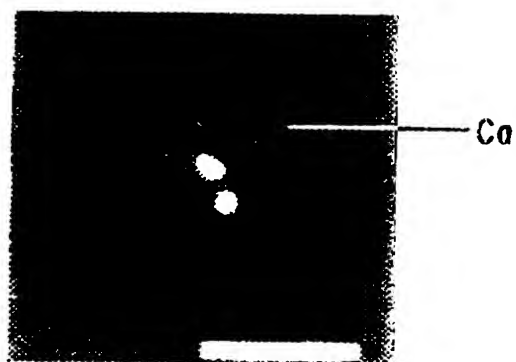


FIG. 3B

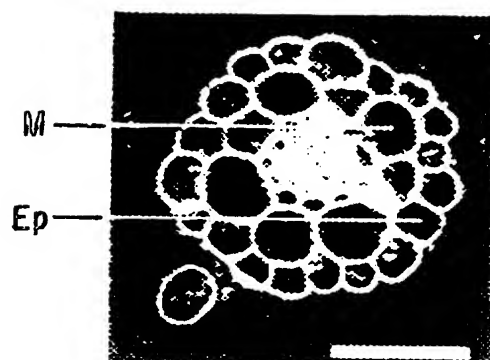


FIG. 3E

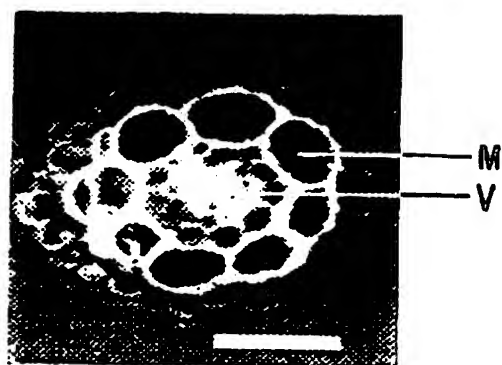


FIG. 3C

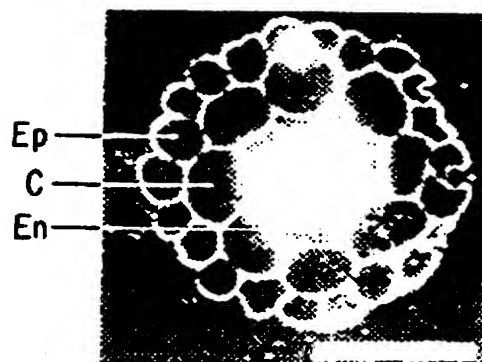


FIG. 3F

6/74

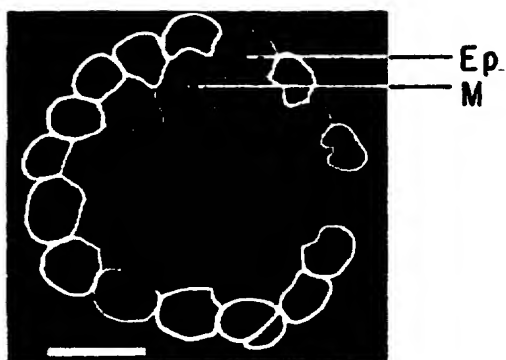


FIG. 4A

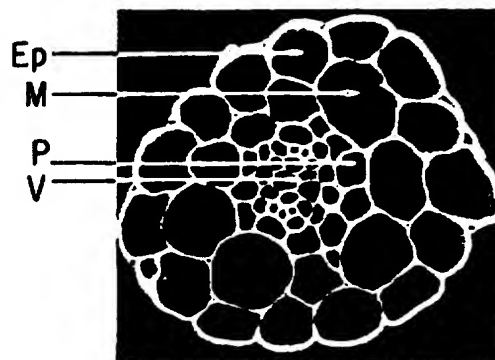


FIG. 4D

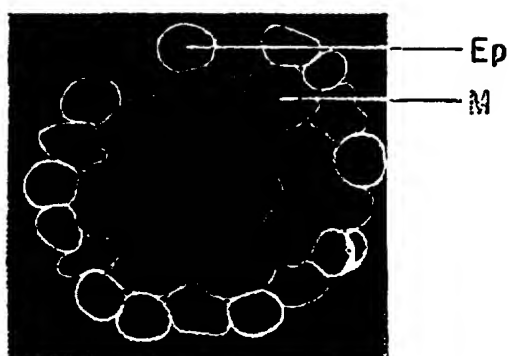


FIG. 4B

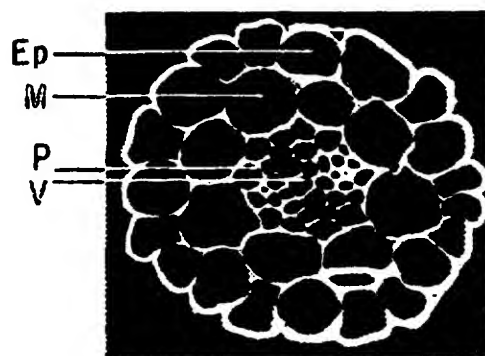


FIG. 4E

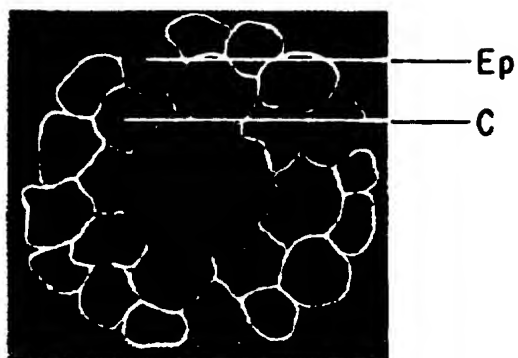


FIG. 4C

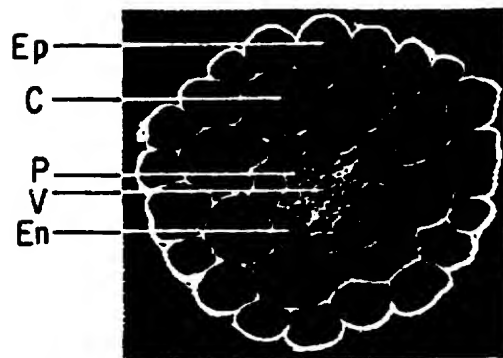


FIG. 4F

7/74

10 20 30 40 50 60 70 80 90 100 110 120
 -41
 CCTTATTATAACCATGCAATCTCAGACCAACAACCCTTCAATCTCCATGCCGGAATCCGGCGATTTCACCGTCGTCACACCTCCCTCATAGTCCCTCGAGACAACACTTCTTCCCGT
 +1
 M A E S G D F N G G Q P P P H S P L R I I S S G 24
 AGTAGCAGCAGCAACAACCGTGGTCCCTCCCTCCCTCCCTTTAGTATGGTGAGAAAAGATTACCTTCCGAGATGCTTCTAACCCCTGACTACACAACACTCTCTCGTCCCT
 S S S N N R G P P P P P P L V H V R K R L A S E M S S H P D Y H N S S R P 64
 CCTCGCGTGTCTCACCCTTCTTGACTCCAACACTACAAATACGTACAGCACAACACCACCGTCTCTTACGGCGGGGTACTGTCTCTCAACCAACCCACCACCTCTCTGTTGT
 P R R V S H L L D S N Y N I V I P Q Q Q P P S L I A A I V S S Q P N P P L S V C 104
 GCGTCTCTGGTCTCCCGTTTTCCCTTCAGACCGTGGTGGGAATGATGATGTCGGTACAAACCAATGGATCAAGACTCTTCACTCTTCTGCTTCCCTTACTGTATGGGTGAC
 G F S G L P V F P S D R G G R N V M S V Q P M D Q D S S S S A S P I V W V D 144
 GCCATTATCAGAGACCTTATCCATTCCCTCAACTTCTATTCCTCAACTTATCCAAACGTTAGAGACATTATCTCCCTTGTAAACCAATCTCGTGTCTTCTTGAATACAGG
 A I I R D L I H S S I S V S I P Q L I Q N V R D I I F P C H P N L G A L L E Y R 184
 CTCGATCTCTCATGCTCTTGATCCCTTCTCTGACCCCTTCTCTCAAACTTTCGAACCTCTCTATCAGATCTCCAACAATCTCTCTCCACACAGCAACAGCAGCACCAA
 L R S L M L L D P S S S S D P S P Q I F E P L Y Q I S N N P S P P Q Q Q Q Q H Q 224
 CAACAACAACACAGCATAAGCCCTCCCTCCGATTACAGCAGCAAGAAGAGAAAATCTTCTACCGATGCACCACCGCAACAGAGACAGTACGGCCACTGTCCCGCCGTCCAA
 Q Q Q Q Q H K P P P P I Q Q Q E R E N S S I D A P P Q P E I V I A I V P A V Q 264
 ACAAAATACGGCGGAGGCTTAAGAGAGAGGAAGAGAGATTAGAGGCAGAGCAAGCAGCAAGAGGATTACACCTTCTCACATTGCTGTCTGCTGCTGCTGCTGCTGCTGAT
 I N I A E A L R E R K E E I K R Q K Q D E E G L H L L T L L L Q C A E A V S A D 304
 AATCTCGAAGCAAGCAAGCTTCTTGTGAGATCTCTGAGTATCAACTCTTACGGACCTCAGCGCAGAGAGTAGCTGCTTACTTCTCGAAGCTATGTACGGAGATTACTCAAC
 N L E E A N K L L L E I S Q L S T P Y G T S A Q R V A A Y F S E A M S A R L L N 344

FIG.5A-1

TCGTGCTCGGAATTTACGCGGCTTGGCTTACCGTGGATGCCCTCAAAGCCATAGCTTGAATAATGGTCTCGCTTTCAGGCTCTTTAAATGGGATAAGCCCTTTAGTGAATTCCTCACAC
S S C L G I Y A A L P S R W M P Q T H S L K M V S A F Q V F N G I S P L V K F S H 384

TTTACAGCGAATCAGGCGATTCAAGAAGCATTTGAGAAAGACAGACAGTGTACACATCATTTGACTTGGACAICATGCCAGGACTTCAATGGCCCTGGTTTATTCACATTCITGCTTCTTAGA
F F T A N Q A I Q E A F E K E D S V H I I D L D I M Q G L Q W P G L F H I L A S R 424

CCCTGGAGGACCTCCACACGTGGGACTCACGGGACTTGGTACTTCCATGGAAGCTCTTCAGGCTACAGGAAAGCTCTTTTCGGATTTACAGATAAGCTTCCCTCGCTTTTTCAGTCTTCGC
P G G P P H V R L T G L G T S M E A L Q A T G K R L S D F T D K L G L P F E F C 464

CCCTTAGCTGAGAAAGTTGGAACCTTGGACACTGAGAGACTCAATGTGAGGAAAGCGAAGCTGTGCTGTTCAGCTGGCTTCAACATTCCTTTAATGATGTCACATGCCCTCTGATGCCACAC
P L A E K V G N L D T E R L N V R K R E A V A V H W L Q H S L Y D V T G S D A M 504

ACTCTCTGGTTACTCCAAAGGTAAATAAACATTACCTTTTAATCACTCTTTAICTATAAATATTTTAAGATTATATAGSAAAGATACTGTCTAAAGCTGGCTTTTTCGTTAATGA
T L W L L Q R 511

TTTCGGGAAAGACAGATTAGCTTCCTAAAGTTGACAGTAGTGGAGCAAGATTGACGCCCGCTGGTCTTCTTCTTAGGAAGATTGTAGAGGCAATACATTACTACTGTCGCACTCTTTGA
L A P K V V T V V E Q D L S H A G S F L G R F V E A I H Y Y S A L F D 546

CTCACTGGGACCAAGCTACGGCGAAGACAGAGTGAACAGACAGACATGTCTGTCGAACAGCAGCTATTATCGAAAGACAGATACGGAAATGATTAGCGGTTGGAGGACCATTGAGAGCGGTGAAGT
S L G A S Y G E E S E R H V V E Q Q L L S K E I R N V L A V G G P S R S G E V 586

GAACTTTGAGAGCTGCAGGAGAAAAATGCAACAAATGTGGGTTTAAAGGTATATCTTTAGCTGGAAATGCGAGTACACAAGGACACTACTGTGGGAATGTTTCCTTCGGATCGGTTACAC
K F E S W R E K M Q Q C G F K G I S L A G N A A T Q A T L L L G H F P S D G Y T 626

TTTGGTTGATGATAATGGTACACTTAAGCTTGGATGGAAGAGATCTTTCGTTACTCACTGCTTCAGCTTGGAGCGCTCGTTCTTAGTTTCTTCCTTTTTCACAAACAATGTCGCCATA
L V D D N G T L K L G W K D L S L L T A S A W T P R S STOP 653

2163
AAT

FIG. 5A-2

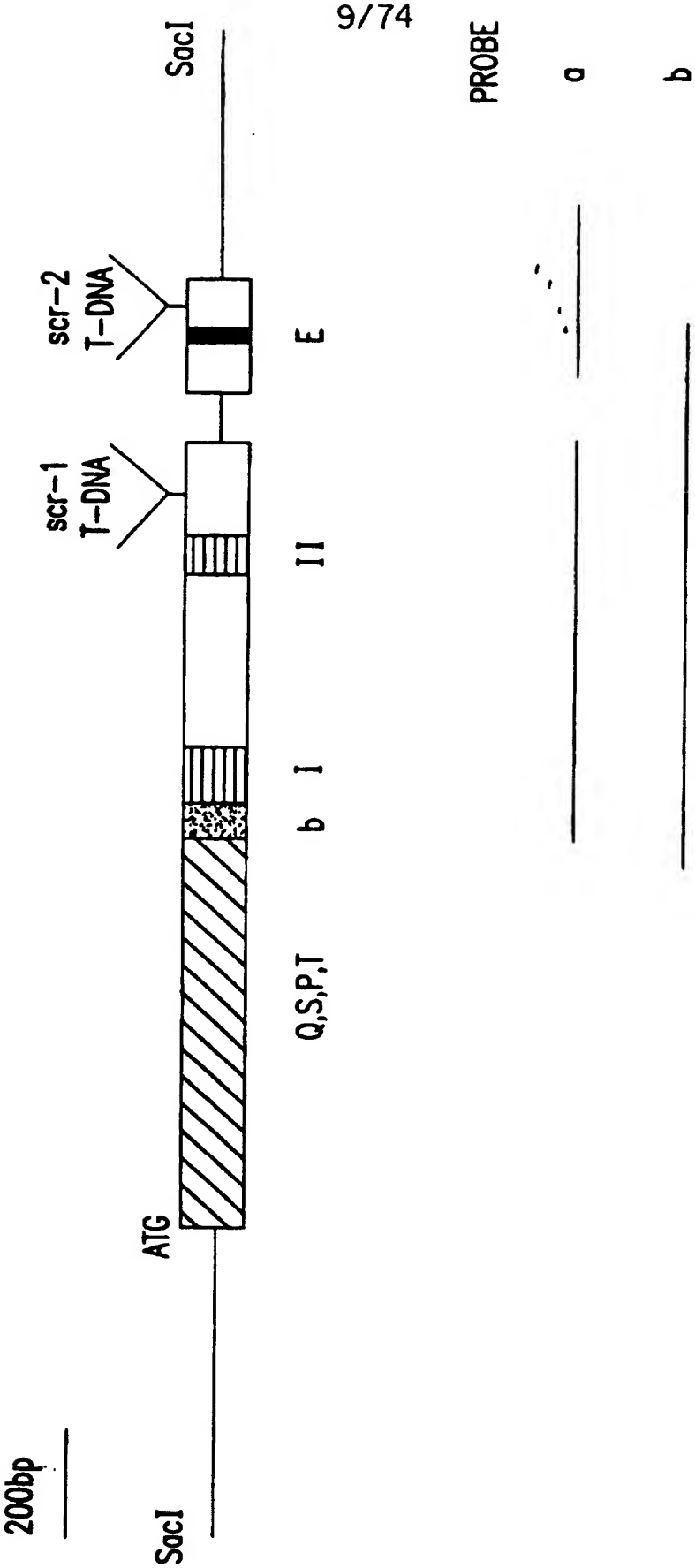


FIG.5B

10/74

SCR bZIP-like domain	PAVQTNTAEALRERKEEIKRQKQ	1
	:	D
GCN4 (yeast)	LKRARNTEAARRSRARKLQRMKQ	L
TGA1 (Arabidopsis)	RRLAQNREAARKSRLRKKAYVQQ	L
C-Fos (mouse)	IRRERNKMAAAKCRNRRREL TDT	L
c-JUN (human)	RKRMRNR IAASKCRKKLERIAR	L
CREB (human)	VRLMKNREAARECRKKKEYVKC	L
Opaque-2 (maize)	KRKESNRESARRSRYRKA AHLKE	L
OBF2 (maize)	MRQIRNRDSAMKSREKKSYIKD	L
RAF-1 (rice)	RRMVS NRESARRSRKKKQ AHLAD	L

FIG.5C

11/74

SCR	VH11D domain	
SCR	AFEKEDSVH11DDLIMQGLQWPGLFHILASRPGGPPHVRLTGL	1
F13896	AVKNESFVH11DFQISQGGQWVSLIRALGARPGGPPNVRLTGI	
Z37192	AMEGEKMHVH11DLDASEPAQWLALLQAFNSRPEGPPHLRLITGV	
Z25645	AIKGEEVH11DFDINQGNQYMTLIRSLA	
D41474	IHV1DFXLGVGGQWASFLQELAHRRG	
T18310	VH11FXLMQGLQWPALMDVFSARKGGPPKLRITGI	

FIG. 5D

12 / 74

FIG. 5E-1

MetAlaGluSerGlyAspPheAsnGlyGlyGlnProProProHisSerProLeuArgThr
 ThrSerSerGlySerSerSerSerAsnAsnArgGlyProProProProProProPro
 LeuValMetValArgLysArgLeuAlaSerGluMetSerSerSerAsnProAspTyrAsnAsn
 SerSerArgProProArgArgValSerHisLeuLeuAspSerAsnTyrAsnThrValThr
 ProGlnGlnProProSerLeuThrAlaAlaAlaThrValSerSerGlnProAsnProPro
 LeuSerValCysGlyPheSerGlyLeuProValPheProSerAspArgGlyGlyArgAsn
 ValMetMetSerValGlnProMetAspGlnAspSerSerSerSerSerAlaSerProThr
 ValTrpValAspAlaIleIleArgAspLeuIleHisSerSerThrSerValSerIlePro
 GlnLeuIleGlnAsnValArgAspIleIlePheProCysAsnProAsnLeuGlyAlaLeu
 LeuGluTyrArgLeuArgSerLeuMetLeuLeuAspProSerSerSerSerAspProSer
 ProGlnThrPheGluProLeuTyrGlnIleSerAsnAsnProSerProProGlnGlnGln
 GlnGlnHisGlnGlnGlnGlnHisLysProProProProIleGlnGlnGln
 GluArgGluAsnSerSerThrAspAlaProProGlnProGluThrValThrAlaThrVal
 ProAlaValGlnThrAsnThrAlaGluAlaLeuArgGluArgLysGluGluIleLysArg
 GlnLysGlnAspGluGluGlyLeuHisLeuLeuThrLeuLeuGlnCysAlaGluAla
 ValSerAlaAspAsnLeuGluGluAlaAsnLysLeuLeuLeuGluIleSerGlnLeuSer
 ThrProTyrGlyThrSerAlaGlnArgValAlaAlaTyrPheSerGluAlaMetSerAla

13/74

FIG. 5E-2

ArgLeuLeuAsnSerCysLeuGlyIleTyrAlaAlaLeuProSerArgTrpMetProGln
ThrHisSerLeuLysMetValSerAlaPheGlnValPheAsnGlyIleSerProLeuVal
LysPheSerHisPheThrAlaAsnGlnAlaIleGlnGluAlaPheGluLysGluAspSer
ValHisIleAspLeuAspIleMetGlnGlyLeuGlnTrpProGlyLeuPheHisIle
LeuAlaSerArgProGlyGlyProProHisValArgLeuThrGlyLeuGlyThrSerMet
GluAlaLeuGlnAlaThrGlyLysArgLeuSerAspPheThrAspLysLeuGlyLeuPro
PheGluPheCysProLeuAlaGluLysValGlyAsnLeuAspThrGluArgLeuAsnVal
ArgLysArgGluAlaValAlaValHisTrpLeuGlnHisSerLeuTyrAspValThrGly
SerAspAlaHisThrLeuTrpLeuLeuGlnArgLeuAlaProLysValValThrValVal
GluGlnAspLeuSerHisAlaGlySerPheLeuGlyArgPheValGluAlaIleHisTyr
TyrSerAlaLeuPheAspSerLeuGlyAlaSerTyrGlyGluGluSerGluGluArgHis
ValValGluGlnGlnLeuLeuSerLysGluIleArgAsnValLeuAlaValGlyGlyPro
SerArgSerGlyGluValLysPheGluSerTrpArgGluLysMetGlnGlnCysGlyPhe
LysGlyIleSerLeuAlaGlyAsnAlaAlaThrGlnAlaThrLeuLeuLeuGlyMetPhe
ProSerAspGlyTyrThrLeuValAspAspAsnGlyThrLeuLysLeuGlyTrpLysAsp
LeuSerLeuLeuThrAlaSerAlaTrpThrProArgSerSTOP

14/74

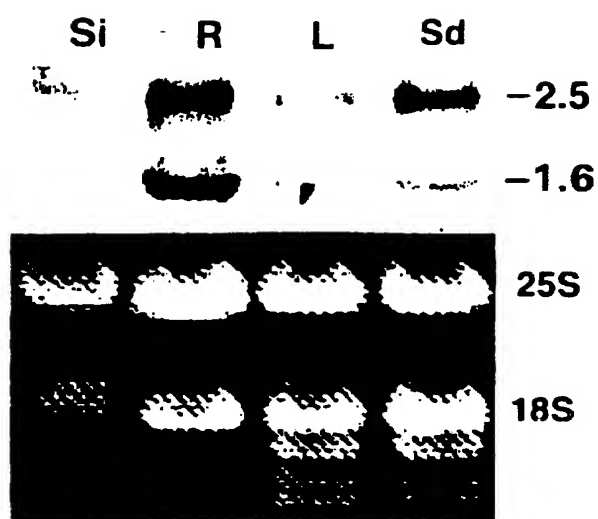


FIG.6A



FIG.6B

15/74

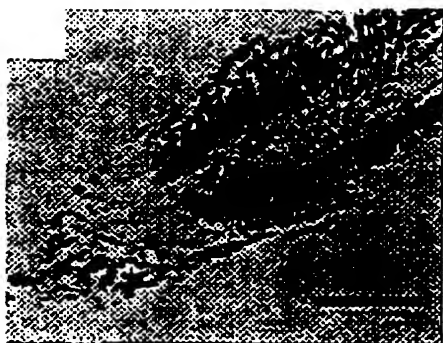


FIG.7A

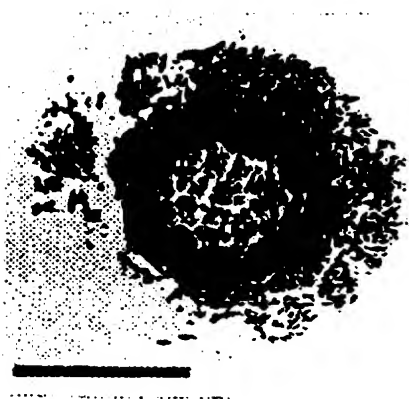


FIG.7B



FIG.7C

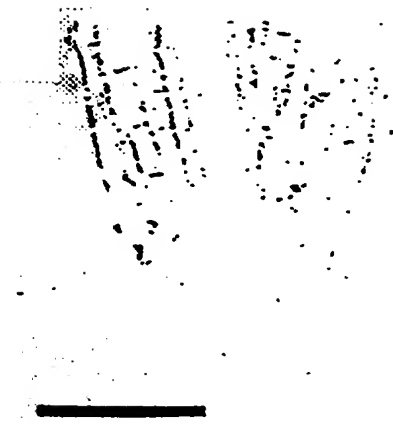


FIG.7D

16/74



FIG. 7E

FIG. 7F

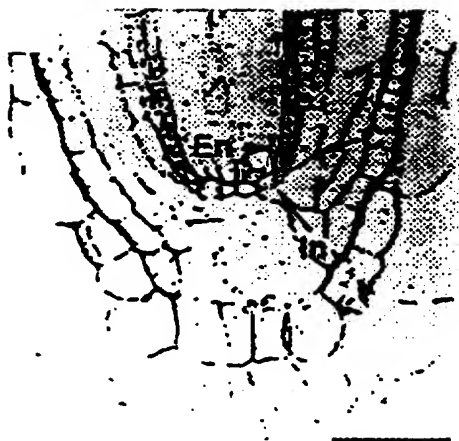
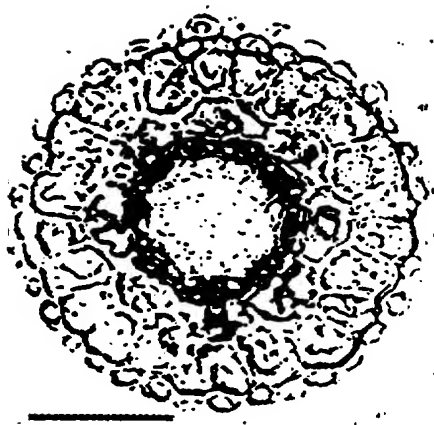


FIG. 7G

17/74

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
GGCACGAGCC	CAACGGGTCC	TGAGCTTCTT	ACTTATATGC	ATATCTTGT	50
G T S P	T G P	E L L	T Y M H	I L Y	
TGAAGCCTGC	CCTTATTTCA	AATTCCGTTA	TGAATCTGCT	AATGGAGCTA	100
E A C	P Y F K	F G Y	E S A	N G A I	
TAGCTGAAGC	TGTGAAGAAC	GAAAGTTTTC	TGCACATTAT	CGATTTCACG	150
A E A	V K N	E S F V	H I I	D F Q	
ATTTCTCAAG	GTGGTCAATG	GGTGAGTTTG	ATCCGTGCTC	TTGGTGCTAG	200
I S Q G	G Q W	V S L	I R A L	G A R	
ACCTGGTGGA	CCTCCGAACG	TTAGGATAAC	GGGAATTGAT	GATCCGAGAT	250
P G G	P P N V	R I T	G I D	D P R S	
CATCGTTTGC	TCGTCAAGGA	GGACTTGAGT	TAGTTGGACA	AAGACTTGGG	300
S F A	R Q G	G L E L	V G Q	R L G	
AAGCTAGCTG	AAATGTCCCG	TGTTCCGTTT	GAGTTCCATG	GAGCTGCTTT	350
K L A E	M C G	V P F	E F H G	A A L	
ATGCTGCACG	GAAGTCGAAA	TCGAGAAGCT	AGGAGTTAGA	AATGGAGAAG	400
C C T	E V E I	E K L	G V R	N G E A	
CGCTCGCGGT	TAACTTCCCG	CTTGTTCTTC	ACCACATGCC	TGATGAGAGT	450
L A V	N F P	L V L H	H M P	D E S	
GTAAGTGTGG	AGAATCACAG	AGATAGATTG	TTGAGATTGG	TCAAACACTT	500
V T V E	N H R	D R L	L R L V	K H L	
GTCACCAAAC	GTTGTGACTC	TGGTTGAGCA	AGAAGCGAAT	ACAAACACTG	550
S P N	V V T L	V E Q	E A N	T N T A	
CGCCGTTTCT	TCCCCGTTT	GTCGAGACAA	TGAACCATT	CTTGGCAGIT	600
P F L	P R F	V E T M	N H Y	L A V	

FIG.8A

18/74

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TTCCAATCAA	TAGATGTGAA	ACTCGCTAGA	GATCACAAGG	AAAGGATCAA	650
F E S I	D V K	L A R	D H K E	R I N	
TGTTGAGCAG	CATTGTTTGG	CTAGAGAGGT	TGTGAATCIT	ATAGCTTGTG	700
V E Q	H C L A	R E V	V N L	I A C E	
AAGGTGTTGA	AAGAGAAGAG	AGGCACGAGC	CACTAGGGAA	ATGGAGGTCT	750
G V E	R E E	R H E P	L G K	W R S	
CGGTTTCACA	TGGCGGGATT	TAAACCGTAT	CCTTTGAGCT	CGTATGTGAA	800
R F H M	A G F	K P Y	P L S S	Y V N	
CCCAACAATC	AAAGGATTGC	TTGAGAGTTA	TTCAGAGAAG	TATACACTTG	850
A T I	K G L L	E S Y	S E K	Y T L E	
AAGAAAGAGA	TGCAGCATTG	TATTTAGGAT	GGAAGAATCA	ACCTCTTATC	900
E R D	G A L	Y L G W	K N Q	P L I	
ACTTCTTGTG	CTTGGAGGTA	ACTAATAAAA	ACCTTGTTGG	GTTTCAGAAG	950
T S C A	W R X				
AGATTAGAAA	CTTCTTTTAA	AGTTTGCAGA	ATCTGTTTGT	AAAAGTAAAA	1000
CTCATGCATG	ATCCGNAGGA	ACAAGTTGTC	AAATGTTGTA	GTAAGTAACTG	1050
ATATGTTGAT	GACCCAAAAA	AAAAAAAAAA	AAAAA		1085

FIG.8B

19/74

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
GCTATGGAAG	GAGAGAAGAT	GGTTCATGTG	ATTGATCTCG	ATGCTTCTGA	50
A M E G	E K M	V H V	I D L D	A S E	
GCCAGCTCAA	TGGCTTGCTT	TGCTTCAAGC	TTTAACTCT	AGGCCTGAAG	100
P A Q	W L A L	L Q A	F N S	R P E G	
GTCCACCTCA	TTTGAGAATC	ACTGGTGTTT	ATCACCAGAA	GGAAGTGCTT	150
P P H	L R I	T G V H	H Q K	E V L	
GAACAAATGG	CTCATAGACT	CATTGAGGAA	GCAGAGAAAC	TCGATATCCC	200
E Q M A	H R L	I E E	A E K L	D I P	
GTTTCAGTTT	AATCCCGTTG	TGAGTAGGTT	AGACTGTTTA	AATGTAGAAC	250
F Q F	N P V V	S R L	D C L	N V E Q	
AGTTGCGGGT	TAAAACAGGA	GAGGCCTTAG	CCGTTAGCTC	GTTTCTTCAA	300
L R V	K T G	E A L A	V S S	V L Q	
TTGCATACCT	TCTTGGCCTC	TGATGATGAT	CTCATGAGAA	AGAACTGCGC	350
L H T F	L A S	D D D	L M R K	N C A	
TTTACGGTTT	CAGAACAACC	CTAGTGGAGT	TGACTTGCAG	AGAGTTCTAA	400
L R F	Q N N P	S G V	D L Q	R V L M	
TGATGAGCCA	TGGCTCTGCA	GCTGAGGCAC	GTGAGAATGA	TATGAGTAAC	450
M S H	G S A	A E A R	E N D	M S N	
AACAATGGGT	ATAGCCCTAG	CGGTGAGTCG	GCCTCATCTT	TGCCTTTACC	500
N N G Y	S P S	G D S	A S S L	P L P	
AAGTTCAGGA	AGGACTGATA	GCTTCCTCAA	TGCTATTTGG	GGTTTGTCTC	550
S S G	R T D S	F L N	A I W	G L S P	
CAAAGGTCAT	GGTGSTCACT	GAGCAAGACT	CAGACCACAA	CGGCTCCACA	600
K V M	V V T	E Q D S	D H N	G S T	

FIG.9A

20/74

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
CTAATGGAGA	GGCTATTAGA	ATCACTTTAC	ACCTACGCAG	CATTGTTTGA	650
L M E R	L L E S	L Y T Y	A A L F	D	
TTGCTTGGA	ACAAAAGTTC	CAAGAACGTC	TCAAGATAGG	ATCAAAGTGG	700
C L E T	K V P R	T S Q D	R I K V	E	
AGAAGATGCT	CTTCGGGGAG	GAGATCAAGA	ACATCATATC	CTCGGAGGGA	750
K M L F	G E E I	K N I I	S C E G		
TTTGAGAGAA	GAGAAAGACA	CGAGAAGCTT	GAGAAATGGA	GCCAGAGGAT	800
F E R R	E R H E	K L E K	W S Q R	I	
DGATTTGGCT	GGTTTTGGGA	ATGTTCCCTCT	TAGCTATTAT	GCGATGTTC	850
D L A G	F G N V	P L S Y	Y A M L	Q	
AGGCTAGGAG	ATTGCTTCAA	GGGTGCGGTT	TTGATGGGTA	TAGAATCAAG	900
A R R L	L Q G C	G F D G	Y R I K		
GAAGAGAGCG	GGTGCGCAGT	AATTTGCTGG	CAAGATCGAC	CTCTATACTC	950
E E S G	C A V I	C W Q D	R P L Y	S	
GGTATCAGCT	TGGAGATGCA	GGAAGTGAAT	GATATATTAC	AGTTTGTCTT	1000
V S A W	R C R K	X			
CTATTTTGGT	TATGAGCAGA	GTCCCTTTCT	TTTTTGATA	CATGGGGACA	1050
CAATCTTAGT	TGTTTTGTGA	TGGTGACTTT	CTGTCTCTTT	ATGCTATTTT	1100
GGCTTAAATG	CTTCTACTGC	CTCTGCATGT	AAAGCCTTTG	TGTGTTGGTT	1150
CAATTTGGTC	TGGTGTGGGT	GTAATACCAA	ACCAAATCCA	ATTTGAGCTG	1200
AAGATAACTA	ATTTGATGAT	CGGCTCGTGC	C		1231

FIG.9B

21/74

FIG. 10

CTTTGTCAAT GGTAAATGAG CTGAGGCAGA TAGTTTCTAT CCAAGGAGAC 50
CCTTCTCAGA GAATCGCAGC TTACATGGTG GAAGGTCTAG CTGCAAGAAT 100
GGCCGCTTCA GGAAATTC A TCTACAGAGC ATTGAAATGC AAAGAGCCCTC 150
CTTCGGATGA GAGGCTTGCA GCTATGCAAG TCCTGTTTGA AGTCTGCCCT 200
TGTTTCAAGT TCGGGTTTTT AGCAGCTAAT GGTGCGATAC TTGAAGCAAT 250
CAAAGGTGAA GAAGAAGTTC ACATAATCGA TTTCGATATA AACCAAGGA 300
ACCAATACAT GACACTGATA CGAAGCATTG CTGAGTTGCC TGGTAAACGA 350
CCTCGCCTGA GGTAAACAGG AATTGATGAC CCTGAATCAG TCCAACGCTC 400
CATTGGAGGG CTAAGAATCA TCAATCTAAG ACTCGAGCAA CTCGCAGAG 450
ATAATGGAGT ATCCTTCAAA TTCAAAGCAA TGCCCTTCAAA GACTTCGATT 500
GTCTCTCCAT CAACACTCGG TTGCAAAACCA GGAGAAACCT TAATCAGTGA 550
ACTTTGCATT CCAACTTCAC CACATGCCCTG ACGAGAGTGT CACAACAGTA 600
AACCAGCGGG ACGAGCTACT TCACATGGTC AAAAGCTTAA ACCCGCTTGT 650
CACGGTCGTT GAACAAGACG TGAACACAAA CACTTCACCG TTCTTTCCCA 700
GATTCATAGA GGCTTACGAA TACTACTCAG CAGTTTTCGA GTCTCTAGAC 750
ATGACACTTC CAAGAGAAAG CCAAGAGAGG ATGAATGTAG AAAGACAGTG 800
TCTCGCTAGA GACATAGTCA ACATTGTTGC TTGCGAAGGA GAAGAACGGA 850
TAGAGAGATA CGAGGCTGCG GGAAAATGGA GAGCAAGGAT GATGATGGCT 900
GGATTCAATC CAAAACCAAT GAGTGCTAAA GTAAACCAACA ATATACAAA 950
CCTGATAAAG CAACAATATT GCAATAAGTA CAAGCTTAAA GAAGAAATGG 1000
GTGAGCTCCA TTTTGTCTGG GAGGAGAAA GCTTAATCGT TGCTTCAGCT 1050
TGGAGGTAAG ATAAGTGACA AGAGCATATA GTCTTTATGT TTCATAAAAC 1100
ATAAATTATGT TTTTACTGTA ATCTTGGGTT ATTGTGTAAAC TGGTTAAATC 1150
ATCTCCATGT ATTATTACCA GAGGTTAGGG GTGATCACAG GTACTAAAAG 1200
CTAATCTAAC ACTTATGGAA GAATTTTCTT TTCTTTTCTT TCCCTATTAT 1250
ATAAAAATAA TTAGAGTTTT GTTCTTAAC CTATTTGCTA AGTGTAATG 1300
AGTCTTTTACA TGTTCAATTT TCAGTTCAAA TGGTTAAATT TGTTAAGGTT 1350
CTCACTTAAA AAAAAA

22/74

Zm-sc11

10	20	30	40	50
CCAGGAGGCGTTCGAGCGGGAGGAGCGTGTGCACATCATCGACCTCGACA				
Q E A F E R E E R V H I I D L D I				
60	70	80	90	100
TCATGCAGGGGCTGCAGTGGCCGGGCCTCTTCCACATCCTTGCCTCCCGC				
M Q G L Q W P G L F H I L A S R				

FIG.11A

23/74

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
CACGCGTCCG	TCAAAGGATA	CAACCATGTA	CACATAATTG	ACTTTTCCCT	50
H A S V	K G Y	N H V	H I I D	F S L	
GATGCAAGGT	CTCCAGTGGC	CGGCACTCAT	GGATGTCTTC	TCCGCCCCGTG	100
M Q G	L Q W P	A L M	D V F	S A R E	
AGGGTGGGCC	ACCAAAGCTC	CGAATCACAG	GCATTGGCCC	GAACCCAATA	150
G G P	P K L	R I T G	I G P	N P I	
GGTGGCCGTG	ACGAGCTCCA	TGAAGTGGGA	ATTGCGCTCG	CCAAGTATGC	200
G G R D	E L H	E V G	I R L A	K Y A	
ACACTCGGTG	GGTATCGACT	TCACTTTCCA	GGGAGTCTGT	GTCGATCAAC	250
H S V	G I D F	T F Q	G V C	V D Q L	
TTGATAGGTT	GTGCGACTGG	ATGCTTCTCA	AACCAATCAA	AGGAGAGGCA	300
D R L	C D W	M L L K	P I K	G E A	
GTTGCCATAA	ACTCCATCCT	ACAACTCCAT	CGCCTCCTCG	TTGACCCAGA	350
V A I N	S I L	Q L H	R L L V	D P D	
TGCAAACCCA	GTGGTGCCCG	CACCAATAGA	TATCCTCCTC	AAATTGGTCA	400
A N P	V V P A	P I D	I L L	K L V I	
TCAAGATAAA	CCCCATGATC	TTCACGGTGG	TTGAGCATGA	GGCAGATCAC	450
K I N	P M I	F T V V	E H E	A D H	
AACAGACCAC	CACTACTAGA	GAGGTTCAC	AATGCCCTCT	TCCACTATGC	500
N R P P	L L E	R F T	N A L F	H Y A	
GACCATGTTT	GACTCTTTGG	AGGCCATGCA	TCGTTGTACC	AGTGGTAGAG	550
T M F	D S L E	A M H	R C T	S G R D	
ACATCACCGA	CTCACTCACA	GAGGTGTACC	TTGAGGTGA	GATTTTGTAC	600
I T D	S L T	E V Y L	R G E	I F D	

FIG.11B1

24/74

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
ATTGTCTGCG	GCGAGGGCAG	TGCACGCACC	GAACGTCATG	AGTTGTTTGG	650
I V C G	E G S	A R T	E R H E	L F G	
TCACTGGAGG	GAGAGGCTCA	CCTATGCTGG	GCTAACTCAA	GTGTGGTTCC	700
H W R	E R L T	Y A G	L T Q	V W F D	
ACCCCGATGA	GGTTGACACG	CTAAAAGACC	AGTTGATCCA	TGTGACATCC	750
P D E	V D T	L K D Q	L I H	V T S	
TTATCTGGCT	CTGGGTTCAA	CATCCTAGTG	TGTGATGGCA	GCCTTGCACT	800
L S G S	G F N	I L V	C D G S	L A L	
AGCGTGGCAT	AATCGCCCGT	TATATGTGGC	AACAGCTTGG	TGTGTGACAG	850
A W H	N R P L	Y V A	T A W	C V T G	
GAGGAAATGC	TGCCAGTTCC	ATG GTTGGCA	ACATCTGTAA	GGGTACAAAT	900
G N A	A S S	M V G N	I C K	G T N	
GATAGTAGAA	GAAAGGAAAA	CCGTAATGGA	CCCATGGAGT	AGCAGGAAGA	950
D S R R	K E N	R N G	P M E X		
ATAACCATGT	CATGAGCAAA	TCGATCAAGT	AATAAAATGC	ACTGATGACA	1000
TGCATGGTGA	TCTAAAGTTT	TTTTGCGTGA	ATGTGCAATG	ACGAATTGTT	1050
CAATTTGAAT	AACCTAATCA	TGAGACTCAA	AAAAAAAAAA	AAA	1093

FIG.11B2

25/74

CCCAACTTGG GAAGCCCTTC CTCCGCTCCG CCTCCTACCT CAAGGAGGCC
CTCCTCCTCG CACTCGCCGA CAGCCACCAT GGCTCCTCCG GCGTCACCTC
GCCGCTCGAC GTTGCCCTCA AGCTGCGAGC ATACAAGTCT TTCTCTGACC
TGTCACCTGT GCTCCAGTTC ACTAACTTA CCGCAACAAG GCGCTTCTTG
ATGAGATTGG TGGCATGGCA ACTTCCTGCA TCCATGTCAT TGACTTTGAT
CTCGGTGTG GTGGTCAGTG GGCTTCCTTC TTGCAGGAGC TTGCCCAACG
CCGGGAGCT GGAGGTATGG CCTTGCCGTT GTTGAAGCTC ACGGCTTTCA
TGTCGACTGC TTCTCACCAT CCACTGGAGC TGCACCTTAC CCAGGATAAC
CTCTCTCAGT TTGCCGCAGA GCTCAGAATT CCTTTCGAAT TCAATGCCGT
CAGTCTTGAT GCATTCAATC CTGCGGAATC TATTTCCTCC TCTGGTGATG
AAGTTGTTGC TGTTAGCCTC CCTGTTGGCT GCTCTGCTCG TGCACCAACG
CTGCCAGCGA TTCCTCGGTT GGTGAAACAG CTTTGTCCTA AGGTTGTCGT
GGCTATTGAT C

FIG. 12A

26/74

TTTTTTTTTT TTTTTTTTTT TTTTTTTTTT TTTTTTTTTT TTTTTTTTTT TTTTTTTTTT
ATATTAAATC TGTACCACAC AACCATTGGA TAGGTTAAAT TACAGAGCAA CAGCAGTATA
TCTCTACTCA TAAGCAGTGT TTCCAATGAG ATGATCATGG CTAATTGAGC
AGAGCATGGC AACAACTAA AGCAACATCA TTAGCTATAG AGACTGACAC
CAATATTCTT AAATCCACTA GGCTAGCTAA TAAGCTGCAA CGAAAGCAA
TATGAAGAGT TCAACAGCTC AAGACAACAA TTTCATTGTC AACATTTAAT
TGCAAGAATA AATGGACATT ACTGGAGTGG TCGATGCTTG CAAACGGTGG
TGGAACCTTG GTGGAGTGAA GCTTATGGCT GATCAGCACC GCCAAGATGA
TATGGATACA AGCTCCCCAC GCTGCCAGTA GAGCGTAAGA GCAGCTCCGC
GTTTCTCCAC ATGGAATCCT CGGACCTGCA CCCGCTTCAG GAGGCAGTCT
GC

FIG. 12B

27/74

SCR MAESGDFNGGQPPPHSPLRTTSSGSSSSNNRGPPPPPLVMVRKR----LASEMSS
 TF1 MKRD---HHQFQGRLSNHGTSSSSSSISKDK--MMVKKKEEDGGCNMDDLAV----
 TF4 MKRDHHHHHQ-----DKKTMMM--NEEDDGNGM-DELLAV----

|----- MOTIF I -----|
 NPDYNNSSRPPRRVSHLLDSNYNTVTPQQPPSLTAAATVSSQPNPPLSVCGFSG
 TF1 -LG YKVRSEMAEVALKLEQLETMSNAQEDGLSHLATDAAHYNPSELYS----
 TF4 -LG YKVRSEMAADVAKLEQLEVMSNVQEDDLSQLATETVHYNPAELYT----

SCR LPVFPSDRGGRNVMSVQPMDDQSSSSASPTVWDAIIRDLIHS----STSVSIPQL
 TF1 -----WLDNMLSELNPPPLPASSNGLDPVL
 TF4 -----WLD SMLTDLNPP-----SSN-AEYDL

SCR IQNVRDIIFPCNPNLGALLEYRLRSLMLDPSSSDPSPQTFEPLYQISNNPSP
 TF1 PSPEICGFPXSDYDLKVI PXNAIYQFP AIDSSSSNN--Q-----
 TF4 -----KAI-P-----GDILNQF-AIDSASSN--Q-----

FIG. 13A

28/74

SCR	PQQQQHQQQQQHKPPPPPIQQQERENSSDAPPQPETVTATVPVQNTAE	-----
TF1	-----NKRLKSCSSPDSMTSTSTGTQIGGVIGTIVTTTTTTTAAES	
TF4	-----GGGDTYTTNKRLKCSNGVVETTTTATAES	

SCR	----- MOTIF II (DIMERIZATION?) -----	
1110	LREKEIKRQKQDEEGLHLLTLLQCAEAVSADNLEEANKLLEISQLSTPYG	
TF1	-----TRSVILVDSQENGVRVHALMACAEAIQNNLTAEALVKQIGCLAVSQA	LSMVNELRQIVSIQG
TF4	-----TRHVVLVDSQENGVRVHALLACAEAVKENLTVAEALVKQIGFLAVSQI	
3898		QLGKPF

SUBSTITUTE SHEET (RULE 26)

SCR	----- -----	
4818	TSAQRVAAYFSEAM SARLLNSCLGIYAALPSRWMPQTHSLKMVSAFQVFN	GTSP-TGPELLTYMHILYEACP
1110	DPSORIAAYMVEGLAARMAASGFIYRAL-KCKEPPS--DERLAAMQVLF	
TF1	GAMRKVATYFAEALARR-----IY-RL-SPPQNQIDHCLSDTLQMHFYETCP	
TF4	GAMRKVATYFAEALARR-----IY-RL-SPSQSPIDHSLSDTLQMHFYETCP	
3989	-----RSASYLKEALLALADSHHGSSGVT-SPLDVA-----LKLAAYKSFSDLSP	

FIG. 13B

29/74

```

----- MOTIF III (VHIID) -----
LVKFSHTANQAIQAEFEK--EDSVHIIDLDIMQGLQWPGLFHILASRPGGPP-----HVR
YKFGYESANGAIAEAVKN--ESFVHIIDFQISQGGQWVSLIRALGARPGGPP-----NVR
CFKFGFLAANGAILEAIKG--EEEVHIIDFDINQGNQYMTLIRSI AELPGKRP-----RLR
      AMEG--EKMVHVVIDLDASEPAQWLALLQAFNSRPEGPP-----HLR
YLKFAHFTANQAILEAFEG--KKRVHVVIDFSMNQGLQWPALMQALALREGGPP-----TFR
YLKFAHFTANQAILEAFQG--KKRVHVVIDFSMSQGLQWPALMQALALRPGGPP-----VFR
VLQFTNFTANKALLDEIGGMATSCIHVVIDFNLGVGQWASEFLQELAHRRGAGGMALPLLK
      HASVKG--YNHVHIIDFSLMQGLQWPALMDVFSAREGGPP-----KLR
      QEAFER--EERVHIIDLDIMQGLQWPGLFHILASR
      FAG--CRRVHVVDGFKQGMQWPALLXDLAL
      GRNGRTL--WLGECHIDLWPLQGLLSQGLQALCARPLGAP-----HVF-

```

FIG. 13C

SCR
4818
1110
3935
TF1
TF4
3989
18310
Zm-Sc11
Zm-Sc12
Human

30/74

	--- ---	-----	MOTIF	IV (DIMERIZATION)	-----	MOTIF	V
SCR	LTG	LGTSMEA	LQATGKR	LSDFTDK	LGLPFEFCPLAEKVGNLTERLNV		
4818	ITG	IDDPRSSFARQGG	LELVGQR	LGKLAEM	CGVPFEFHGAALCCTEVEIEKLG		
1110	LTG	IDDPESVQRSIGG	LRIINLR	LEQLAED	NGVSFKFKAMPSKTSIVSPSTLGC		
3935	ITG	VHHQKEV	LEQMAHR	LIEEAEK	LDIPFQFNPVVSRDCLNVEQLRV		
TF1	LTG	GIGPPAPDNSDH	LHEVGCK	LAQLAEA	IHVEFEYRGF	VANSLAD	LDASMLELRP
TF4	LTG	GIGPPAPDNFDY	LHEVGCK	LAHLAEA	IHVEFEYRGF	VANTLAD	LDASMLELRP
3989	LTAF	MSTASHHPLE	LHLTQDN	LSQFAAE	LRIPFEFNAVSLDAFNPAESISSGDE		
18310	ITG	GIGNPIGGRDE	LHEVGIR	LAKYAH	VGIDFTFQGVCDQLDRLCDWMLLKPI		
Human	LPGL	HTLS...	LGLQXRH	LLVHMMA	LSYSYGRXP...		

SCR	RKREA	VHQLQHS	LYDVTGSDAHTLWLL	---QRLAPK---
4818	RNGEAL	AVNFPLVLH	HMPDES	VTVENHR---DRLLRL---
1110	KPGETL	VNFAFQLH	HMPDES	VTVNQR---DELLHM---
3935	KTGEAL	AVSSVLQLHT	FLASDDDL	MRKNC-ALRFQNNPSGVDLQRLMMSHGS
TF1	SDTEA	AVAVNSVFEL	HKLLGRXGGIEKVLG	-----
TF4	SEIESV	AVNSVFEL	HKLLGRPGAIDKVLG	-----
18310	K-GEA	VAIN	SILQLHRL	LLVDPDANPVVPAPIDILK---
3989	VWAV	SLPVGCSA	RAPPLPAILRL	VKQLCPKVVAID

FIG. 13D

31/74

```

|-----
-----VTV-
-----VKHSPN-VVTL-
-----VKSLNPK-LVTV-
AAEARENDMSNNNGYSPSGDSASSLPSPSSGRDTSFLNAIWGLSPKVMVVT-
-----VVKQD*TGDFHXW
-----VWNQIKPEIFTV-
-----LVIKINPMIFTV-

```

SCR
4818
1110
3935
TF1
TF4
18310

```

----- MOTIF VI -----
VEQDLSHAGS--FLG-RFVEAIHYYSALFDSLGSYGEESE---ERHVVEQQ
VEQEANTNTAP-FLP-RFVETMNHYLAVFESIDVKLARDHK---ERINVEQH
VEQDVNTNTSP-FFP-RFIEAYEYYSAVFESLDMTLPRESQ---ERMNVERQ
-EQDSDHNGS--TLMERLLESLYTYAALFDCLETKVPRTSQ---DRIKVEKM
XRQEPNHNG-PGFLD-GXTESLHYYSTXFDSELEG--XPNSQ---DKLMSEXY
VEQESNHNS-PIFLD-RFTESLHYSTLFDSLEG--VPSGQ---DKVMSEVY
VEHEADHNR-PPLLE-RFTNALFHYATMFDSLEAMHTCTSGRDITDSLTEVY

```

SCR
4818
1110
3935
TF1
TF4
18310

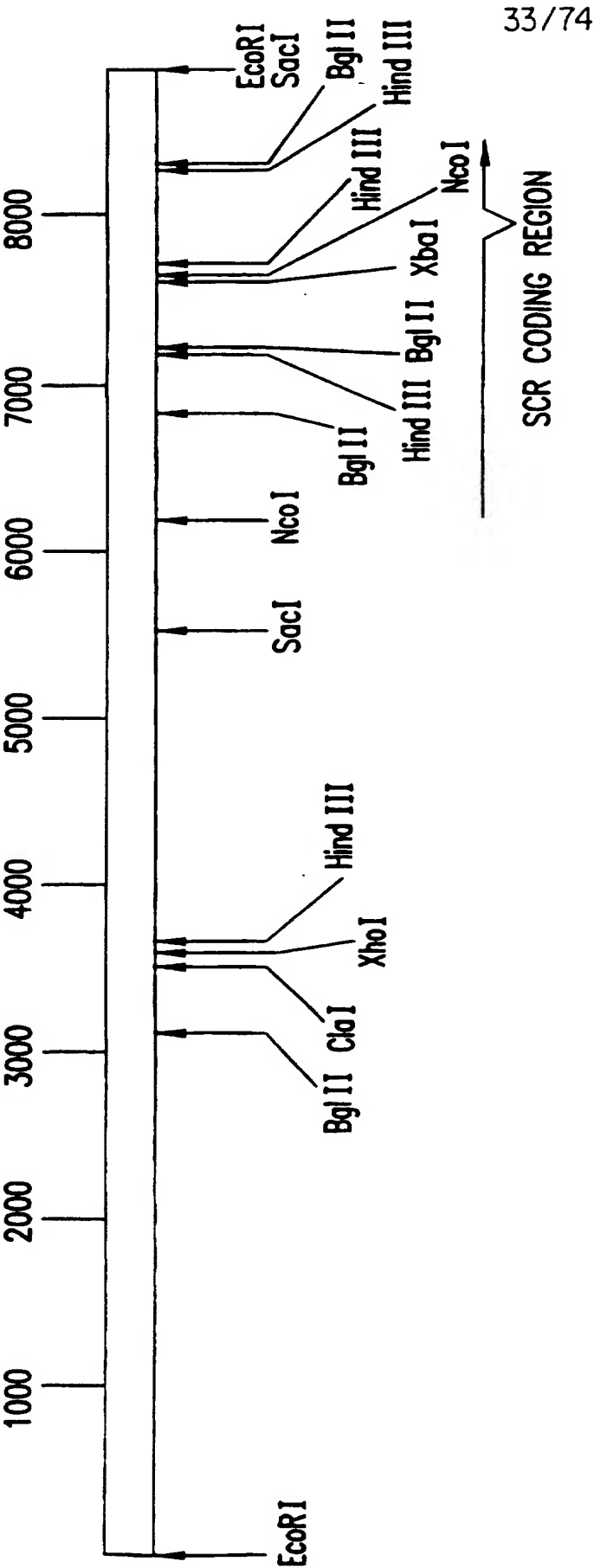
FIG.13E

SCR LLSKEIRNVLAVGGPSRSGEVKFE-SWREKMQQCGFKGIS-
4818 CLAREVVNLIACEGVEREERHEPLGKWSRFRHMA GFKPYP-
1110 CLARDIVNIVACEGEEERIEREYEAAGKWRARMMAGFNPKP-
3935 LFGEEIKNIIISCEGFERRERHEKLEKWSQRIDLAGFGNVP-
TF1 -LGXQICNLVACEGPD RVERHETLSQWGNRFGSSGLAPAH-
TF4 -LGKQICNVVACDGPDRVERHETLSQWRNRFGSAGFAAAH-
18310 -LRGEIFDIVCGESARTERHELFGHWRERLTYAGLTQVWF

-----|-----
SCR LAGNAATQATLLGMFPS-DGYTLVDDN-GTLKLGWKDLSLLTASAWTPRS*
4818 LSSVYNATIKG LLES-YS-EKYTL-EERD GALYLGWKNQPLITSCAWR*
1110 MSAKVTNNIQNLIKQYC-NKYKLKEEM-GELHFCWEEKSLIVASAWR*
3935 LSYYAMLQARRLLQGCGF-DGYRIKEES-GCAVICWQDRPLYSVSAWRCRK*
TF1 LGSNAFKQASMLLSVFNSGQGYRV-EESNGCLMLGWHTRPLITTSAWKLSTAAH*
TF4 IGSNAFKQASMLLALFN GGE GYRV-EESD GCLMLGWHTRPLIATS AWKLSTN*
3989 ADCLL-KRVQVRGFHV-EKRG AALTYWQRGELVSISSWRC*
18310 DPDEVDTLKDQLIHVTSLSGSGFNILVCDGSLALAWHNRPLYVATAWCVTGGNAA

FIG. 13F

18310 SSMVGNICKGTNDSRRKENRNGPME*



SacI GENOMIC FRAGMENT

HIII-SacI CLONE

FIG.14

34/74

Old Name	New Name
SCR	SCR
3989	SRPo3
12398	SRPa6
4871	SRPa5
11846	SRPo4
2504	SRPo2
3935	SRPa3
11261	SRPa10
713	SRPo1
10964	SRPa9
23196	SRPa12
Tf1	SRPa8
Tf4	SRPa2
18310	SRPm1
18652	SRPa11
4818	SRPa4
21729	SRPa7
1110	SRPa1
174	SRPb1
33/08	SRPa13
-150	-101

FIG. 15A

36/74

```

Scr
3989
12398
4871
11846
2504
3935
11261
713
10964
23196
Tf1
Tf4
18310
18652
4818
21729
1110
174
33/08

```

FIG. 15C

37/74

SCF	MAESGDFNGG	QPPPHSPLRT	TSSGSSSSNN	RGPPPPPPPP	LVMVRKRLAS
3989
12398
4871
11846
2504
3935
11261
713
10964
23196	CMFHDALALQ	AAEKSLYEAL	GEKDPSSSSA	SSVDHPERLA	SHSPDGCSCG
Tf1
Tf4
18310
18652
4818
21729
1110
174
33/08TSDSA	SSFNIPTSAQ	NHYATGSFST
	1				50

FIG.15D

38/74

	←-----Motif I-----→	
Scr	EMSSNPDYNN SSRPPRRVSH LLDSNYNTVT PQQPPSLTAA ATVSSQPNPP	
3989
12398
4871
11846
2504
3935
11261
713
10964
23196	GAFSYASTT TTTSSDSHWS VDGLNRPSPW LHTPMPNSFV FQSTSRNSV	
Tf1
Tf4	.MKRDHHQFQ GRLSNHTSS SSSSISKDKM MMVKKEEDGG	
18310QDKK TMMNEEDDG
18652
4818
21729
1110
174
33/08	NSRTTNVATA TTNSATAHWV ATDAEHTDTI IAQP	
	51	100

FIG. 15E

39/74

SCR	LSVCGFSGLP	VFPSDRGGRN	VMMSVQPMQ	DSSSSASPT	VWVDIIRDL
3989
12398
4871
11846
2504
3935
11261
713
10964
23196	TGGGGGNSA	VYGSFGDDL	VSNMFKDDEL	AMQFKKVEE	ASKFLPKSSQ
Tf1	GNMDELLAV	LGKVRSSSEM	AEVALKLEQL	ETMMSNAQED	GLSHLATDAA
Tf4	NGM.DELLAV	LGKVRSSSEM	ADVAQKLEQL	EVMMSNVQED	DLSQLATETV
18310
18652
4818
21729D
1110
174
	101				150

FIG.15F

SCI	IHSSTSVSIP	QLIQNVRDII	FPCNPNLGAL	LEYRLRSLML	LDPSSSSDPS
3989
12398
4871
11846
2504
3935
11261
713
10964
23196	LFIDVDSYIP	MNSGSKENG	EVFVKTEKD	ETEHHHHSY	APPPNRLTGK
Tf1	HYNPSELYSW	LDNMLSELNP	PPLPASSNGL	DPVLPSPEIC	GFPXSDYDLK
Tf4	HYNPAELYTW	LDSMLTDLNP	P.....SSNA.EYDLK
18310
18652
4818
21729	LTSVNDMSLF	GGSGSSQRYG	LPVPRSQTQQ	QQSDYGLFGG	IRMGIGSGIN
1110
174
151
					200

FIG.15G

41/74

Scf	PQTFEPLYQI	SNNPSPQQQ	QQHQQQQQQH	KPPPPPIQQQ	ERENSSDAP
3989
12398
4871
11846
2504
3935
11261
713
10964
23196	KSHWRDEDED	VEERSNKQSA	VYVEEELSE	MFDNMFLCGP	GKPVCIILNQ
Tf1	VIPXNAIYQF	PAIDSSSSN	NQ.....	NKRLKSCSSP	DSMVTSTG
Tf4	AIPGDAILNQ	FAIDSASSN	QGGGDTYTT	NKRLKCS...
18310
18652
4818
21729	NYPTLTGVPC	IEPVQNRVHE	SENMLNSLRE	LEKQLDDDD	ESGDDDDSV
1110
174
201
					250

FIG. 15H

42/74

	←--- bZIP like domain ---	
	←--- Motif II (dimerization)	-----
Scf	PQPETVTATV PAVQTNTAEA LRRKEEIKR QKQDEEGLHL	LTLLQCAEA
3989
12398
4871AAIFYG HHHHTPPPAK	RLNPGPVGIT
11846
2504
3935
11261
713
10964
23196	NFPTESAKV TAQSNQAKIR GKKSTSTSHS	NDSKKETADL RTLLVLCAQA
Tf1	TQIGGVIGTT VTTTTTTTA AEESTRSVIL	VDSQENGVRV VHALMACAEA
Tf4	...NGVVE.. .TTTA TAESTRHVVL	VDSQENGVRV VHALMACAEA
18310
18652
4818
21729	ITNSNSDWIQ NLVTPNPFPN PVLSPSPSS	SSSSSPSTAS TTTSVCSRQT
1110
174
251	300

FIG.15I

43 / 74

	----- Motif II (dimerization) ----->	
SCR	VSADNLEEAN KLLLEISQLS TPYGTSQORV AAYFSEAMSA RLLNSCLGIY	
3989
12398
4871	EQLVKAAEVI ESDTCLAQGIL ARLNQQLSS PVGKPLERAA FYFKEALNNL	
11846
2504
3935
11261
713
10964
223196	VSVDRRRTAN EMLRQIREHS SPLNGSERL AHYFANSLEA RLAGTGQTQIY	
Tf1	IQQNNLTAE ALVKQIGCLA VSQAGAMRKV ATYFAEALAR RIYRLSP PQN	
Tf4	VQKENLTVAE ALVKQIGFLA VSQIGAMRQV ATYFAEALAR RIYRLSPSQS	
18310
18652
4818GT
221729	VMEIATAIAE GKTEIATEIL ARVSQTPNLE RNSEEKL VDF MVAALRSRIA	
1110	...LSMVNEL RQIVSIQ GDP SQRIAYMVE GLAARMAASG KFIYRALCKCK	
174
	301	350

FIG. 15J

44/74

SCR				←--- Motif III (SCR VHIID) ---	
3989	AALPSRWMPQ	THSLKMVSFA	QVFNGISPLV	KFSHFTANQA	IQEAFEKEDS
12398LYRNKALL	DEIGGMATSC
4871	LHNVSQTLA	CSLIFKVAAY	KSFSEISPLV	QFANFTSNQA	LLESFHGFHR
11846
2504
3935AMEGEKM
11261
713
10964
23196	TALS...SKK	TSAADMLKAY	QTYMSVCPFK	KAIIIFANHS	MMRFTANANT
Tf1	QIDHCLSDT.LQ	MHFYETCPYL	KFAHFTANQA	ILEAFEGKKR
Tf4	PIDHSLSDT.LQ	MHFYETCPYL	KFAHFTANQA	ILEAFQGKKR
18310HA	SVKGYN...H
18652ANVE	ILEAIAGETR
4818	SPTGPELLT.YM	HILYEACPYF	KFGYESANGA	IAEAVKNESF
21729	SPVTELYGKE	HLISTQL...	..LYELSPCF	KLGFEEAANLA	ILDAADNNDGGMMI
1110	EPPSDERLA.AM	QVLFEVCPCF	KFGFLAANGA	ILEAIKGEET
174
	351				400

FIG. 15K

45/74

```

-- Motif III (VHIID) ----->|+ -- Motif IV ---
VHIIDLDIMQ GLQWPGLFHI LASRPGGPPH VRLTGLGTSM EA.....LQ
IHVIDFDLGV GGQWASFLQE LAHRRGAGGM ALPLLKLTAF MSTASHHPLE LH
.....
LHIIDFDIGY GGQWASLMQE LVLRDNAAPLSLKITVFASPA NHVQLELG..
.....
.....
VHVIDLDASE PAQWLALLQA FNSRPEGPPH LRITGVHHQK EVLE.....
.....
.....
.....
IHIIDFGISY GFQWPALIHRLSLSRPGGSPK LRITGIELPQ RGFRPAE....
VHVIDFSMNQ GLQWPALMQA LALREGGPPT FRLTGIGPPA PDNSDHLH..
VHVIDFSMSQ GLQWPALMQA LALRPGGPPV FRLTGIGPPA PDNFDYLH..
VHIIDFSLMQ GLQWPALMDV FSAREGGPPK LRITGIGPNP IGGRDELH..
VHIIDFQIAQ GSQYMFLIQE LAKRPGG... ...PPLLRVT GVDDSQSTYARGGGLS
VHIIDFQISQ GGQWVSLIRA LGARPGG... ...PPNVRLT GIDDPRSSFARQGGLE
VHIIDFDINQ GNQYMTLIRS IAELPGK... ...RPRLRLT GIDDPESVQRSIGGLR
PHVIDFDIGE GGQYVNLRLT LSTRNGKSQ SQNSPVVKIT AVANNVGDCLVDDGGGEERLX
.....
401
450

```

FIG.15L

46/74

	--- Motif IV ---	← --- Motif V ---
Scr	ATGKRLSDFT	DKLGLPFEFC
3989	LHLTQDNLSQ	FAAELRIPFE
12398	FTQDNLKHFA	SEINISLDIQ
4871	VL..SDDL	LG
11846	QMAHRLIEEA	EKLDIPFQFN
2504	QMAHRLIEEA	EKLDIPFQFN
3935	QMAHRLIEEA	EKLDIPFQFN
11261	QMAHRLIEEA	EKLDIPFQFN
713	QMAHRLIEEA	EKLDIPFQFN
10964	QMAHRLIEEA	EKLDIPFQFN
23196	QMAHRLIEEA	EKLDIPFQFN
Tf1	QMAHRLIEEA	EKLDIPFQFN
Tf4	QMAHRLIEEA	EKLDIPFQFN
118310	QMAHRLIEEA	EKLDIPFQFN
118652	QMAHRLIEEA	EKLDIPFQFN
4818	QMAHRLIEEA	EKLDIPFQFN
21729	QMAHRLIEEA	EKLDIPFQFN
1110	QMAHRLIEEA	EKLDIPFQFN
174	QMAHRLIEEA	EKLDIPFQFN

FIG. 15M

47/74

	----- Motif V -----	
Scr	...QHS...
3989P VG.
12398
4871AA.
11846
2504R SL.
3935	LQLHTFLASD DDLMPKNCAL RFHNPPSGVD LQRVLMMSGH SAAEARENDM	
11261	NCIHRLLQYTP DE.
713
10964
23196	FRFRNLL... DE.
Tf1	FELHKLLGRX GG.
Tf4	FELHKLLGRP GA.
18310	LQLHRLLVDP DA.
18652	YVLHMH...P DE.
4818	LVLHMH...P DE.
21729	FKLYRV...P DE.
1110	FQLHMH...P DE.
174
	501	550

FIG. 15N

48/74

	Motif V	----->		←--- Motif VI	---
Scr	LYDVTGSD	AHTLWLLQRL	APKVVTVVEQ
3989	CSARAPPL	PAILRLVKQL	CPKVVAIDH
12398	GGDRADLPFS
4871
11846	SFSLPLV	LRFVKHLSPT	IIVCSDRGCE
2504	RTDLPFSQQL
3935	WTARSLVPSSPST	DSF.....
11261	DSASSLPSSGRT	DSFLNAIWGL	SPKVMVTEQ
713	DSDHNGSTLM
10964	TVSLDSPR	DTVCLKFRDI	NGMYNSPFFM
23196	NGSYNAPFFV
Tf1
Tf4AYNAPFFV
18310	TVLVNSPR	DAVLKLIRKI	SGNYNAPFFV
18652	EPNHNGPGFL
4818	KGPEIFTVVEQ
21729	NPVVPAPI	DILKLVIKI	NPMIFTVVEH
1110	EADHNRPPLL
174	SVSVEKYR	DRLLHLIKSL	EDTNTSPLV
	SVTVENHR	DRLLRLVKHL	SPNVVTLVEQ
	SVCTENPR	DELLRRVKGL	EANTNTAPFL
	SVTTVNQR	DELLHMKSL	EMNSNTAPFL
	DVNTNTSPFF

	551				600

FIG.150

49/74

```

----- Motif VI -----
GRFVEAIHY SALFDSLGA Y..GESEER HVEQQLSK EIRNVLA VGG
QHFLNCFQSC VFDSLDAAG I..DADSA.. CKIERFLIQP RVEDAVIG..
..... SLEPN L..DRDSKER LRVERVLFG RIMDLVRSDD
AHSLSHTAL FESLDAVNAN L..DAM.... QKIERFLIQP EIEKLVLD..
DRFTEALFYY SAVFDSLDA N..NNNNNNN QRMEEAYLQR EICDIVCGEG
.....
ERLLESLYTY AALFDCLETK V..PTSQDR IKVEKMLFGE EIKNIIISCEG
TRFREALFHY SSLFDMFDTT IHADEYKNR SLLERELLVR DAMRVISCEG
TRFREALFHY SAIFDMLETN I..PKDNEQR LLIESALFSR E.XNVISCEG
TRFREALFHP SSIFDMLETI V..PREDEER MFLEMEVFGR EALNVIACEG
TRFREALFHY SAVFDMCD SK L..AREDEMR LMVVFEPFYGR EIVNVVASEG
DGXTESLHY STXFDSLEGX ...PNSQD.. KLMSEXYLGX QICNLVACEG
DRFTESLHY STLFD SLEGV ...PSGD.. KVMSEVYLGK QICNVVACDG
ERFTNALFHY ATMFD SLEAM HRCTSGRDIT DSLTEVYLRG EIFDIVCGEG
SRFVETLDY TAMFESIDAA R..PRDDKQR ISAEQHCVAR DIVNMIACEE
PRFVETMNHY LAVFESIDVK L..ARDHKE R INVEQHCLAR EVENLIACEG
GRVSESCACY GALLESVES T V..PSTNSDR AKVE.EGIGR KLVNAVACEG
PRFIEAYEY SAVFESLDM T L..PRESQER MNVERQCLAR DIVNIVACEG
..... .RXFDSLEHD A..SKGEPRE DERGRXCLAR NIVNIVXCKX
601 650

```

FIG.15P

50/74

SCI	PSRSGEVKF.ESWRE	KMQQCGFKGI	SLAG..NAAT	QATLLGMFEP
3989	.RHKA..Q..	...KAIAWRS	VFAATGFKPV	QLSN..LAEA	QADCLLKRVO
12398	DNNKPGTRFG	LMEKEQWRV	LMEKAGFEPV	KPSN..YAVS	QAKLLWNYN
4871	.RSRPIER..	...PMTWQA	MFLQMGFSPV	THSN..FTES	QAECLVQRTF
11846	AARXERHE..	...PLSRWRD	RLTRAGLSAV	PLG....SNA
2504
3935	FERRERHE..	...KLEWSQ	RIDLAGEFNV	PLSY..YAML	QARRLLQCGG
11261	AERFARPE..	...TYKQWRV	RILRAGFKPA	TIS....KQI	MKEAKEIVRK
713	LERMERPE..	...TYKQWQV	RNQRVGFQQL	PLN....QDM	MKRARXEGQV
10964	WERVERPE..	...TYKQWHV	RAMRSGLVQV	PFD....PSI	MKTSLHKVHT
23196	TERVESRE..	...TYKQWQA	RLIRAGFRQL	PLE....KEL	MQNLKLKIEN
Tf1	PDRVERHE..	...TLSQWGN	RFGSSGLAPA	HLGS...NAF	KQASMLLSVF
Tf4	PDRVERHE..	...TLSQWRN	RFGSAGFAAA	HIGS...NAF	KQASMLLALF
18310	SARTERHE..	...LFGHWRE	RLTYAGLTQV	WFDPEVDTL	KDQLIHVTSL
18652	SERVERHE..	...VLGKWRV	RMMAGFTGW	PVSTSAFAA	SE....MLK.
4818	VEREERHE..	...PLGKWS	RFHMAGFKPY	PLSSYVNATI	KG....LLE.
21729	IDRIERCE..	...VFGKWRM	RMSMAGFELM	PLSEKIAESM	KS....RGNR
1110	EERIERYE..	...AAGKWRA	RMMAGFNPK	PMSAKVTNNI	QN....LIKQ
174	EERIERYE..	...VTGKWRA	RMMAGFSPR	PMSGRVTSNI	ES....LIKR
	651				700

FIG.15Q

51/74

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----- Motif VI ----->
Scr      .SDGYTLVD. DNGTLKLGWK DLSLLTASAW TPRSX.....
3989     VRGFH..VEK RGAALTLYWQ RGELVSISSW RCX.....
12398    YSTLYSLVES EPGFISLAWN NVPLLTVSSW RX.....
4871     VRGFH..VEE KHNSLLLCWQ RTELVGVS AW RCRSSX....
11846    .....
2504    .....
3935     FDGYR..IKE ESGCAVICWQ DRPLYSVSAW RCRKX.....
11261    RYHRDFVIDS DNNWMLQGWK GRVIYAFSCW KPAEKFTNNN LNIX.....
713      LPTRTFIIDE DNRWLLQGWK GRILFALSTW KPDNRSSSX. ....
10964    FYHKDFVIDQ DNRWLLQGWK GRTVMALSVW KPESX.....
23196    GYDKNFDVDQ NGNWLQGWK GRIVYASSLW VPSSSX.....
Tf1      NSGQGYRVEE SNGCLMLGWH TRPLITTS AW KLSTAAHX.. ....
Tf4      NGGEGYRVEE SDGCLMLGWH TRPLIATSAW KLSTNX.....
18310    .SGSGFNILV CDGSLALAWH NRPLYVATAW CVTGGNAASS MVGNICKGTN
18652    AYDKNYKLG GEGALYLFWK RRP MATCSVW KPNPNYIGX. ....
4818     SYSEKYTLEE RDGALYLGWK NOPLITSCAW RX.....
21729    VHPG.FTVKE DNGGVCFGWM GRALTVASAW RX.....
1110     QYCNKYKLKE EMGELHFCWE EKSLIVASAW RX.....
174      DYCSKYKVKE EMGELHFSWE EKSLIVASAW SX.....
701
750

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FIG.15R

SCR
3989
12398
4871
11846
2504
3935
11261
713
10964
23196
Tf1
Tf4
18310	DSRRKENRNG	PMEX
18652
4818
21729
1110
174
	751	764

FIG. 15S

53/74

SRPa1 (1110)

CTTTGTCAAATGGTAAATGAGCTGAGGCAGATAGTTTCTATCCAAAGGAGACCCCTTCTCAGA
GAATCGAGCTTACATGGTGAAGGTCTAGCTGCAAGAAATGGCCGCTTCAGGAAATTCATCA
TCTACAGAGCATTTGAAATGCAAGAGCCCTCTCGGATGAGAGGCTTGCGAGCTATGCAAG
TCCTGTTTGAAGTCTGCCCTTGTTTCAAGTTCCGGGTTTTTAGCAGCTAATGGTGGATAC
TTGAAGCAATCAAAGGTGAAGAAGTTCAATATCGATTTCGATTAATAACCAAGGGA
ACCAATACATGACACTGATACGAAGCATTTGCTGAGTTGCCCTGGTAAACGACCTCGCCTGA
GGTTAACAGGAATTGATGACCCCTGAATCAGTCCAAACGCTCCATTTGGAGGCTAAGAATCA
TCGGTCTAAGACTCGAGCAACTCGCAGAGGATAATGGAGTATCCTTCAAAATTCAAAGCAA
TGCCTTCAAAGACTTCGATTGTCTCTCCATCAACACTCGGTTGCAACCCAGGAGAAACCT
TAATAGTGAACTTTGCATTCCAACTTCACCACATGCTGACGAGAGTGCACACAGTAA
ACCAGCGGACGAGCTACTTCACATGGTCAAAAGCTTAACCCAAAGCTTGTACGGTCG
TTGAACAAGACGTGAACACAACACTTCACCGTTCTTTCCAGATTTCATAGAGGCTTACG
AATACTACTCAGCAGTTTTTCGAGTCTCTAGACATGACACTTCCAAGAGAAAGCCAAAGAGA
GGATGAATGTAGAAAGACAGTGTCTCGCTAGAGACATAGTCAACATTGTTGCTTGCGAAG
GAGAAACGGATAGAGAGATACGAGGCTGCGGGAAATGGAGAGCAAGGATGATGATGG
CTGGATTCAATCCAAACCAATGAGTGCTAAAGTAACCAACAATATACAAAACCTGATAA
AGCAACAATAATTGCAATAAGTACAAGCTTAAGAGAAATGGGTGAGCTCCATTTTGCT
GGAGGAGAAAGCTTAATCGTTGCTTCAGCTTGAGGTAAGATAAGTGACAAGAGCATA
TAGTCTTTATGTTTCATAAAACATAATTATGTTTTTACTGTAATCTTGGGTTATTGTGTA
ACTGGTTAAATCATCTCCATGTATTATTACCAGAGGTTAGGGTGATCACAGGTACTAAA
AGCTAATCTAACACTTATGGAAGAATTTTCTTCTTTTCTTTTCCCTATTATATAAAAT
AATTAGAGTTTTTGGTCTAAACCTATTTCCTAAGTGTGAATGAGTCTTTACATGTTTCATA
TTTCAGTTCAAATGGTTAAATTGTGTTAAGGTTCTCACTTAAAAAAAA

FIG. 16A

54/74

SRPa3 (3935)

GCTATGGAAGGAGAGATGGTTCACTGTGATTGATCTCGATGCTTCTGAGCCAGCTCAA
TGGCTTGCTTTGCTTCAAGCTTTTAACTCTAGGCCCTGAAGTCCACCCTCATTTGAGAAATC
ACTGGTGTTCATCACAGAGGAGAGTGCTTGAAACAAATGGCTCATAGACTCATTGAGGAA
GCAGAGAAACTCGATATCCCGTTTCAGTTTAAATCCCGTTGTGAGTAGGTTAGACTGTTTA
AATGTAGAAACAGTTGGGGTTAAACAGGAGAGGCCCTTAGCCGTTAGCTCGGTTCTTCAA
TTGCATACCTTCTTGGCCCTCTGATGATCTCATGAGAAAGAACTGCGCTTTACGGTTT
CAGAAACAACCTAGTGGAGTTGACTTGCAGAGAGTTCTAATGATGAGCCATGGCTCTGCA
GCTGAGGCACGTGAGAAATGATAGTAACAACAAATGGGTATAGCCCTAGCGGTACTCG
GCCTCATCTTTGCCCTTACCAAGTTCAGGAAGGACTGATAGCTTCCCTCAATGCTATTGG
GGTTTGTCTCCAAAGGTCATGGTGGTCACTGAGCAAGACTCAGACCAACGAGCTCCACA
CTAATGGAGAGGCTATTAGAAATCACTTTACACCTACGCAGCATGTTTGTGATTGCTGGAA
ACAAAGTTCCAAGAACGCTCAAGATAGGATCAAGTGGAGAGATGCTCTTCGGGGAG
GAGATCAAGAACATCATATCCGCGAGGATTTGAGAGAGAGAAAGACACGAGAAAGCTT
GAGAAATGGAGCCAGAGATCGATTTGGCTGGTTTGGGAATGTTCCCTTAGCTATTAT
GCGATGTTGCAGGCTAGGAGATTGCTTCAAGGGTGGGTTTGTGATGGGTATAGAATCAAG
GAAGAGAGCGGTGCGCAGTAATTGCTGGCAAGATCGACCTCTATCTCGGTATCAGCT
TGGAGATGCAGGAAGTGAATGATATATTACAGTTTGTCTTCTATTTTGGTTATGAGCAGA
GTCCCTTCTTTTGTATACATGGGGACACAATCTTAGTTGTTTGTGATGGTGACTTT
CTGTCTCTTTATGCTATTTGGCTTAAATGCTTCTACTGCCCTCTGCATGTAAAGCCTTG
TGTGTTGGTTCAATTTGGTCTGGTGTGGGTGTAATACCAACCAATCCAATTTGAGCTG
AAGATAACTAATTGTGATGATCGGCTCGTGCC

FIG. 16B

55/74

SRPa4 (4818)

GCACGAGCCCAACGGGTCCTGAGCTTCTTACTTATATGCATATCTTGTATGAAGCCTGC
CCTTATTCAAAATTCGGTTATGAATCTGCTAATGGAGCTATAGCTGAAGCTGTGAAGAAC
GAAAGTTTGTGCACATTATCGATTTCAGATTCTCAAGTGGTCAATGGGTGAGTTTG
ATCCGTGCTCTTGGTCTAGACCTGGTGACCTCCGAACGTTAGGATAACGGGAATTGAT
GATCCGAGATCATCGTTTGCTCGTCAAGGAGGACTTGAGTTAGTTGGACAAAGACTTGGG
AAGCTAGCTGAAATGTGCGGTGTTCCGTTTGAGTTCCATGGAGCTGCTTATGCTGCACG
GAAGTCGAAATCGAGAACTAGGAGTTAGAAATGGAGAACGCTCGCGGTTAACTTCCCG
CTTGTTCTTCACCATGCTGATGAGAGTGTAACGTTGTGACTCTGGTTGAGCAAGAGCGAAT
TTCGAACTGCGCGGTTTCTTCCCGGTTTGTGAGACAAATGAACCATTACTTGGCAGTT
TTCGAACTCAATAGATGTGAACCTCGCTAGAGATCAAGGAAGGATCAATGTTGAGCAG
CATTGTTTGGCTAGAGAGGTTGTGAATCTTATAGCTTGTGAAGGTTGAAAGAGAAGAG
AGGCACGAGCCACTAGGGAATGGAGGTCCTCGGTTTCACATGGCGGGATTAAACCGTAT
CCTTTGAGCTCGTATGTGAACGCAACAATCAAAGGATTGCTTGAGAGTTATTCAGAGAAAG
TATACACTTGAAGAAAGAGATGGAGCATTTGTATTAGGATGGAAGAATCAACCTCTTATC
ACTTCTTGTGCTTGGAGGTAACTAATAAAACCTTGTTCGGTTTCAGAAAGAGATTAGAAA
CTTCTTTAAAGTTTGCAGAACTGTGTTGTAAAAGTAAACTCATGTCATGATCCGNAGGA
ACAAGTTGTCAAATGTTGTAGTAGTAAGTGATATGTTGATGACCCAAAATAAAAAA
AAAAA

FIG. 16C

56/74

SRPa5 (4871)

GGGGCTATCTTACGGCCACCACCATACACCTCCGCCGGCAAGCGGCTCAACCCCT
GGTCCCGTGGGATAACAGAGCAGCTGGTTAAGGCAGCAGAGGTCAATAGAGCGACACG
TGCTAGCTCAGGGGATATTGGCGCGGCTCAATCAACAGCTCTCTTCTCCGTCGGGAG
CCATTAGAAAGAGCAGCTTTTACTTCAAGAAGCTCTCAATAATCTCCTTCACAAAGTC
TCCCAAACCCCTAAACCCCTTATTCCTCATCTTCAAGATCGCTGCTTACAAATCCTTCTCA
GAGATCTCTCCGTTCTTCAGTTCGCCAACHTTACCTCCAACCAAGCCCTCTTAGAGTCC
TTCCATGGCTTCCACCGTCTCCACATCATCGACTTCGATATCGGCTACGGTGCCAAATGG
GCTTCCCTCATGCAAGAGCTTGTCTCCGGACAACGCCGCTCTCTCTCCCTCAAGATC
ACCGTTTTCGCTTCTCCGGCGAACCAAGACAGCTCGAACTTGGCTTCACTCAAGACAAC
CTCAAGCACTTCGCTCTGAGATCAACATCTCCCTTGACATCCAAGTTTGGAGCTTAGAC
CTCCTCGGCTCCATCTCGTGGCCTAACTCGTCGGAGAAAGCTGTCGCCGTTAAACATC
TCCGCCGCTCCTTCTCGCACCTCCCTTTGGTCTCCGTTTCGTGAAGCATCTATCTCCG
ACGATCATCGTCTGCTCCGACAGAGGATCGAGAGGACGGATCTGCCCTTCTCTCAACAG
CTCGCCCACTCGTGCACTCACACCGCTCTCTTCGAATCCCTCGACGCCGTCAACGCC
AACCTCGACGCAATGCAGAAAGATCGAGAGGTTTCTTATACAGCCGGAGATAGAGAGCTG
GTGTTGGATCGTAGCCGTCCGATAGAAAGGCCGATGATGACGTGGCAAGCGATGTTTCTA
CAGATGGGTTTCTCACC GTGACGCACAGTAACCTTACGGAGTCTCAAGCCGAGTGTTTA
GTCCAACGGACGCCAGTGAGAGGCTTTCACGTCGAGAAAGAAACATACTCACTCTCCTA
TGTTGGCAAGGACAGAACTCGTCGGAGTTTCAGCATGGAGATGTCGCTCCTGATTT
CCACCGGAGTTTCAATTATTAAAAAATAATTTTCCCTTAATCAATTTATCTTAAATGACA
AATTTTAGTTTCTGATTTTATTTTGCTCAGTGGATTTTAAATTTAAGTTTCAC
ACAAATATATAAATTTT

FIG. 16D

57/74

SRPa6 (12398)

AATCGCTTGAAACCGAATTGGATCGAGATTGGAAGAAGGCTGAGAGTGGAGAGAGTGC
TGTCGGTAGGAGGATTATGGATTGGTCCGATCAGATGATGATAATAAACCGGAA
CCCGTTTGGGTTAATGGAGGAGAAAGAACAAATGGAGAGTGTGATGGAGAAAGCTGGAT
TTGAGCCGGTTAAACCGAGTAATTACCGGTTAGCCAGCGAGCTGCTACTATGGAACT
ACAATTATAGTACATTGTATTCACTTGTGAATCGGAGCCAGGTTTCATCTCCTTGGCTT
GGAACAATGTGCCCTCCTCACCGTTTCCTTGGCGTTGACTACTTGGTCCGATAAGTT
AATCTAGTATTTTGAGTTAGCTTTTAGAATTGAATTGTTTGGGTTAGATTGGATGTTT
AATAGTCTCTAGCCATTCTCTACTCTTTTGTCTAGTGCTTGGAGTGATGATGGTT
TGTCGTTTATGTTTCATTGTAAATATATTGTATGTAAACATTGACTAAAAAATAAAA
AAAAAAA

FIG. 16E

58 / 74

FIG. 16F-1

SRPa7 (21729/3635/17410)

AAAGACTTAGCAGATTTTCAAGCGGCTCAGAAACATCAACAACAACAACAACCG
TTTTATAGTCAAGCAGCTCTCAACGCTTTTCTTCAAGTCTGTGAAGCCTCGAAATTAT
CAGAAATTTCAATCTCCGTCCGCCGATGATTGATCTCACGTCGGTGAATGATAGTTT
GTTTGGTGTCTGGTTCATCTCAGCGTTACGGTTTACCGGTTCCAGGTCCTCAGACGCA
ACAGCAACAATCGGATTACGGTTTATTGGTGGGATCCGAATGGGAATCGGGTCGGGTAT
TAATAATTATCCAAACATTAAACCGGCTTCCGTGTATTGAACCGGTTCAAAACCGGTTCA
TGAATCGGAGAAACATGTTGAATAGTTTAAGAGAGCTTGAGAAACAGCTTTTAGATGATGA
CGATGAGAGTGGTGATGATGACGTGTACGTTATAACAAATTCAAATTCGATTGGAT
TCAAAAATCTCGTGAATCCGAACCCGAACCCGTTTGTCTTTTTCACCGAGCTC
TTCTTCTCGTCTTCTCGCCTTCTACAGCTTCGACGACGACATCGGTATGTTCTAGGCA
AACGGTTATGGAATCGCGACGGCGATCGCGAAGGAAACAGAGATAGCGACGGAGAT
TTTGGCGGTGTTTCTCAACCGCCTAATCTTGAGAGGAATTCAGAGGAGAACTTGTGTTGA
TTTCATGGTGGCTGCGCTTCGATCGAGGATAGCTTCTCCAGTGACGGAATTGTATGGGAA
CGAGCATTTAATCTCGACTCAATTGCTCTACGAGCTCTCTCCTGTTCCTCAAACTCGGTTT
CGAGCGCGGAATCTCGCCATTCTCGACGCCCGCGATAACAACGAGGTGGAATGATGAT
ACCGCACGTTATCGATTTCGATATCGGAGAAGGTGGACAATACGTTAACCTTCTCCGTAC
ATTATCCACGCCCGGAATGGTAAAGTCAGAGTCAGAAATCTCCGGTGAAGAGGTTAA
CGCCGTGGCGAACAACGTTACGGATGTTAGTCGATCGACTCGGTATCTCCGTAAGTTTCAA
AGCCGTGGAGATTGTTGAGCCAACTCGGTGATCGACTCGGTATCTCCGTAAGTTTCAA
CGTGGTGACGAGTTTACGACTCGGTGATCTGAATCGTGAATCTCTCGGTGATCCCGA
CGAGACTTTGGCTGTGAACCTTAGCTTTCAAGCTTTATCGTGTTCGCCGAAAGCGTATG
CACGGAGAAATCCAAGAGACGAACTTCTCCGGCGGTGAAGGACTTAACCGCGCGTGGT
TACTCTAGTGGAGCAAGAAATGAATTCGAATACGGCGCGTCTTTTAGGAGAGTGAGTGA
GTCATGCGGTGTTACGGTGGCTTGCTTGAGTCGGTCGAGTCTACGGTTCTAGTACGAA
TTCCGACCGTGCCAAAGTTGAGGAAGGAATTGCCGGAAGCTAGTAAACCGCGTGCGTG
CGAAGGAATCGATCGTATAGACGGTGCGAGGTGTTCCGGAAATGCGGAATGCGGATGAG
CATGGCTGGGTTGAGTTAATGCCATTGAGTGAGAAAGATAGCGGAGTCGATGAAGAGTCG

59/74

TGGAACCGAGTCCACCCGGGCTTTACCGTTAAAGAAAGATAACGGAGGTTGTGCTTTGG
TTGGATGGGACGGGCACTCACTGTCGCATCCGCTTGGCGTTAACTTCACACACTCTTTT
TTCTTCTTATTATTAACCATATTATTAAATTTTCGAGATTATTCTGATATTATCA
TTGTGATTTTCCGTTTCGAAAAGTGTAGGAATCTTATGTAAACAAGAAAAAAGACT
TTTATGTTTTTCTAATAATAAAGAAAGAGTGATTGGGTTCAAAAAAATAAAAAA
AAAAAAA

FIG.16F-2

60/74

SRPa8 (10964)

TGCATACAACGCACCGTTTTCGTAACACGGTTTCGCCGAGCTCTATTTCATTTCTCCTC
GATTTTGACATGCTTGAGACAAATTGTGCCACGAGAACGAGAGGATGTTCCCTTGA
GATGGAGGTCTTTGGGAGAGAGGCCACTGAATGTGATTGCTTGCGAAGGTTGGGAAAGAGT
GGAGAGGCCCTGAGACATACAAGCAGTGGCACGTACGGGCTATGAGGTCAGGGTTGGTGCA
GGTTCCATTTGACCCCAAGCATTAAGAAGACATCGCTGCATAAGGTCCACACATTCTACCA
CAAGGATTTGTGATCGATCAAGATAACCGGTGGCTCTTGCAAGGCTGGAAGGGAAGAAC
TGTCATGGCTCTTCTGTGTTGGAACCCAGAGTCCAGGCTTGACCGAGAAATCCTCGTTG
GCATATGAGAGACCATCTCTTGATTTTCTCTGTAATTCCCAGAGACAGAAATTACAG
ATGTAAGAAGAGAAATGCTGCACAAAGAACTTGTTCAAAGATAATATGATGTAAGTCCCTG
TTTATAACTTTCTAGCTGTGTTTTTGTGTTTCTCAGCTAGATTCTCCTAACGGTATTC
TTGTAGCTAGGGTATCAGATTGTTTGTAATATGCTAGCAGAGTTAGTTGCTCTAGATTG
TAACACATATAAGAGGGAAGCTTAGAGTTTCTATGGTTTAAAGAGAAAGTTTTCCTCTC
CAATGTAAAAAATAAAAAA

FIG. 16G

61/74

SRPa10 (11261)

AAAAATGGAAACCATCACTCTTGATGAACTTATGATCAATCCAGGAGAGACAACGGTC
GTCAACTGCATTCAATCGGTTACAATACACTCCTGATGAATACTGTGTCATTAGACTCTCCA
AGAGACACGGTTCTGAAAGCTATTTCAGAGATATCAATCCTGACCTCTTTGTGTTGCAGAG
ATTAAACGGAATGTACAACTCTCCTTTCTTCATGACGAGGTTCCGAGAGCGCTTTTTCAT
TACTCTTCACTCTTTGACATGTTTGACACCAATAACGAGAGGATGATACAAAAC
AGGTCACTGTTGGAGAGAGAGTTACTTGTGAGAGACGCGATGAGCGTATTTCTCTGCGAG
GGTGCAGAGCGGTTTGCAGGCCCTGAAACCTACAAGCAATGGCGAGTTAGGATTTTGAGA
GCCGGGTTTAAGCCAGCAACTATTAGCAAAACAGATCATGAAGGAGGCTAAGGAAATTGTG
AGGAAACGTTACCATAGAGATTTTGTGATCGATAGCGATAACAATTGGATGCTTCAAGGA
TGGAAAGGAAGAGTCATCTATGCTTTTCTTGCTGGAAACCTGCTGAGAGTTCACAAAC
AATAATTAAACATCTGAAAAATGTTACTTCTCAATTACATCATTTTGTGTTCCCAATGG
TTTGTAGAAATATGTTGATCCCGTGAGTGGAATGCAACTCTTTTTCCTGCAAGTACATA
TTGTATTCAAATCCTTGTGGAATGATAAATTGTTTAATCAAAAAA

FIG. 16H

62/74

SRPa11 (18652)

CGGAATGTTGAGATCTTGGAAAGCAATAGCTGGGAAACCAGAGTCCACATTAATCGATTTT
CAGATTGCACAGGATCACAATACATGTTTTCAGGATTCAGGAGCTTGCGAAACGCCCTGGT
GGCCCGCTGCTGCGTGACGGGTGGATGATTACAGTCCACCTATGCTCGTGGG
GGAGACTCAGCTTGGTAGGTAGAGGCTTGCAACTTTGGCGCAGTCATGTGGTGTCCCG
TTTGAGTTTCACGATGCCATCATGTCTGGGTGCAAGTGACGCGGAACATCTCGGGTTG
GAACCTGGCTTGTGTTGTGAACCTTCCCATATGTATTACACCATGCGCAGACGAG
AGCGTAAGTGTGAAATAACAGAGACAGGCTGCTGCATCTGATCAAGAGCCTCTCCCCA
AAACTGGTTACTCTAGTAGAGCAAGAAATCCAAACACACCTCGCCATTGGTGTACCGG
TTTGTGGAACACTGGATTACTACACAGCGATGTTTGAGTCGATAGATGCAGCACGGCCA
CGGATGATAAGCAGAGAAATCAGCGCAGAAACAACACTGTGTAGCAAGAGACATAGTGAC
ATGATAGCATGTGAGGAGTCAGAGAGAGTAGAGAGACACGAGTACTGGGAAATGGAGG
GTCAGAAATGATGCTGGGTTACGGGTTGGCCGTCAGCACATCTGCAGCGTTTGCA
GCGAGTGAGATGCTGAAAGCTTATGACAAAACTACAACTGGGAGGCCATGAAGGAGCG
CTCTACCTCTCTGGAAGAGACGACCCATGGCTACATGTTCGTTGGAAGCCAAACCCA
AACTATATTGGTAAGTTATAGTGATGATGGTTACTTGAGTGGATAAAGAGAGACAAC
AAAAACACATCTGCTGTAATTTTATAGGATGTGCAATGATGTTTAAAGTTGTAACA
CAACCTAAGTTATATATATACAAACCAACCTGGTGGTGTCTCTCTTGTAAATTG
TCATGTGTTGTGGTGGAGCTAGTAATGAAATATAACCAAAACATTGATTAGGTCAA
AAAAAAATAAAAA

FIG. 16I

63/74

FIG. 16J-1

SRPa12 (23196) *

TCTTACTCAAGGTTCTTTGTGTCATCTTGTGCCGAATCCACAAGAGGAGAAATAAAGA
TTCGACCTTTATTAGATATTAAACGACTCTGGATTTTGGGTTTGGAGTTGGATCCACA
TGGGTTCTTATCCGGATGGATTCCCTGGATCCATGGACGAGTTGGATTTCATAAAGGACT
TTGATTGGCCCTCCCTCAAAACCAACCTTAGGTTTAGCTAAATGGGTTCTATTAGATG
ACTTAGATTCTCATCCTTGGATCCTCCAGAGGCATATCCCTCCAGAACACAACAACA
ACAACATCAACAACAAGCTGTAGCAGGAGATCTGTTATCATCTTCATCTGATGACGCTG
ATTCTCTGATCTCTGTTTGAAGTATATAAGCCAAAGTTCTTATGGAAGAGGATATGGAAG
AGAAGCCTTGATGTTTCATGATGCTTTGGCTCTTCAAGCTGCTGAGAAATCTCTCTATG
AGGCTCTTGGTGAGAAAGACCCCTTCTTCGCTCTTCTGCTTCTGCTTCTGTTGATCATCCTGAGA
GATTGGCTAGTCATAGCCCTGACGGTTCTTGTTCAGGTGGTGCTTTTAGTGATTACGCTA
GCACCACTACCACTACTTCTCTGATTCTCACTGAGTGTGATGGTTTGGAGAAATAGAC
CTTCTTGGTTACATACACCTATGCCGAGTAATTTTGTTTTCCAGTCTACTTCTAGGTCCA
ACAGTGTACCCGGTGGTGGTGGTAAATAGTGGGTTTACGGTTACGGTTTTCAGGTTTGGCG
ATGATTTGGTTTCCGAATATGTTTAAAGATGATGAATGGCTATGCAGTTCAGAAAGGGG
TTGAGGAAGCTAGTAAGTTCCCTTCCTAAGTCTTCTCAGCTCTTTATTGATGTGGATAGTT
ACATCCCTATGAATTTCTGGTTCCAAGGAAATGGTTCTGAGGTTTGTGTTAAGACGGAGA
AGAAAGATGAGACAGAGCATCATCATCATAGCTATGCACCAACCACCAAGAGATTAA
CTGGTAAGAAAGCCATTGGCCGACGGAAGATGAAGATTTCGTTGAAGAAAGAGTAACA
AGCAATCAGCTGTTTATGTTGAGGAAGCGAGCTTCTGAAATGTTGATAACATGTTCC
TATGTGGCCCTGGGAAACCTGTATGCATTCTTAACCAGAACTTTCCTACAGAAATCCGCTA
AAGTCGTGACCGCACAGTCAAATGGAGCAAAGATTCTGTGGGAGAAATCAACTTCTACTA
GTATAGTAACGATTCTAAGAAAGAAACTGCTGATTTGAGGACTCTTTTGGTGTATGTG
CACAAAGCTGTATCAGTGGATGATCGTAGAACCGCCAACTTTAGCTAAGGCAGATACGAG
AGCATCTTCGCCCTCTAGGCAATGGTTCAGAGCGGTTGGCTCATTTATTTGCAAAATAGTC
TTGAAGCAGGCTTAGCTGGGACCGGTACACAGATCTACACCGCTTATCTTCGAAGAA
CGTCTGCAGCAGACATGTTGAAGGCTTACCAGACATACATGTGGTCTGCCCTTTCAAGA
AAGCTGTATCATATTGCTAACACACAGCATGATCGGTTTCACTGCAACGCCAACACGA
TCCACATAATAGATTTTCGGAATATCTTACGGTTTTCAGTGGCCTGCTGATTCATCGCC
TCTCGCTCAGCAGACCTGGTGTTCGCCTAAGCTTCGAATTACCGGTNNNNNNNNNNNN

64/74

NNNNNNNNNNNNNNNNNNAGTTCAGGAGACAGGTCAATCGCTTGGCTCGATACT
GTCAGCGACACAATGTTCCGTTTGAGTACAACGCAATTGCTCAGAAATGGGAAACGATC
CAAGTCGAAGACTTAAAGCTTCGACAAGGAGAGTATGTGGTTGTGAACTCTTTGTTCCGT
TTCAGGAACCTTCTAGATGAGACCGTTCCTGGTAAACAGCCCGAGAGATGCAGTTTGAAG
CTGATAAGAAAATAAACCCGAATGTCCTTCATTCACGCGATCTTAAGCGGGAATTACAAC
GCGCCATTCTTTGTCAAGAGGTTCAAGAGAGCGTTGTTTCATTACTCGGCTGTGTTGAT
ATGTGTGACTCGAAGCTAGCTAGGGAAGACGAGATGAGGCTGATGTATGTGTTGAGTTT
TATGGGAGAGAGATTGTGAATGTTGTGGCTTCTGAAGGAACAGAGAGAGTGGAGAGCCGA
GAGACATATAAGCAGTGGCAGGCGAGACTGATCCGAGCCGATTTAGACAGCTTCCGCTT
GAGAAGGAACTGATGCAGAACTCTGAAGTTGAAAATCGAAAACGGGTACGATAAAAACCTC
GATGTTGATCAAAAACGGTAACTGGTTACTTCAAGGGTGGAAGGTAGAAATCGTGATGCT
TCATCTCTATGGGTTCCCTTCGTCCTTCAATAGATGTTGTTTCTTACGTTCTAAGCGACTGGG
ATTATGTAGGGCTTTCTGTGTGATAGTCTCTCGCCCAACACGAGTGGATTAAAGTTCAGAG
TTAGGGTCTTGAAACACTAGAAATGTTGTTTATATTATGCTTGTGACATAGCGTGTGAAGA
GTGTAGCCCTAAGAGATATAGTACTCATTCATGTCATGATCTTTTGCTATATATGTTNCATGT

FIG. 16J-2

65/74

SRPd1

TCTGCAGACAATTTNAGGAGGCCAATACCATGCTATTGGAAATTTCAGAACTG
TCCACACCTNNNNNNNNNNNNNNNNNNNNNNNNNNNNNGTACTTCTCAGAGGN
AATGTCGGNNAGATTAGTTAGCTCCTGCTTAGGAATCTATGCTTCTCTCCNGC
AACAGTGGTGCCCTCCTCATGGTCAGAAAGTGGCCTCA

FIG. 16K

SRPq1

TCAACTGAGAACTAGAAAGATGCCAACAAGATGCTTCTGGAGATTCTCAGTTA
TCAACACCGTTTCNNCACTTCAGCACAGCGTGTGGAGCATATTCTCAGAAGCC
ATATCAGCAAGGTTGGTGAGTTCATGTCTAGGGATATACGCAACTTTGCCACAC
ACACCCAAAGCCACAAGGTAGCTTCAGCTTTTCAAGTGTTCATGGTATTAGT
CCTTTAGTGGAGTTCTCACACTTCACAGCAACCAAGCAATTCAAGAAGCCTTC
GAAAGAGAAGAGAGGGTGACATCATAGATCTTGATATAATGCAAGGGTTG

FIG. 16L

66/74

SRPp1

TCTGCAGACAACCTTGAAGAAGCCCAATACAATACTGCCCTCAGATCAGAACTC
TCCACCCCTATNGCAACTCGGTGCAACGAGTGGCTGCCCTATNNNNNNNNNN
NNNNNNNNNNNNNNNNNNNTGCATAGGAATGTATTCTCCTCTCCCTCCT
ATTCACATGTCCCCAGAGCCAGAAATTGTGAAT

FIG. 16M

67/74

FIG. 17A-1

Partial DNA sequence of ZCARECROW gene

GATATCAGCATCAATTTTAAATGTAAGTTGGCAAAAGATCATGAGGGTTCTCATAGT
AATTGGCCACAAGGTATGACACTGTCTCAATTGAGCAATCTAGTAGAGAAACTGATCCA
TCATATATTGCTCATATTGAAGTGAAAAGATATGCTCAAGAACCTAGTAGAGAAAGCTA
AAAATTGAAAAAATCTAGCTCTACTAGAAAATAATGATAGGTTGCCCTGTTCTCATGAAA
TTTATTAGATAATCATATCATGGCTAGATGTCGCTCATGAGGTTGTTCTTGCTAGTTTAG
ATTCCTGTGGCATTTCATCTCTTTTAGATGCACTAACATGATGGAAGTTTCTAATCTGG
TGCTTCACAATTTCTGGTGATTTCATGCTTCCCTTCATTGCAATTGATATGATGCTTGATTC
ATGCTTCAGTCACTTTGTGCGTTTAAATTGGTATTGATGATGATGATGATGATGATGATG
CTGCAACTAGTGTTCACCATGTGGTTTTTTTAGTATCATTCGTATAGTTTCTAACTTTC
TATTGATAATTAAGTGATAAAGTAGTTTTAGAAAATATCTCTTGCGCCATTAAATGCTAC
AACTTGTTTTTAGCGGTACGTTAGCATTAATAATTTCTTATTAAGAAAGCGGAAGAG
AAACGGCCCCAACAGAGCATCCACGTCGTCATTTACCTTCATCGTTGGATCATAGA
TGAGCGGTCCACGGTGAACTCCGTTTGCTGCAAAACCACTCTACGCGCTGTTAAG
TAGCTTCTAGAAACATCACGATGTGTCCTCCCTCCATTCTTTAGGAGGAGCCGGATCCGGC
GCCGAGTCGCCCAAGTCCCGACCGCGGCTCGGCGCGCTCCACCTGCTGAGTGTGACGC
GGAGGTGCAGCGGGAAGCAGCGGACGAGAGGGCTCCACCTGCTGAGTGTGACGC
TGCTGCTGCAGTGCAGCGGAGGCCGTGAACGCGGACACCTCGACGACGCGCACAGACGC
TGCTGGAGATCGCGGAGCTGGCCACGCGTTGCGACCTCGACCCAGCGCGTGCGCGCT
ACTTCGCGGAGGCCATGTGCGCGCGCTGTCAGCTCCTGCTAGGCTGTACGCGCGC
TGCCGCGGGCTCCCGCGCGCGCGCTCCACGGCGCGTGCGCGCGCTTCCAGG
TGTTCAACGGCATCAGCCCCCTTCGTCAAGTTCTCGCACTTCACCGCAACCAAGGCCATCC
AGGAGCGTTCAGCGGAGGAGCGGTGCACATCATCGACCTCGACATCATGAGGGGC
TGCAGTGGCGGCGCTTCCACATCCTTGTCTCCGCGCGCGCGCGCGCGCGCGCGTCA
GGCTCACCGGCGCTGGGGCGTCCATGGACGCGCTCGAGGCGACGGGGAAGCGCTCTCCG
ACTTCGCGACACGCTCGGCTGCGCTTCGAGTTCTGCGCGCTCGCGGAGAGCGCGCGCA
ACGTTGACCCCGCAGAAAGCTGGGCGTCAAGCGGCGGAGCGCTCGCGGAGAGCGCGCGC
ACCACTCGCTTACGACGTCACTGGCTCCGACTCCAAACACGCTCTGGCTCATCCAAAGGT
CCTCCATTTTCTCTGCGCTTCTTCCATGTCAAACTTGATGCAATCATGACCACTT
TTCAGCTGCTGACATTGGATAATGTGAGCTTTACGGCAAGCATCAAGTCGTGAGTACA

68/74

TCCATTACAGCTATTCTCTAAATATTCTCGGAGGTTCTCTGCTCATAGTAAATAAAAT
 CGCGTTTGAAGCTCAAAAGGCGATTCTTCCGAGGTTTCTGCTGTTGAGCGCTATTTTGA
 AACCCCATTTCTCAATGATTTTATTTTAAAGAAAATTAGTTCATTTTCTCTCTG
 TGAATGGAGTCCCAAACTAACCCCTAATATAAAAAAAGCGGCTTGGAGCTCAAAACG
 CTCGTTGTTATGACCAACCGCTTTATAGGTTTAAAAAGTTGAATCTTGACAAATGCTTT
 TGAAAAGGTTGAATCTTGACCAATGCTTTTGAGATGATACTGTAGTGTAGTCTGTAGTGA
 GCATCCTCCATGGTCTTTGGTGATCGAGAAATCCTGCAGCCCCGGGGATCC

FIG. 17A-2

Partial amino acid sequence of ZCARECROW protein

YQH HQFXM XVGKRSXGFSXXFGHKVXHCLNXAIXRXNXSIIYCSYKXKRYAQEPSREAK
 NXKIXLYXKNMIGCLFLMKIYXIIISWLDVAHEVVLASLDSCHSSLLDALTXXEVSNLV
 LHN SGDSCLH CNXXCLIHASVTL CVXLVYVSLDCRV SATSVSPCGFLVSFVLVSNFL
 LIYXSDNXFXKYSLVPLMLQLVFSVVSIIIFPYYESGRETRPTRASTSSHFTFIVGSXM
 SGPRXTPFACKTTSSTRCXVASRNITMCPVHSFRRSRIRRRSRPSRPPRPPRPPRSGR
 RCGSGSATRRASTCXVLTLLLOCAEAVNADNLDDAHQTLLLEIAELATPFGTSTORVAAY
FAEAM SARVVS SCLGLYAPLP PGSPAAARLHGRVAAAFQVFN GISPVFV KFSHFTANQAIQ
EAFEREERVHII DLDMOGLWPGLFHILVSRPGPPRVRLTGLGASMDALEATGKRLSD
FADTLGLPFEFCVAE KAGNVDPQKLG VTRREAVAVHWP HHSLYDVIGSDSNTLWL IORS
 SIFLLCLSSMSNLDAIMTTFQLTLDNVSF TASIKSWXYIHYSYFXN IILRRFPAH SKKS
 RFEAQKAISS EVCXALFWKPHFLNXFLFFKEKLVHFSLVKWSPKLT LILKKTRFGA QNA
 RCYDQPALXVXKGXILTM LLLKRLNLDNAFEMI LXC SLXWSIILHGLWXSRI PAARGI

FIG. 17B

69/74

302	349
SCR	SADNLEEANKLLLEISQLSTPYGTSAQRVAAYFSEAM SARLLNSCLGI
SRPd1	SADNFxEANTMLLEISELSTPXXXXXXVFSxxMSxRLVSSXLXI
SRPg1	STENLEDANKMLLEISQLSTGXXXXXXXXXXXXXXXXXXXXXSCGLI
SRPPi1	SADNFEEANTILPQITELSTPYXNSVGRVAAYXXXXXXXXXXXXXXCIGM

350	Y A A L P S R W M P Q T H - S L K M V S A F Q V F N G I S P L V K F S H F T A N Q A I Q E A F E	396
SCR	Y A S L P A T V V P - - P H G Q K V A S	
SRP d1	Y A T L P - - - - H T H Q S H K V A S A F Q V F N G I S P L V E F S H F T A N Q A I Q E A F E	
SRP g1	Y S P L P P I x M S Q - - - - S Q K I V N	
SRP p1		

397 412
SCR KEDSVHIIDLIMQGL
SRPgl REERVHIIDLIMQGL

FIG. 18

70/74



FIG. 19A



FIG. 19C



FIG. 19B



FIG. 19D

71/74



FIG. 19E



FIG. 19F



FIG. 19G

72/74

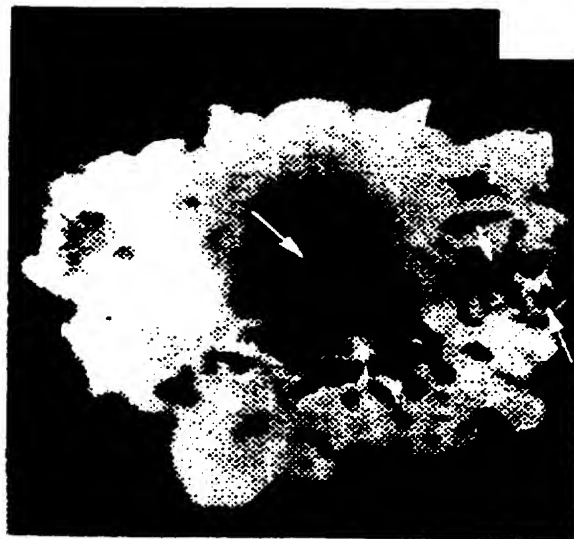


FIG.20A

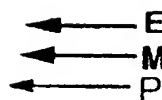


FIG.20B

SCR Promoter::GUS^{73/74}



FIG.21A

SCR Promoter::SCR



FIG.21B



FIG.21C

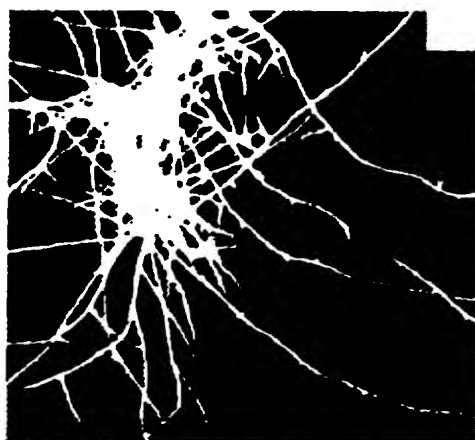


FIG.21D



FIG.21E



FIG.21F

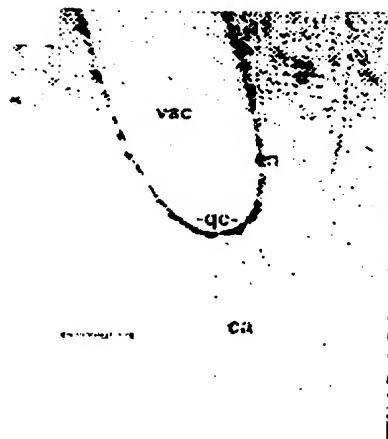


FIG. 22A

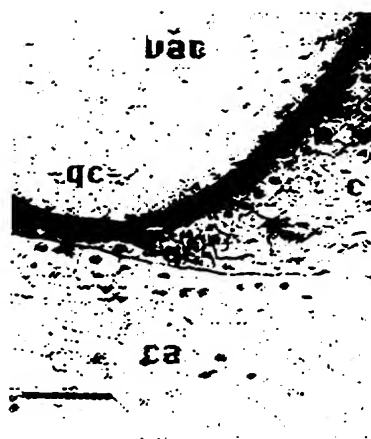


FIG. 22B

INTERNATIONAL SEARCH REPORT

 International application No.
PCT/US97/07022

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : C07K 14/415; C12N 1/21, 5/10, 15/29, 15/63; A01H 5/00

US CL : Please See Extra Sheet.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 536/23.6, 23.1; 435/320.1, 252.3, 419; 530/350, 370, 387.9; 800/205

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, DIALOG - Biotech Files, GenEMBL sequence databases

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	SCHERES et al. Mutations affecting the radial organisation of the <i>Arabidopsis</i> root display specific defects throughout the embryonic axis. Development. 1995, Vol.121, pages 53-62, see entire document.	1-28
Y, P	WYSOCKA-DILLER et al. Root radial organization. Plant Physiology. June 1996, Vol. 111, No. 2, abstract no. 40001, page 12, see entire abstract.	1-28

☐ Further documents are listed in the continuation of Box C.
 ☐ See patent family annex.

* Special categories of cited documents:	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be of particular relevance	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
B earlier document published on or after the international filing date	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*Z* document member of the same patent family
O document referring to an oral disclosure, use, exhibition or other means	
P document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

15 AUGUST 1997

Date of mailing of the international search report

03 SEP 1997

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INTERNATIONAL SEARCH REPORT

International application No.
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A. CLASSIFICATION OF SUBJECT MATTER:

US CL :

536/23.6, 23.1; 435/320.1, 252.3, 419; 530/350, 370, 387.9; 800/205